

# Reconstruction d'embryons à partir d'images 4D

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Module Image - LIRMM - ICAR  
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# Plan présentation

1 Introduction Biologie du développement

2 Protocoles d'acquisition et images 4D

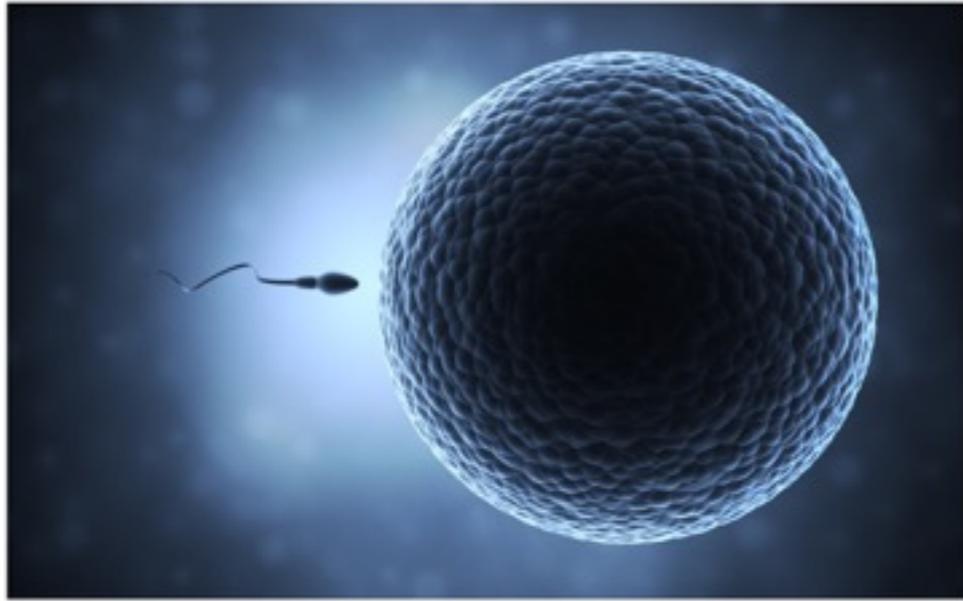
3 Reconstruction phénoménologique de la morphogenèse à partir d'images 4D

( *Filtres, Détection, Segmentation, Suivi cellulaire* )

4 Visualisation de données 4D

5 Validation de données

# Embryogenèse

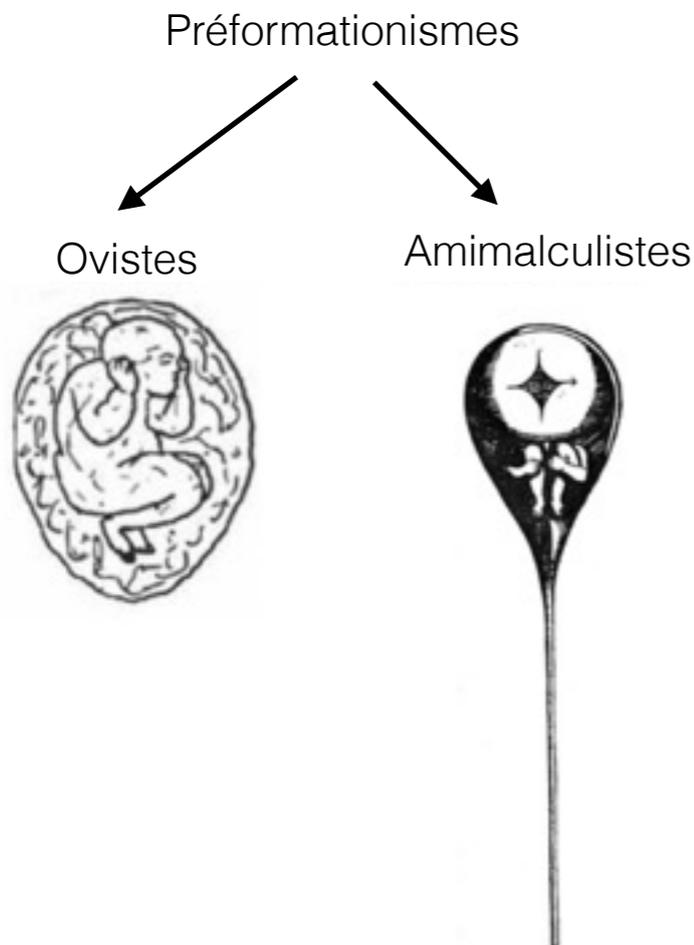


# Un peu d'histoire ...

*IV siècle a.v. J.C.*

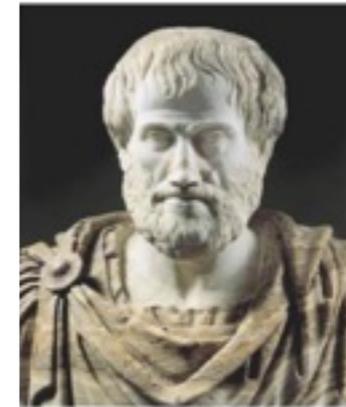
## Théorie de la Préformation

Dans l'œuf, l'adulte préexiste en version miniature



## Théorie Epigénétique

De nouvelles structures s'ajoutent progressivement au cours du développement



Aristote

**Hypocrate** pensait que nous héritons de petits morceaux de toutes les parties de nos parents.

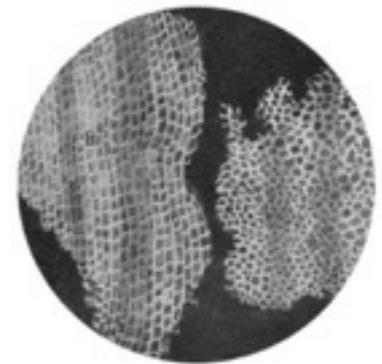
**Aristote** pensait que nous grandissions à partir de formes indistinctes qui se développent à l'intérieur de maman à cause de papa.

## Théorie cellulaire

- Tout organisme vivant est composé d'une ou plusieurs cellules
- La cellule est élémentaire de la vie
- Tout cellule provient d'une autre cellule

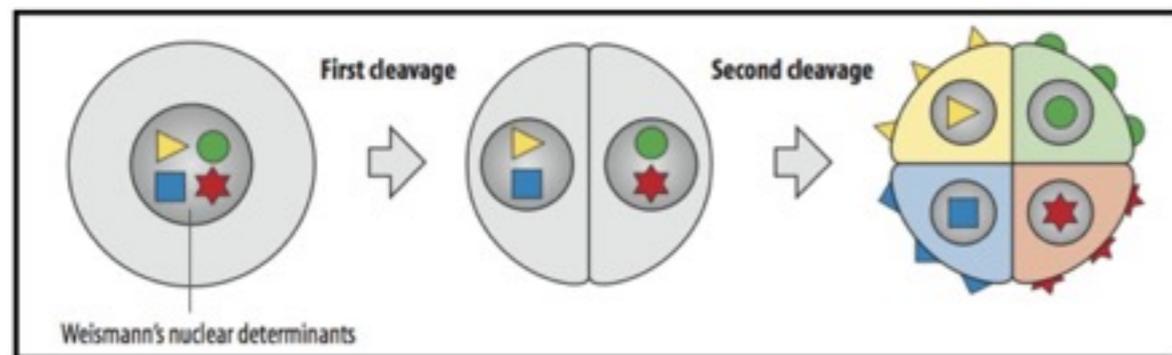


Le microscope de R. Hooke



Cellules de liège

## Théorie du développement mosaïque

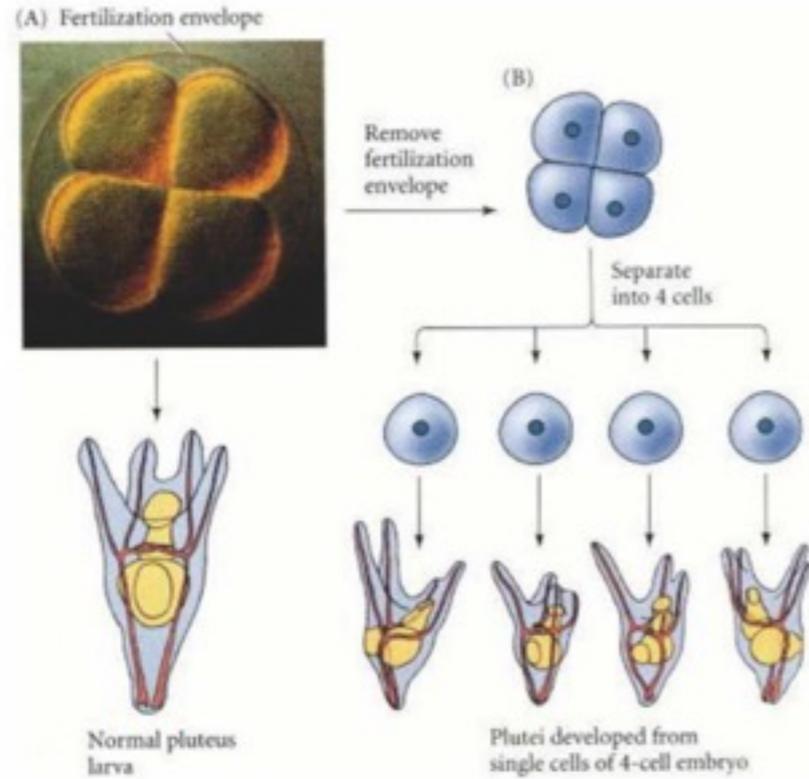


August Weismann

Le noyau comporte des déterminants qui sont partagés de manière asymétrique entre les filles

# Embryologie experimentale

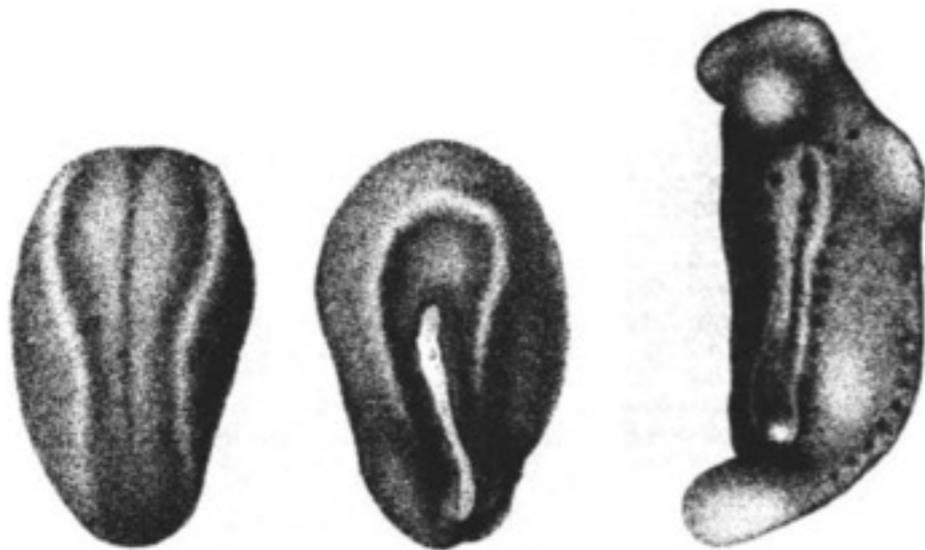
19ème siècle



Hans Driesch

Chaque cellule d'un embryon d'oursin précoce a la capacité de donner un oursin ....

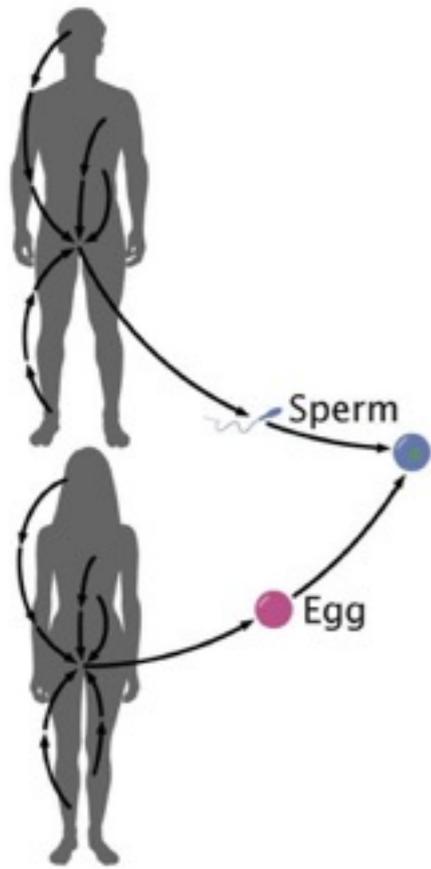
# Embryogenèse



Hans Spemann

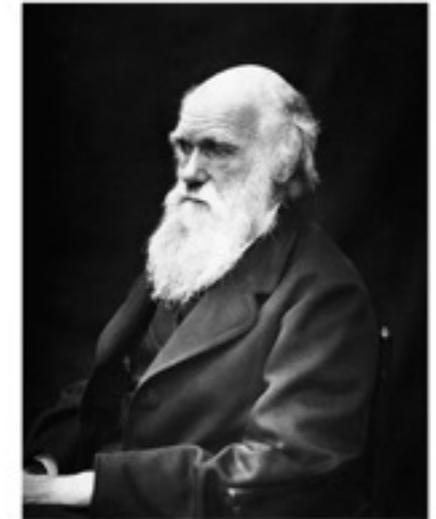
Différentiation cellulaire par signaux chimiques

# Pangenèse



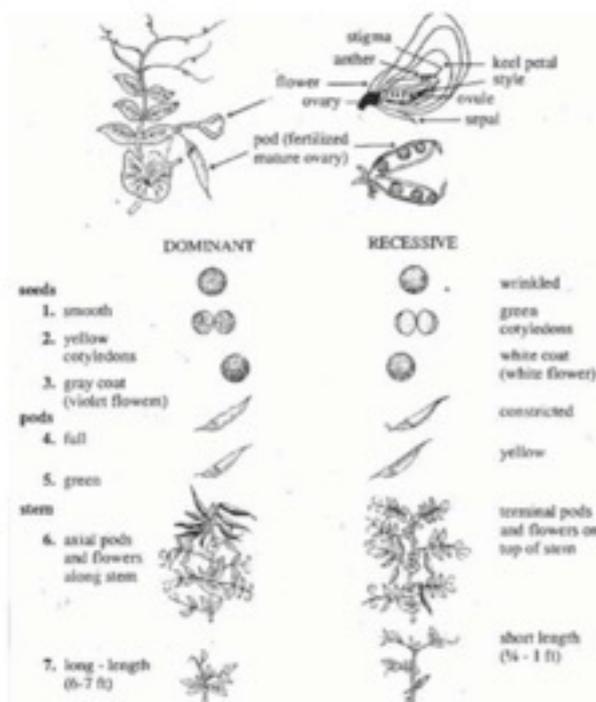
L'ensemble de l'organisme participe à l'hérédité

19ème siècle



Charles Darwin

# Lois de Mendel

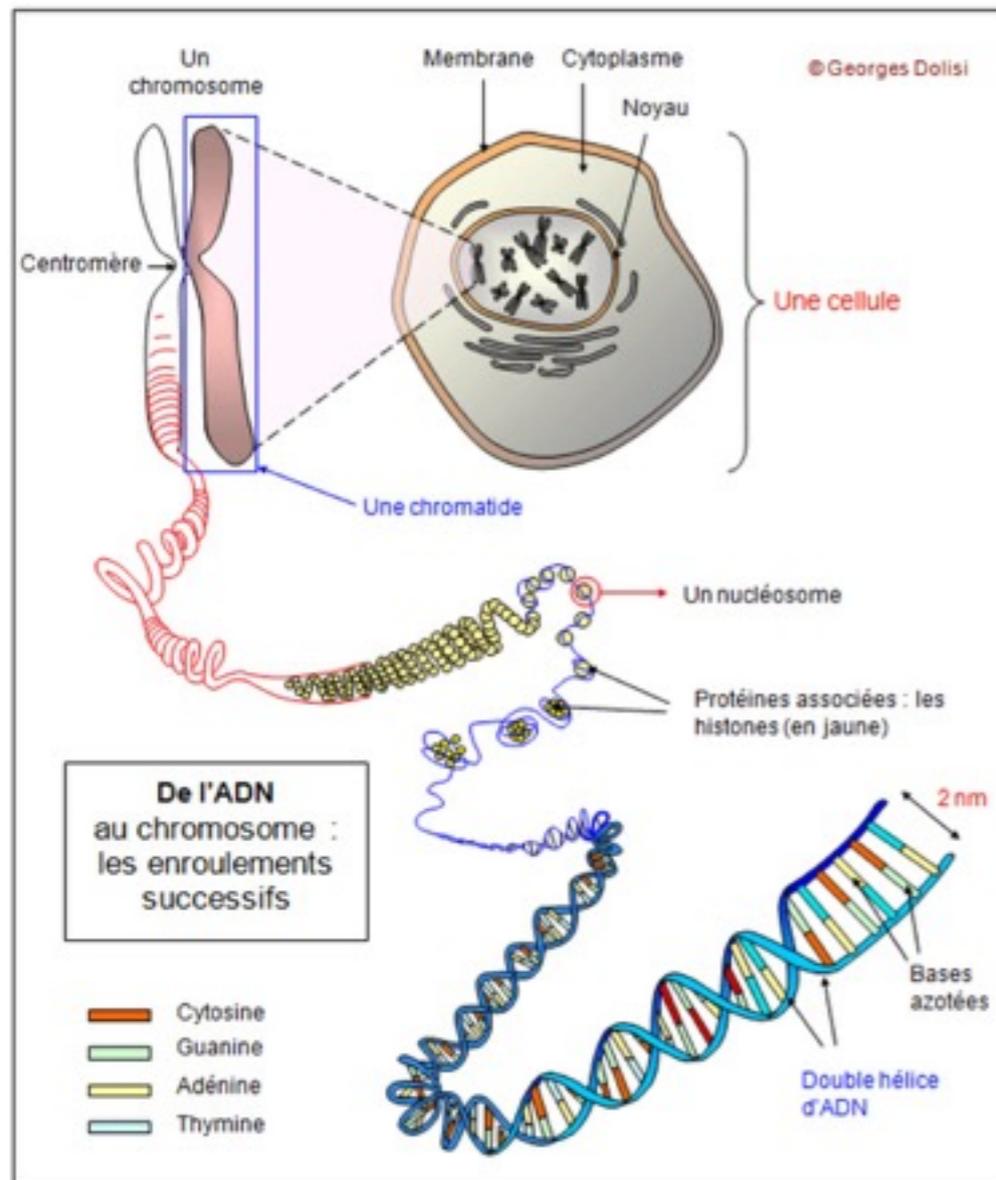


Facteurs héréditaires

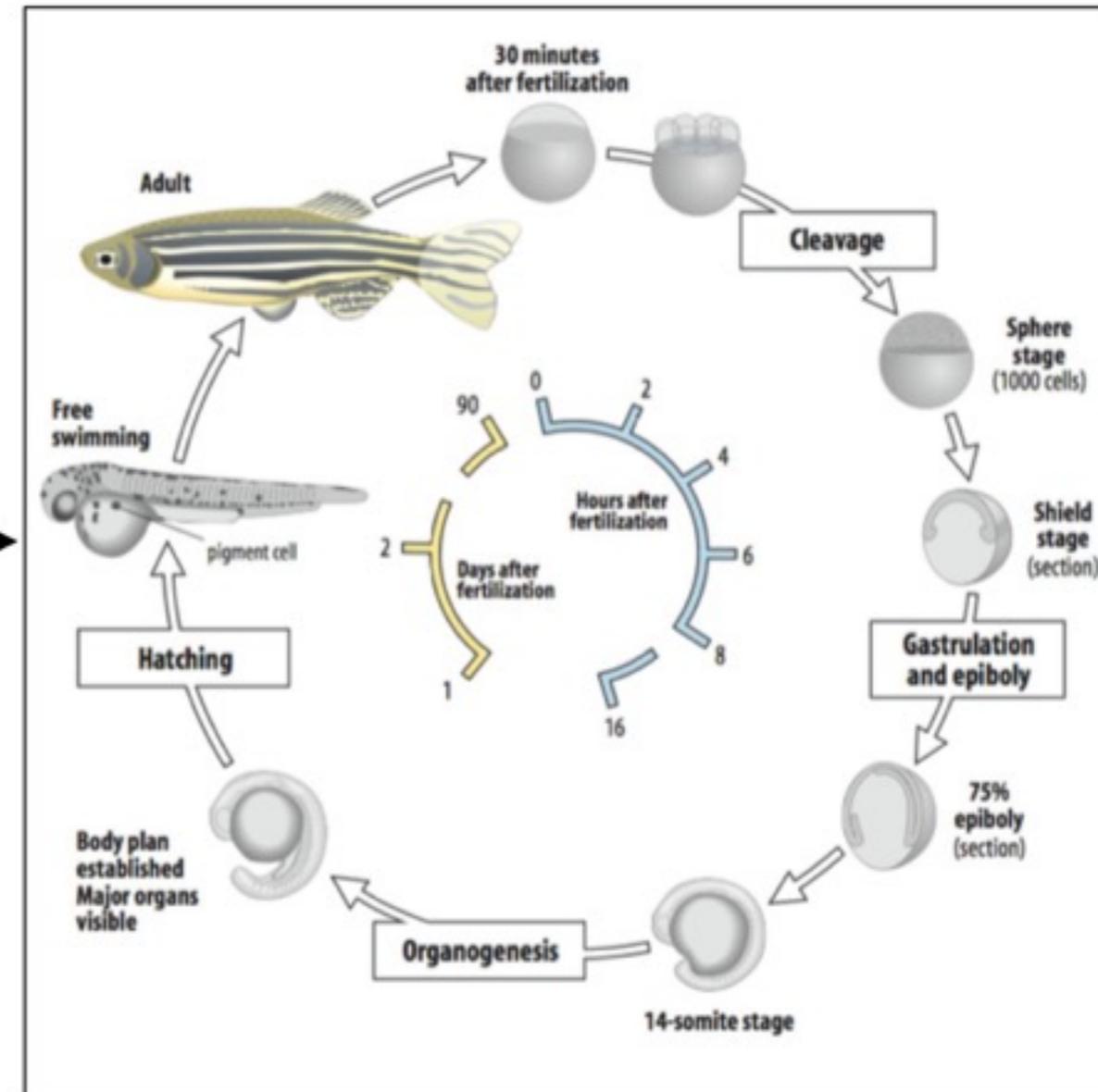


Gregor Mendel

# Comment l'information génétique contrôle le développement ?

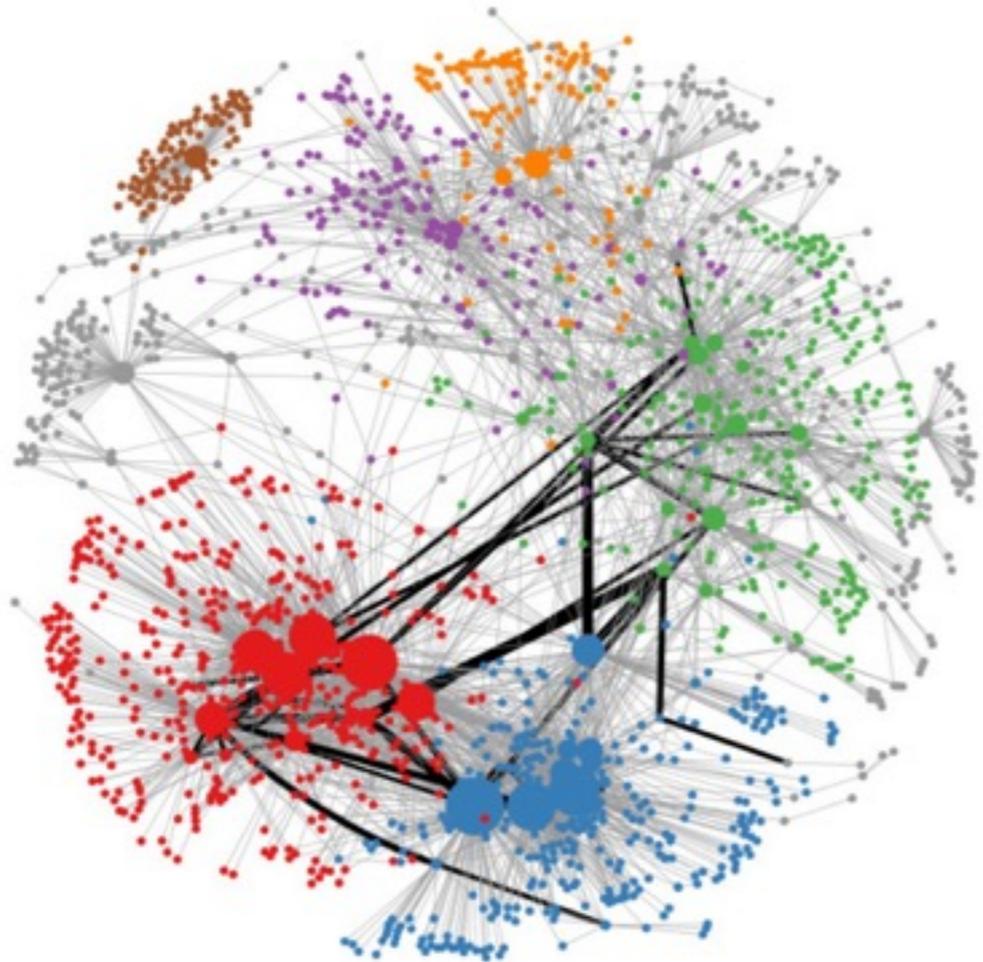


?



# Comment l'information génétique contrôle le développement ?

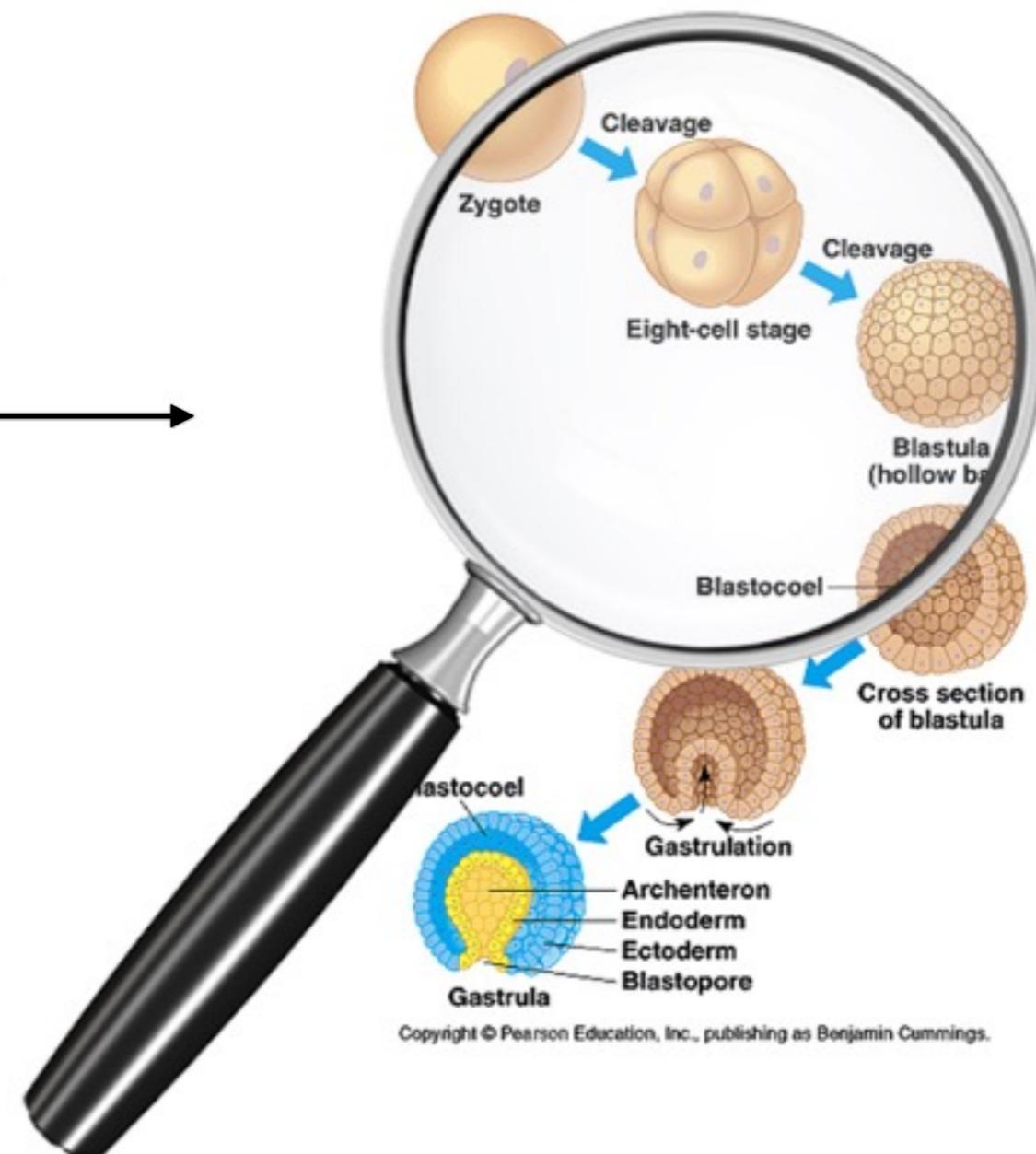
Réseau de régulation génétique



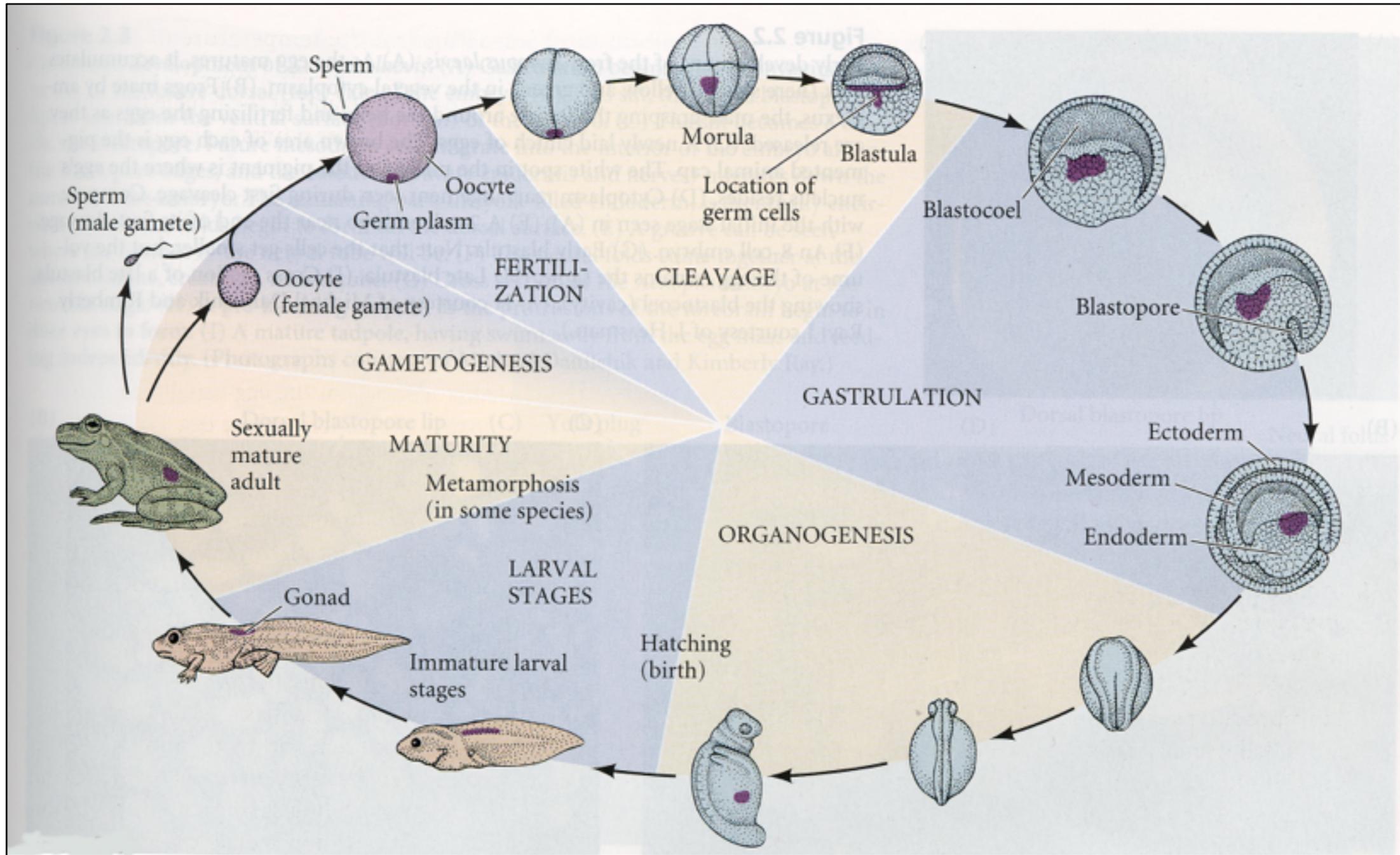
?



Stade de développement



# Que veut on observer ?

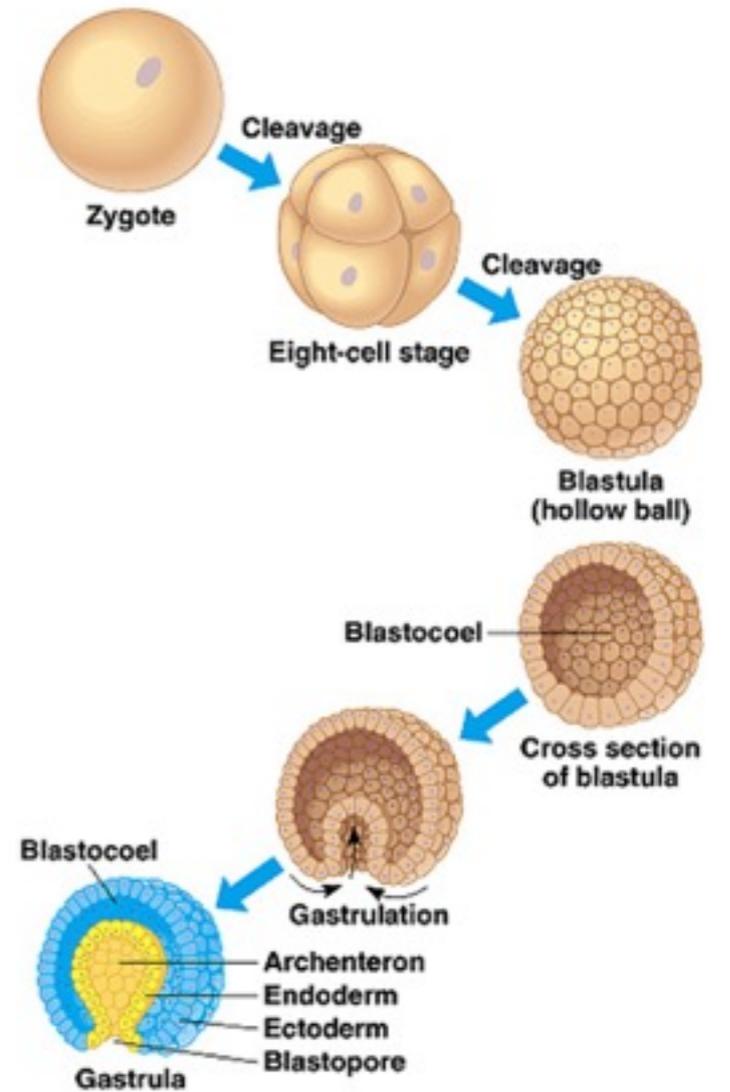
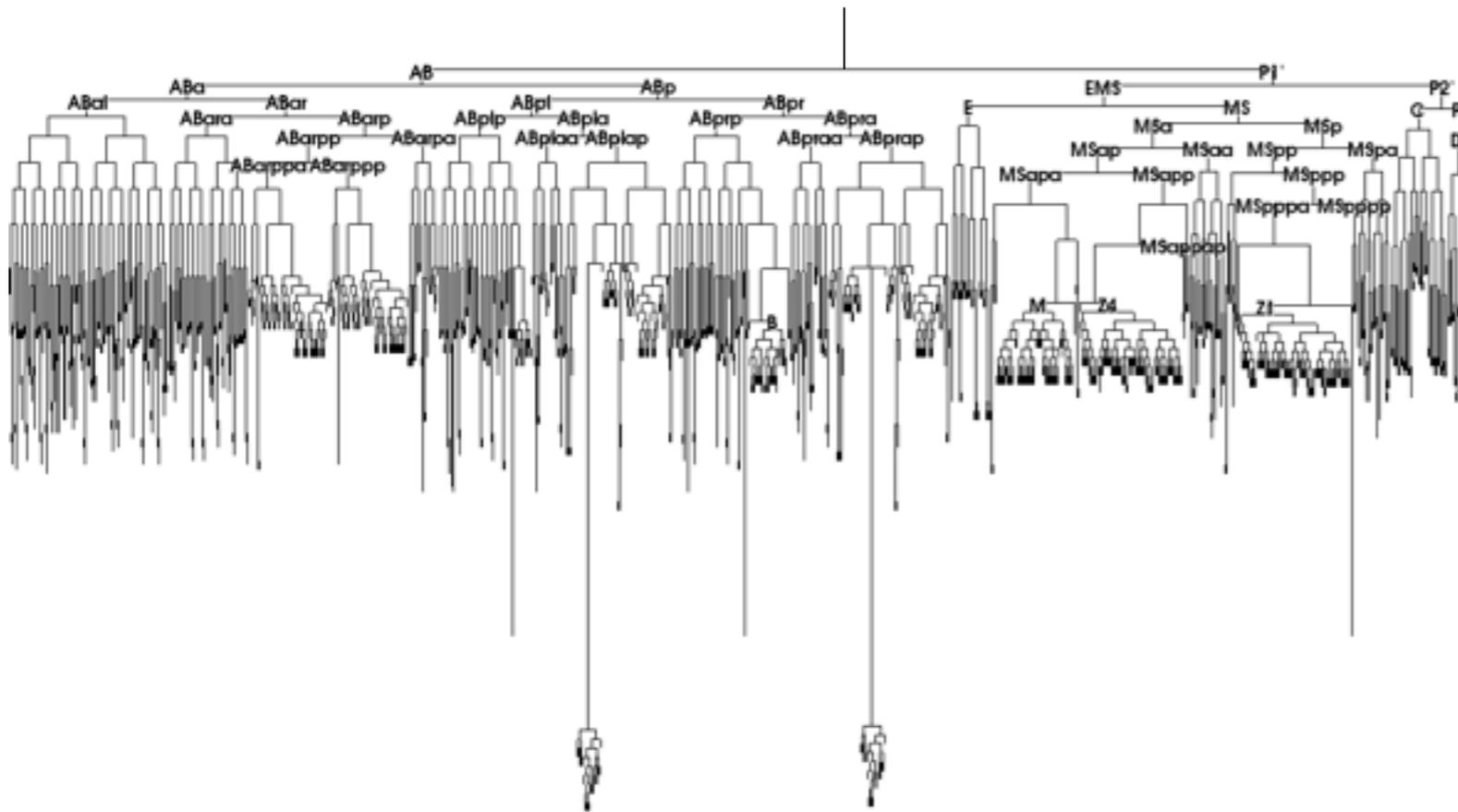


L'embryon à l'échelle cellulaire !

# Suivi cellulaire

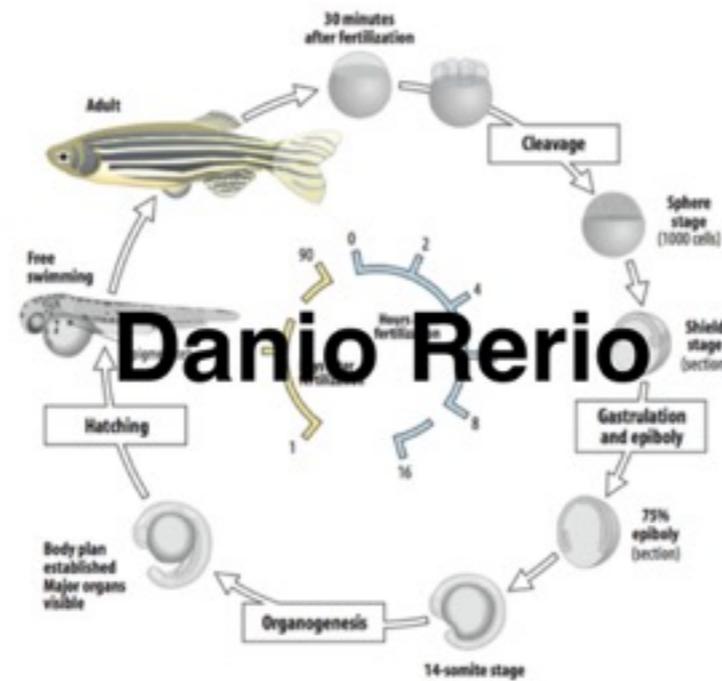
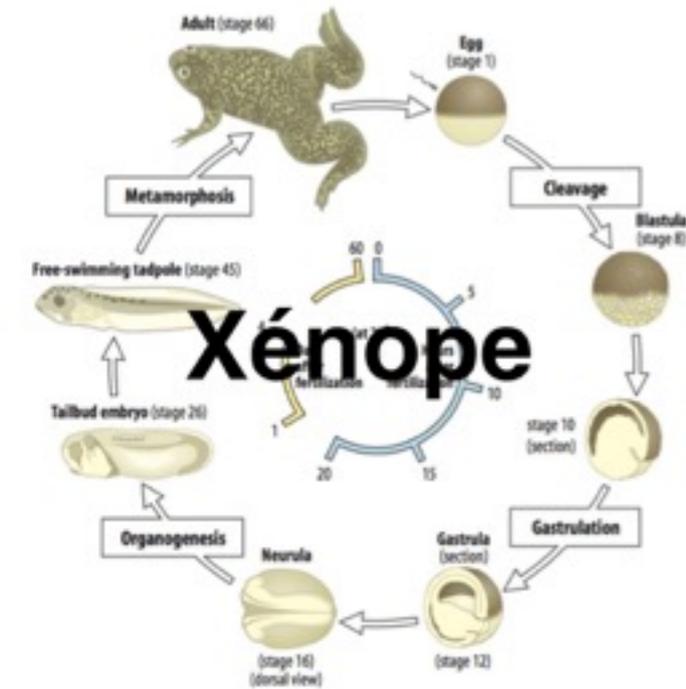
## Arbre de lignage cellulaire

[ *Caenorhabditis elegans* , Sulston 1983 ]

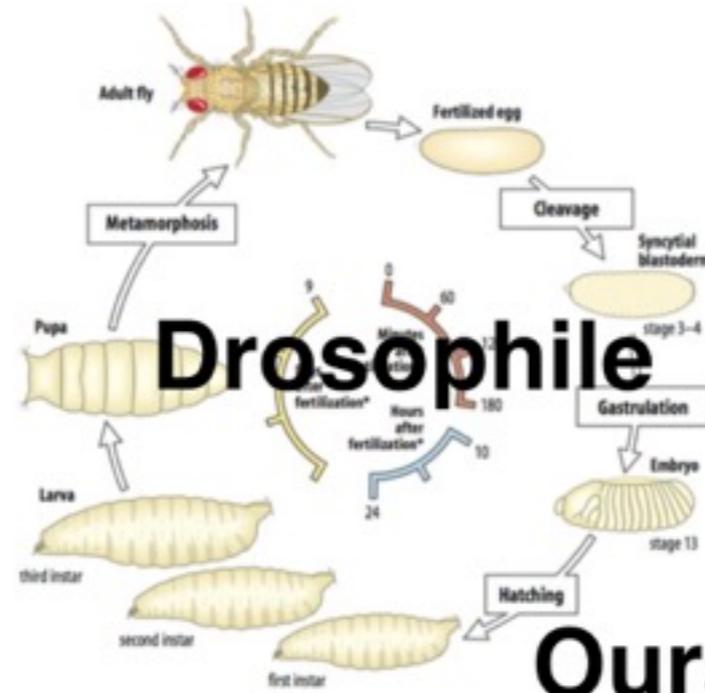
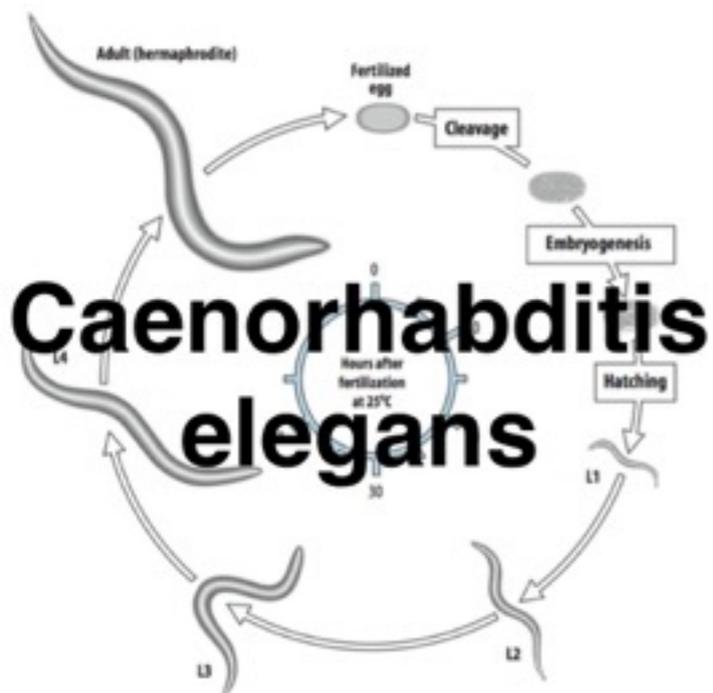


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# Modèle de la biologie du développement



- taille
- position phylogénétique
- transparence
- accessibilité
- cycle de vie
- ....



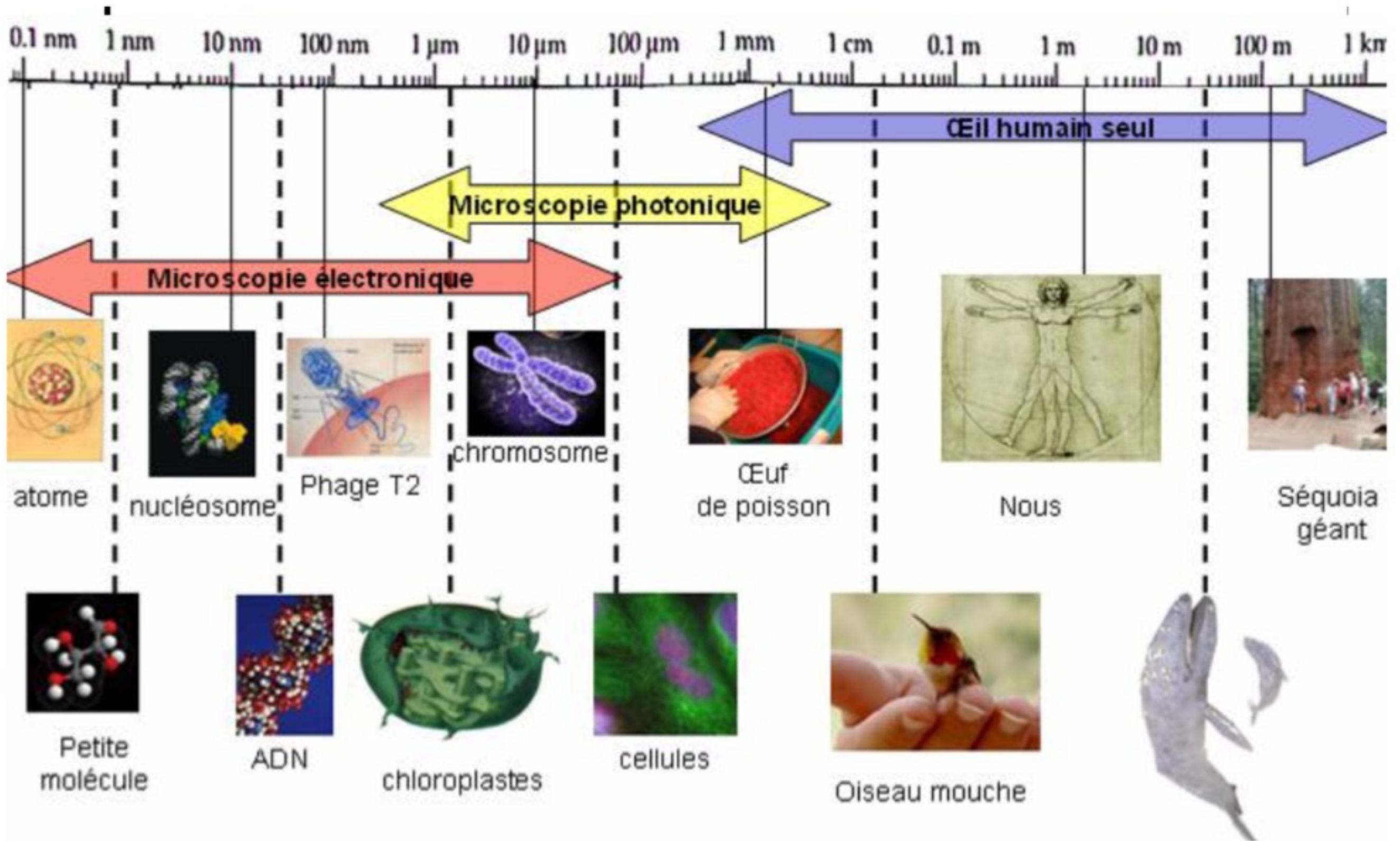
**Oursin, Ascidies, Souris ! ...**

Comment peut-on l'observer ?



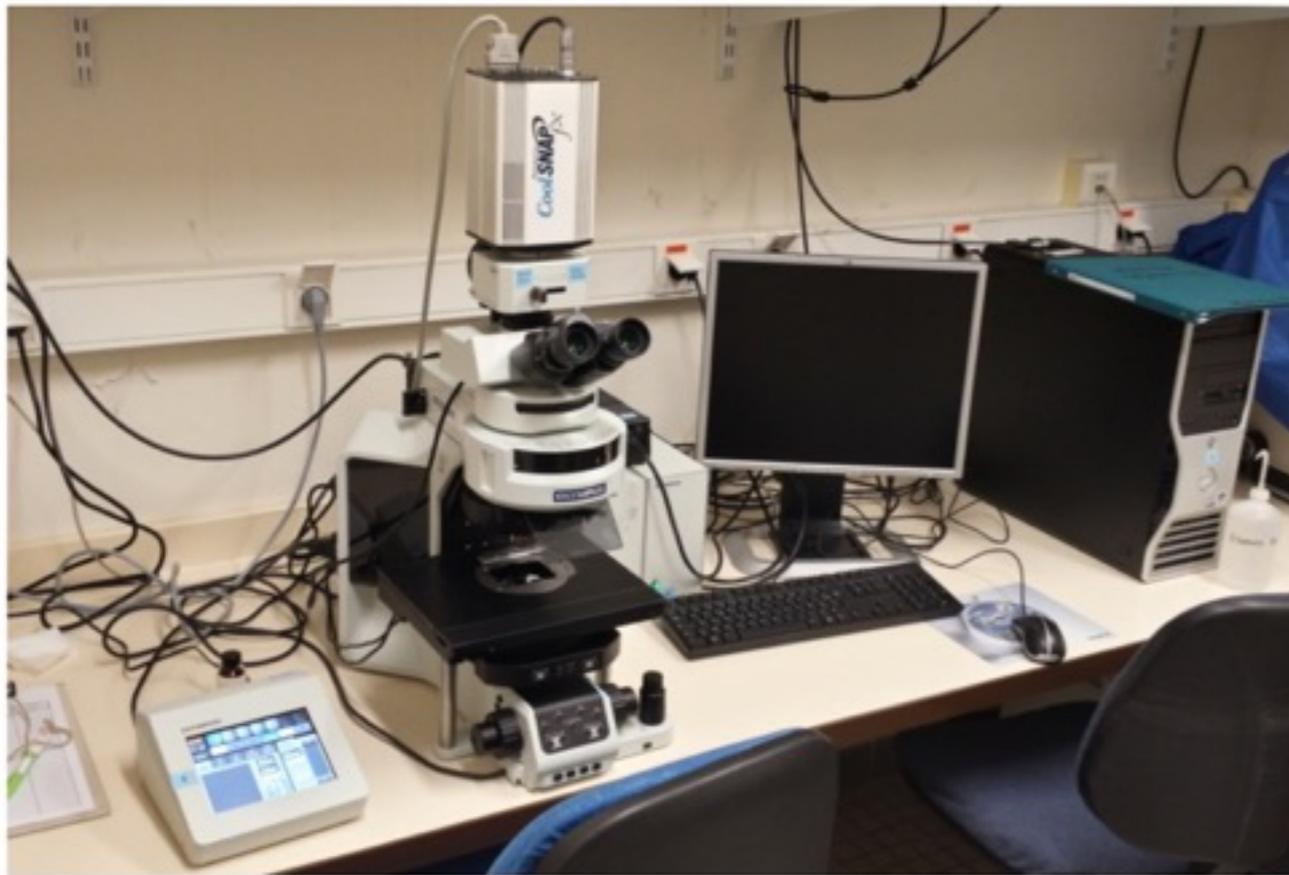
au microscope !

# Quel microscope ?



# Microscope à contraste interférentiel (*Normarski 1950*)

exploite les interférences de deux faisceaux d'une onde lumineuse traversant un échantillon

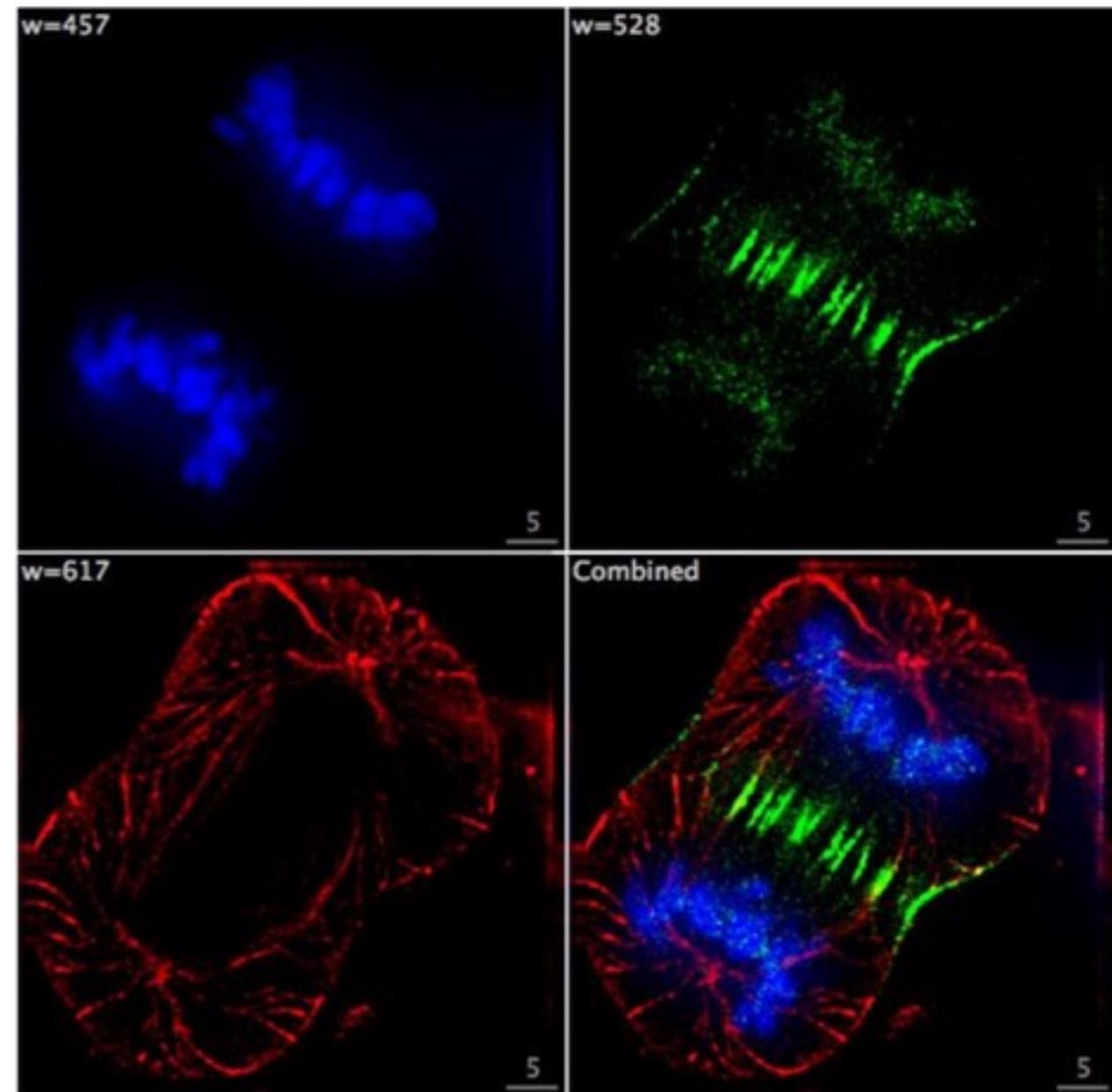
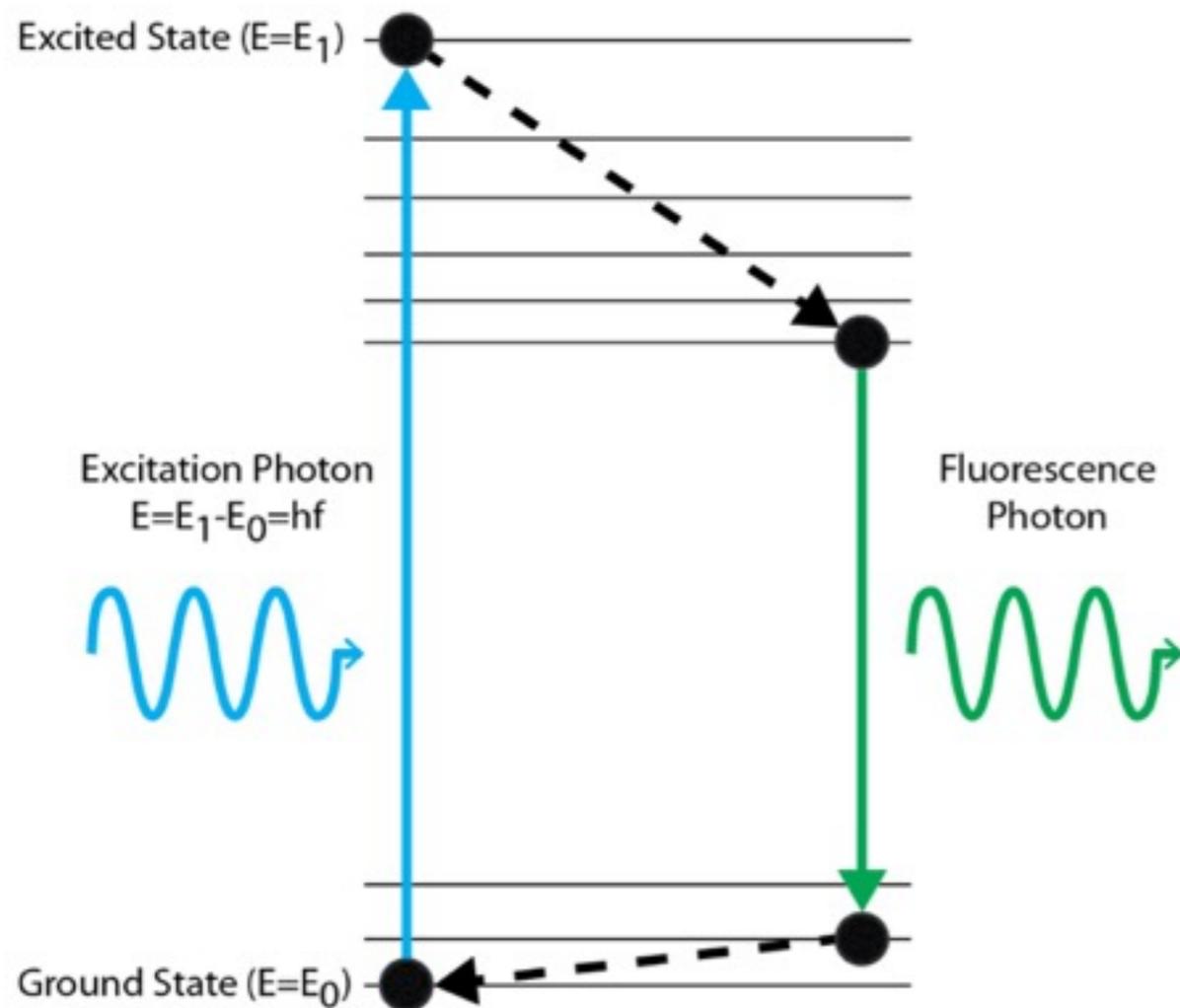


Kimmel 1995

—————> Images 2D + temps

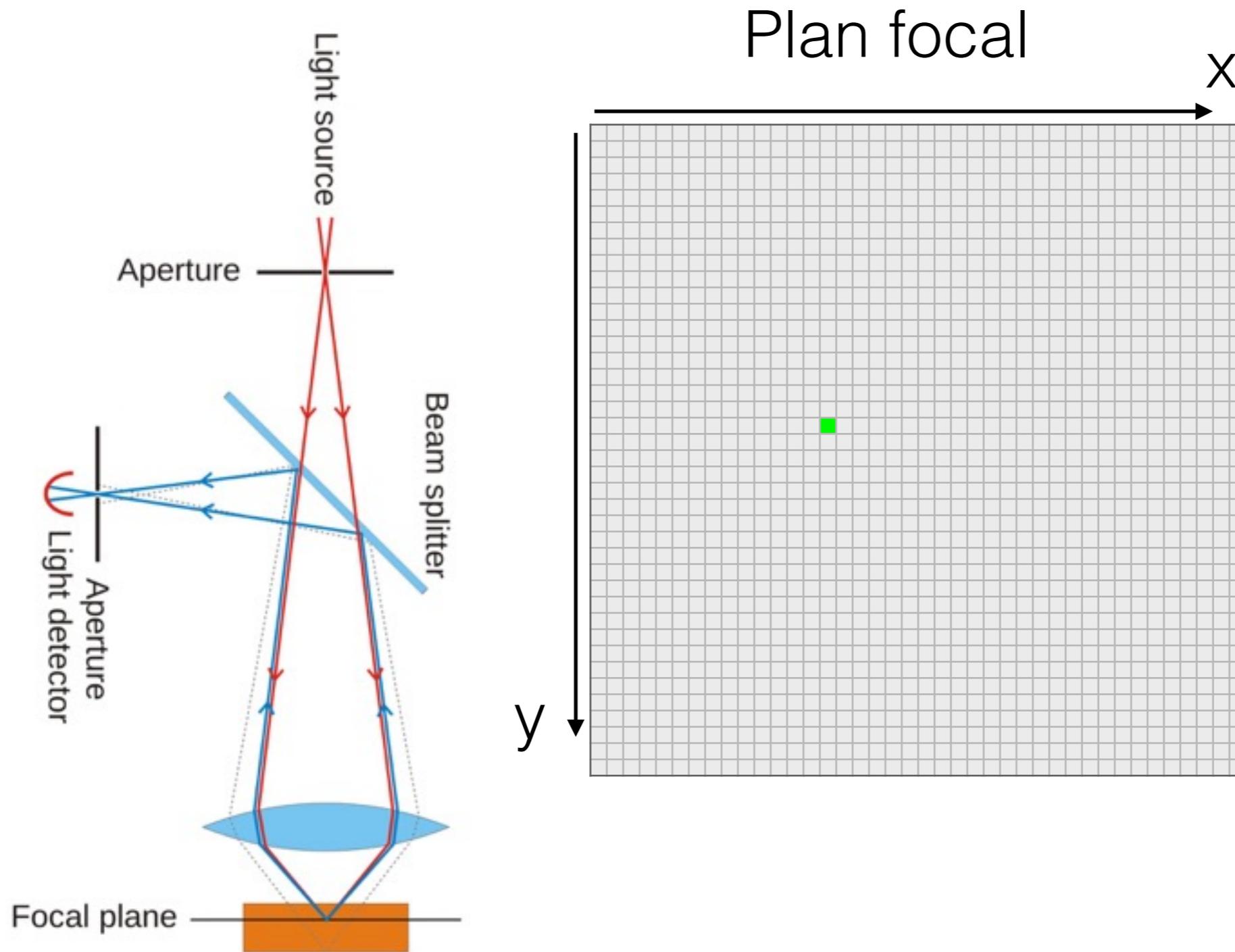
# Fluorescence (Stokes 1852)

La fluorescence est la propriété que certains corps ou **molécules** ont à émettre une **lumière** après avoir été excités avec une lumière d'énergie supérieure. Ainsi, un objet excité par une longueur d'onde émettra une **fluorescence** à une longueur d'onde supérieure.



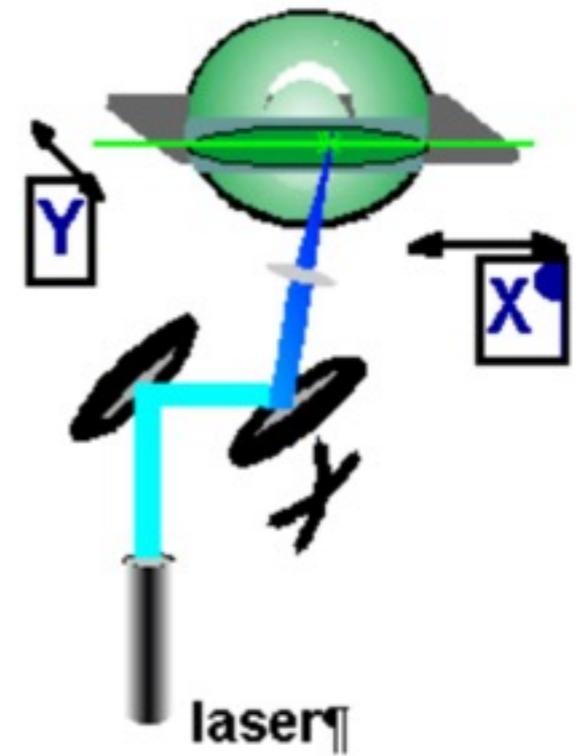
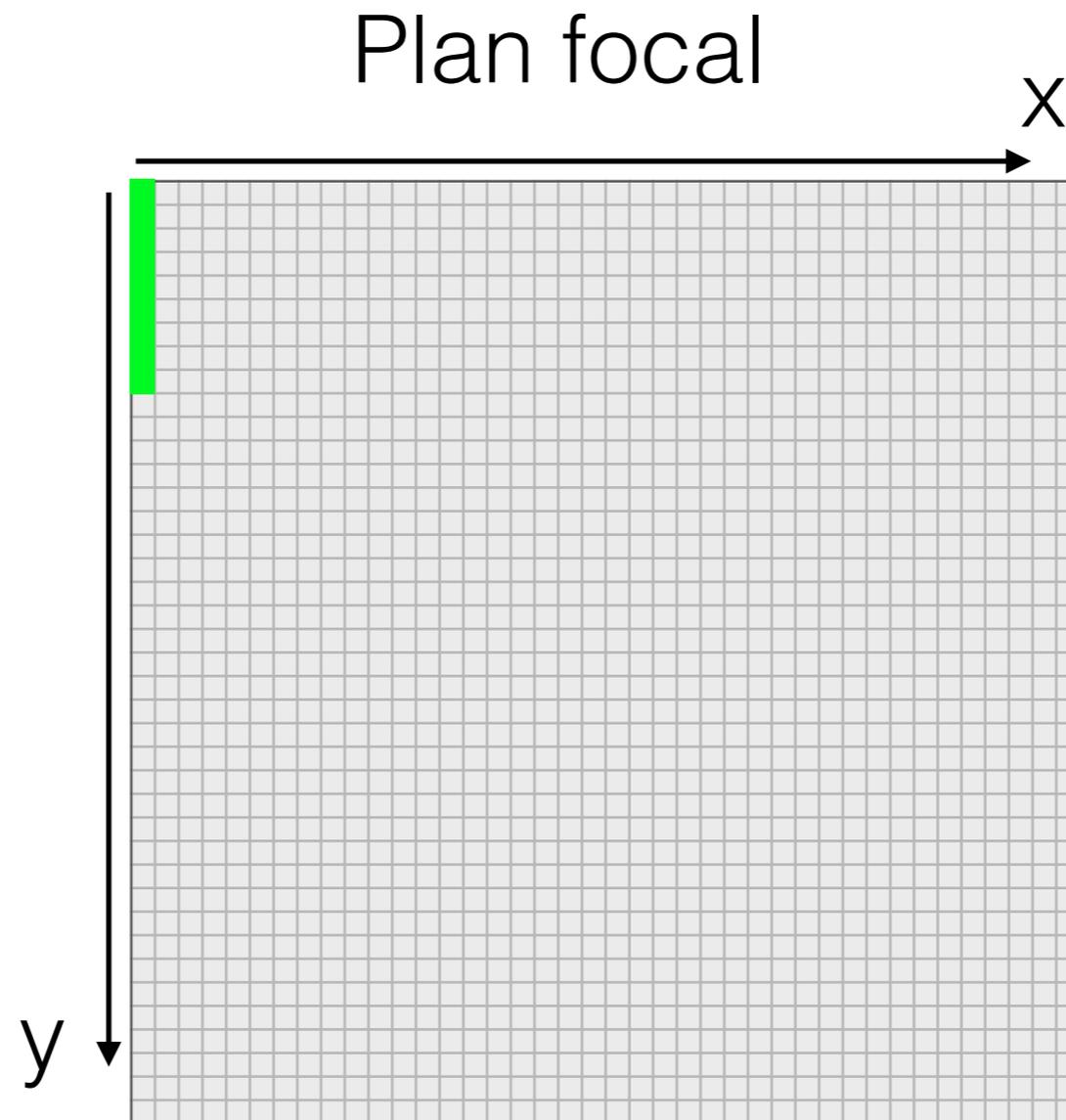
# Microscope confocal

*(Minsky 1953 , commercialisé en ~1980)*



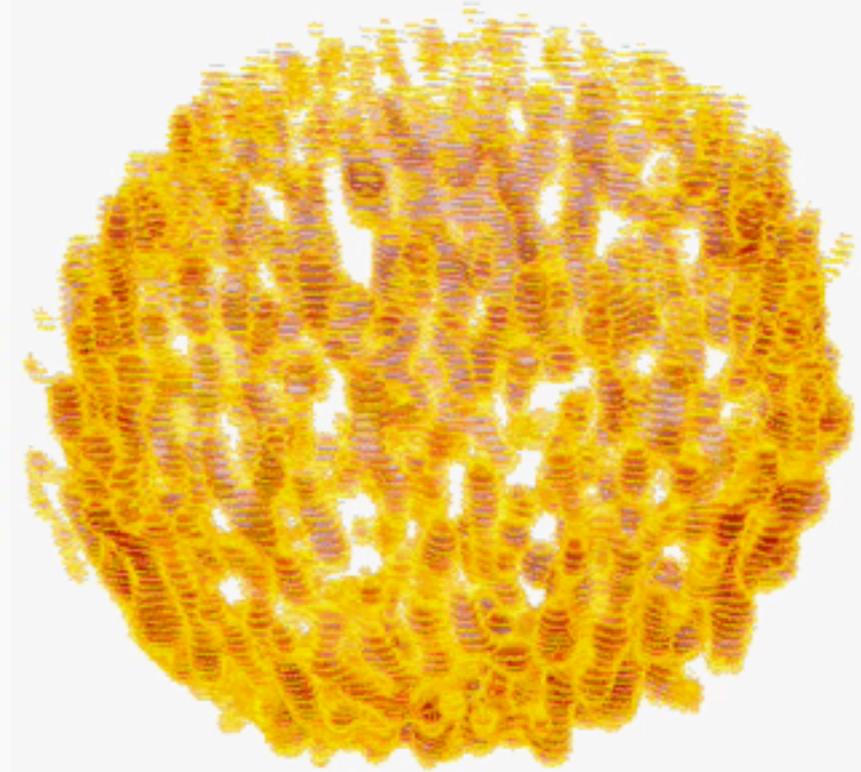
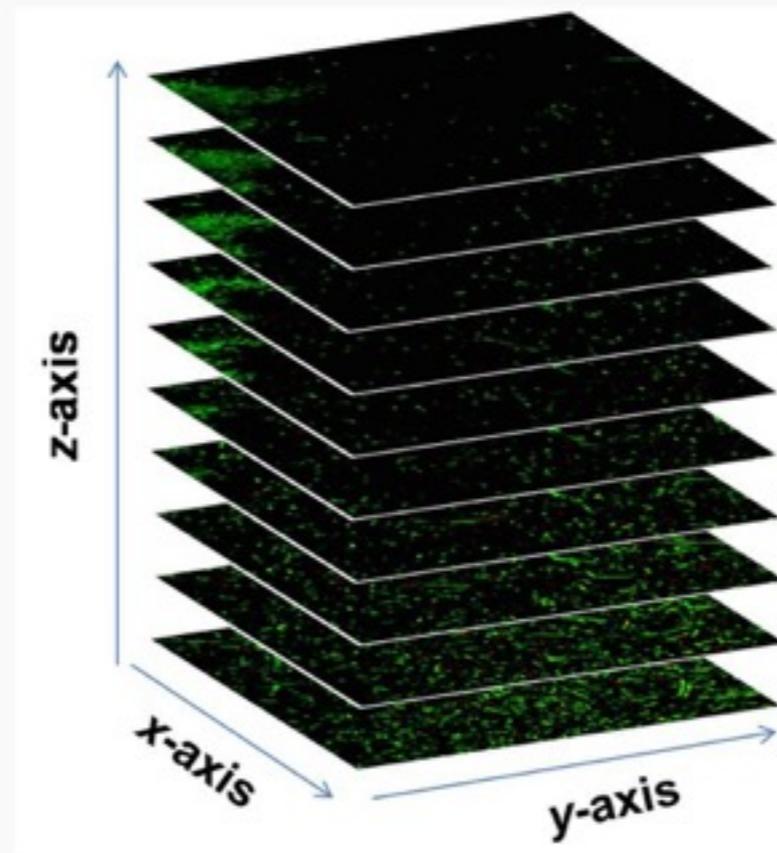
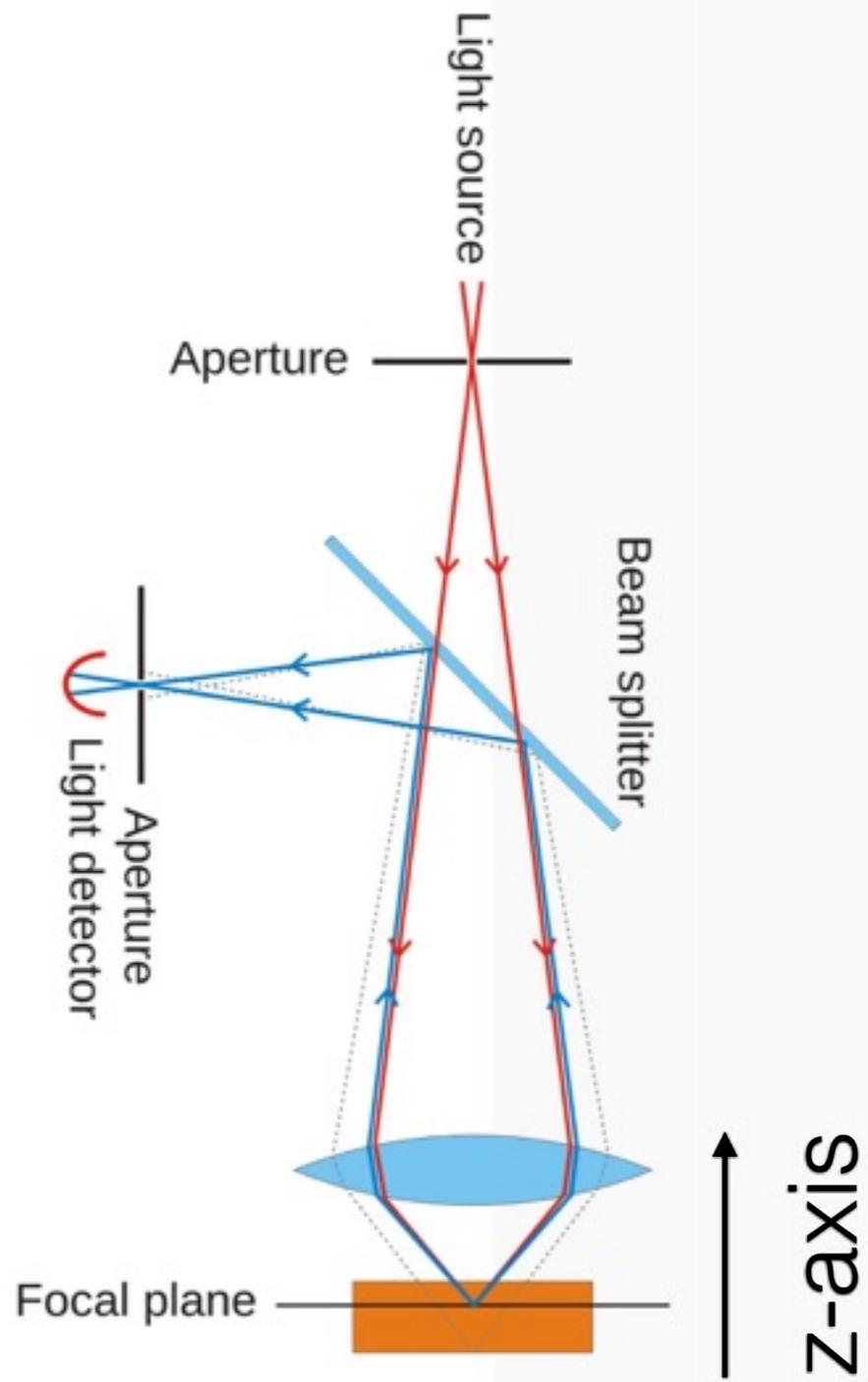
# Microscope confocal

balayage laser

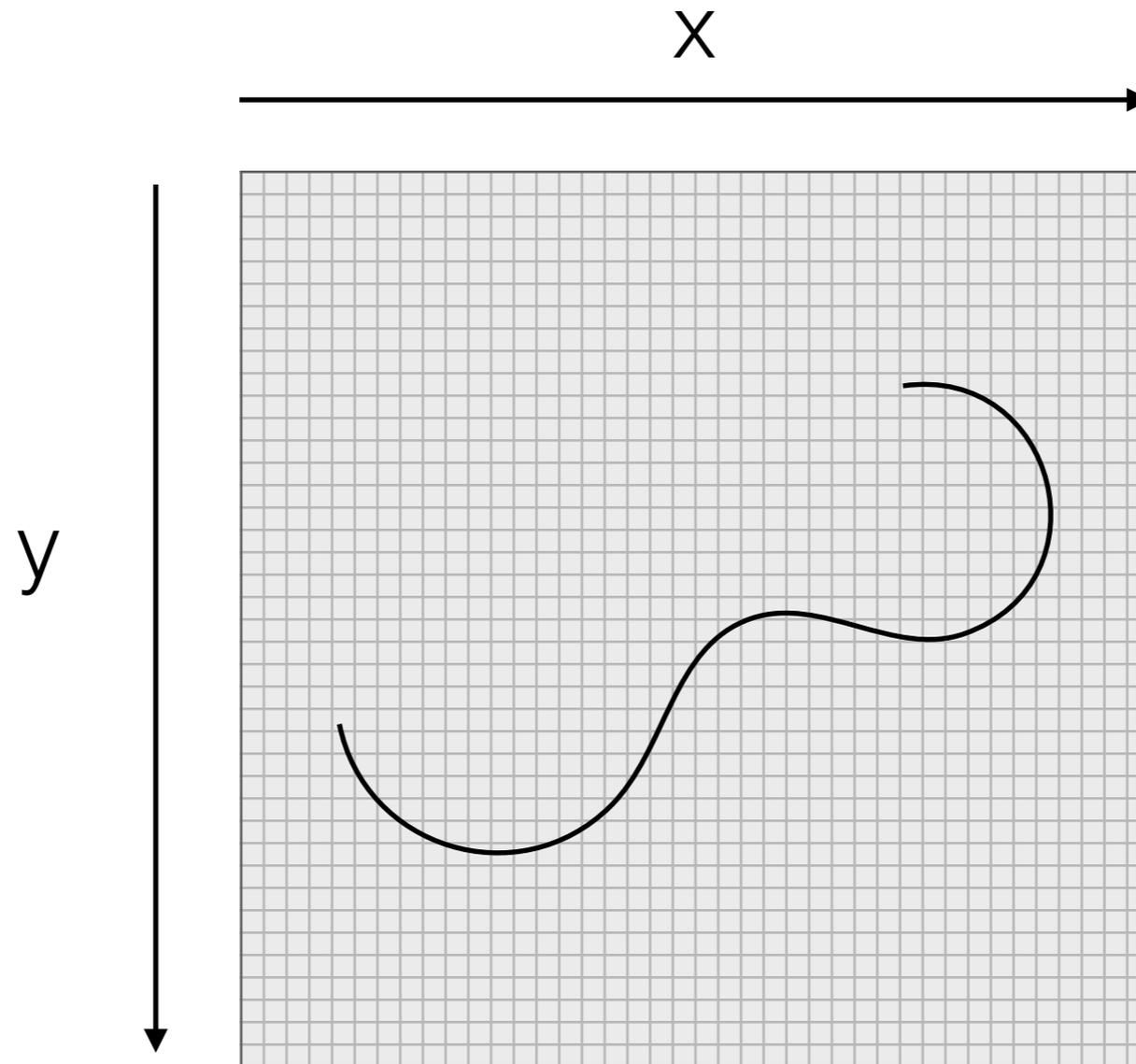


# Microscope confocal

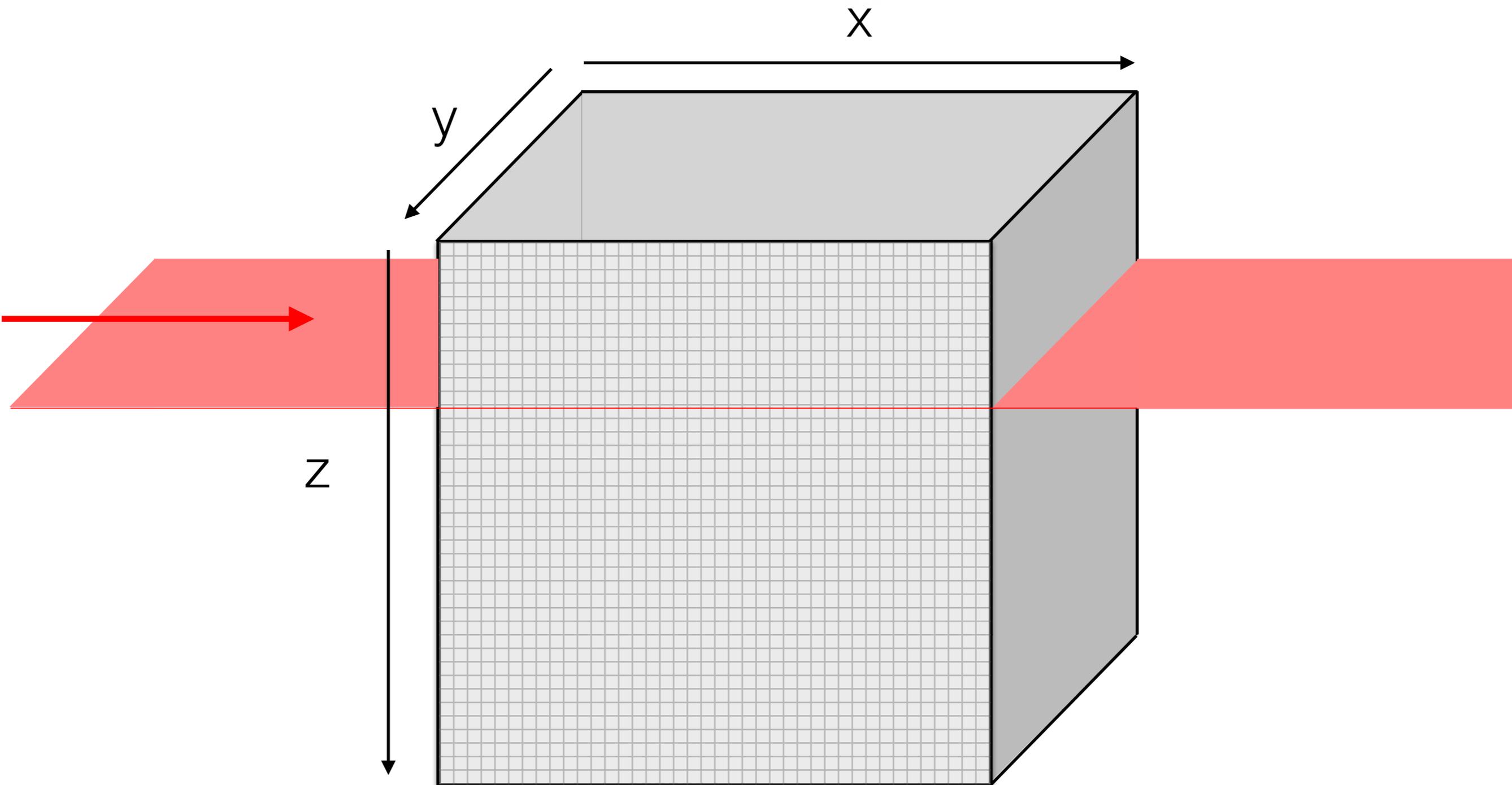
Image 3D

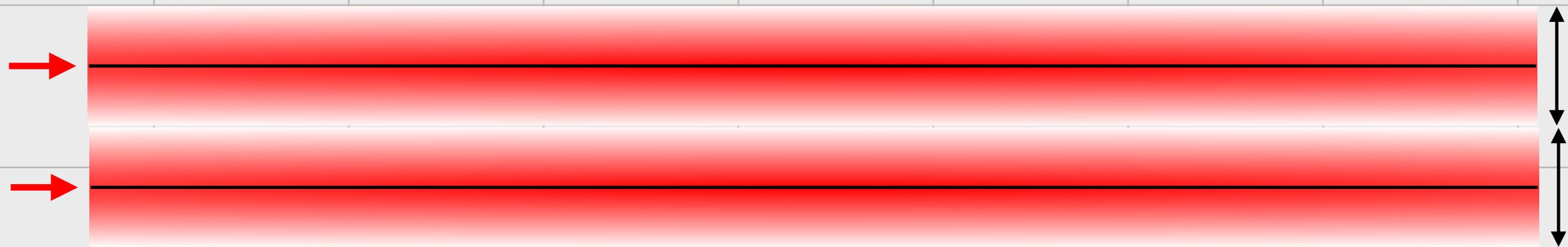


# Découpage Image 2D



# Découpage Image 3D

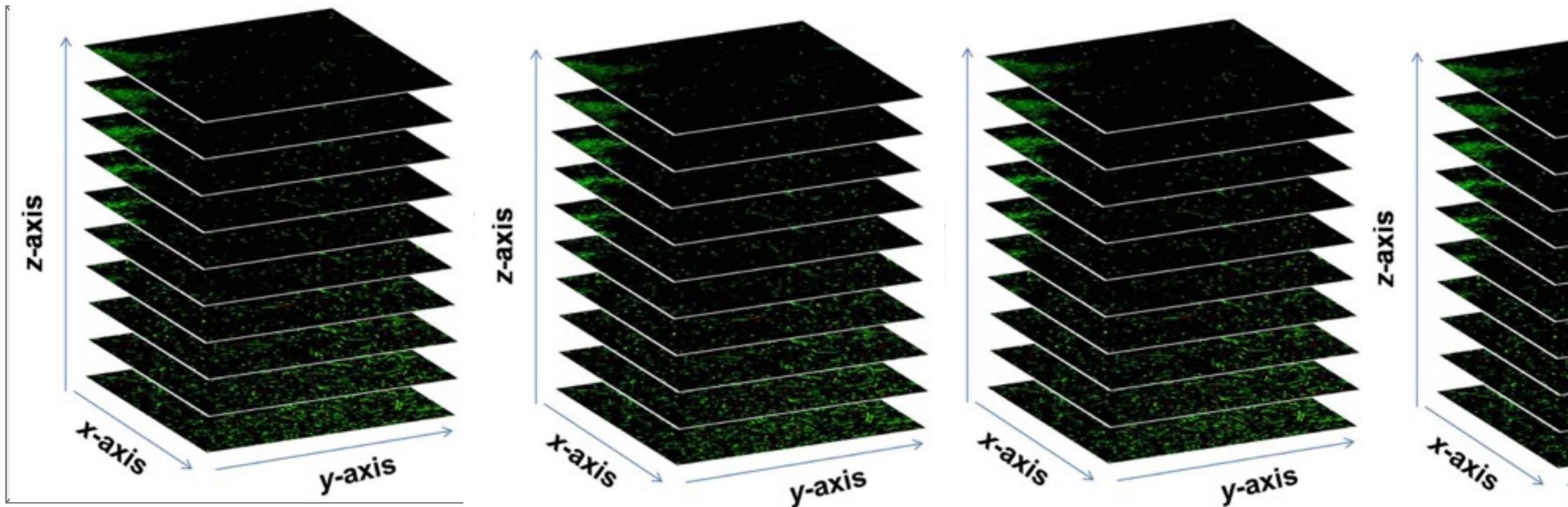




# Microscope confocal

Image 4D

2min



temps

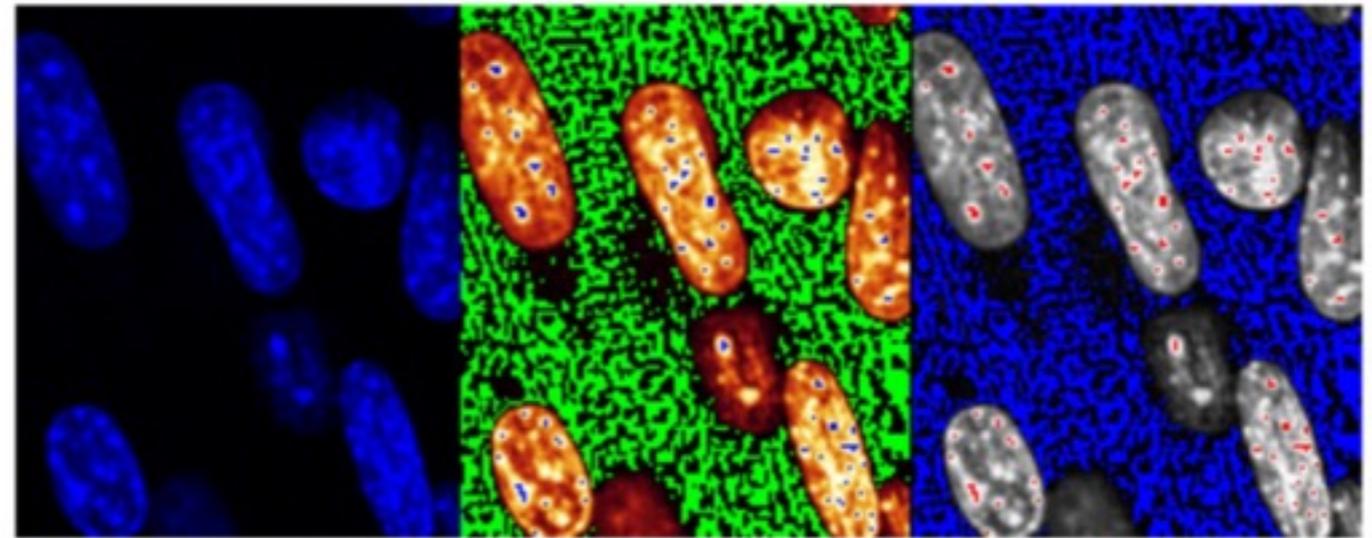
# Microscope confocal



# Microscope confocal

Puissance Laser

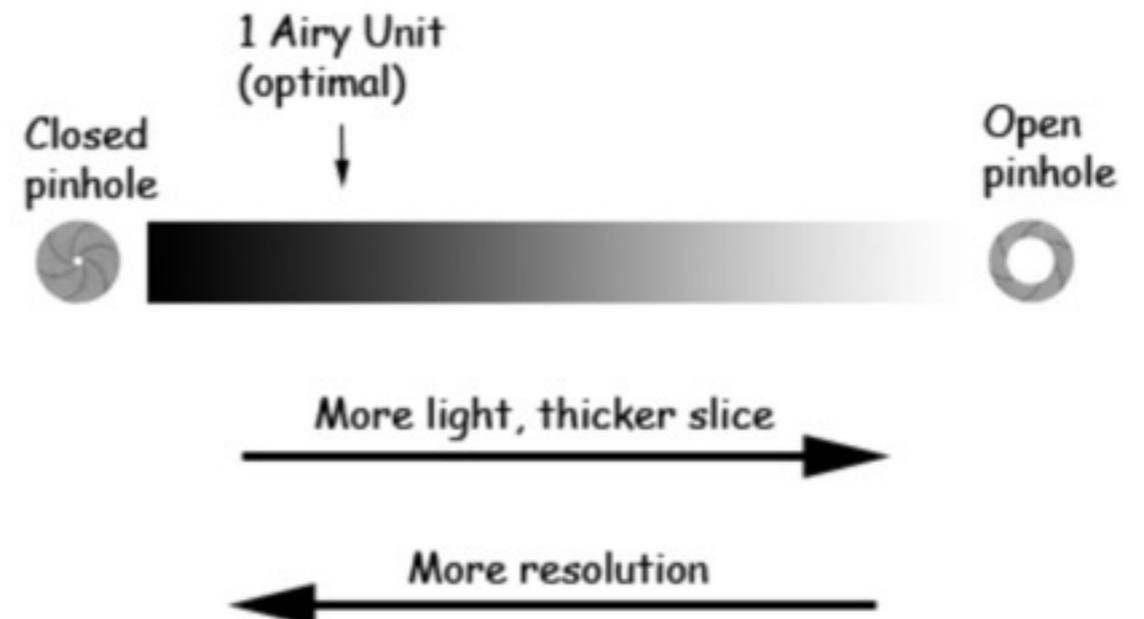
Sensibilité des  
détecteurs



Green = 0  
Blue = max

Blue = 0  
red = max

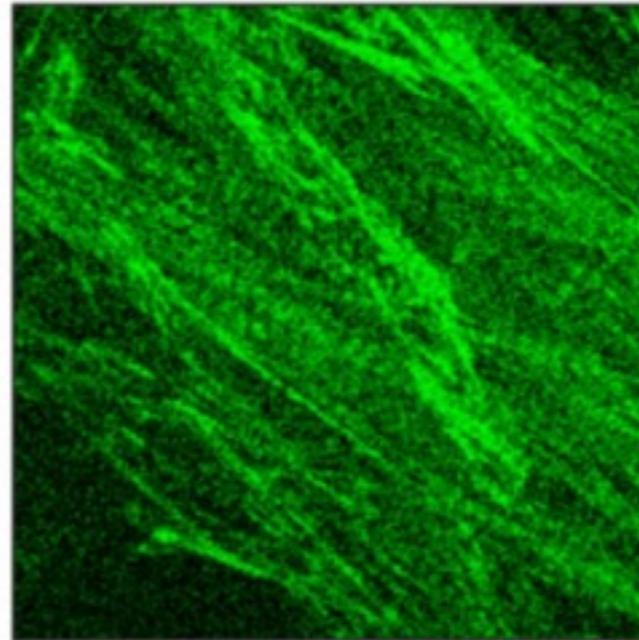
Ouverture



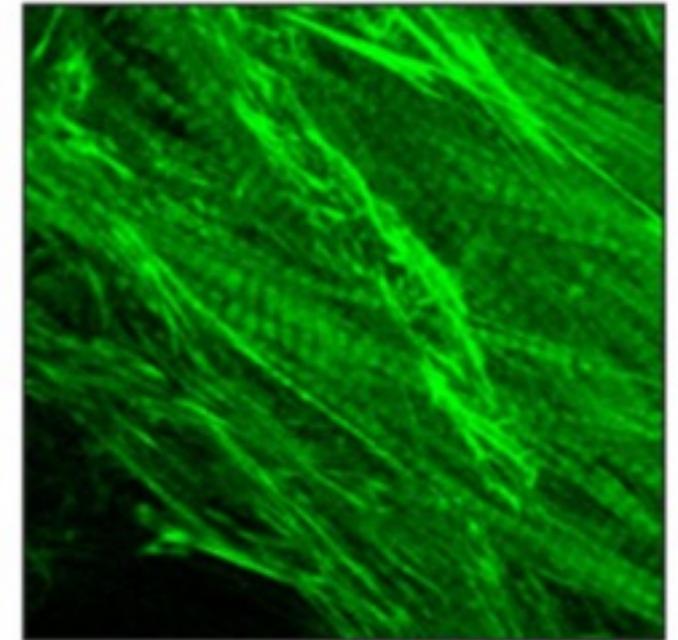
# Microscope confocal

Nombre de  
passes

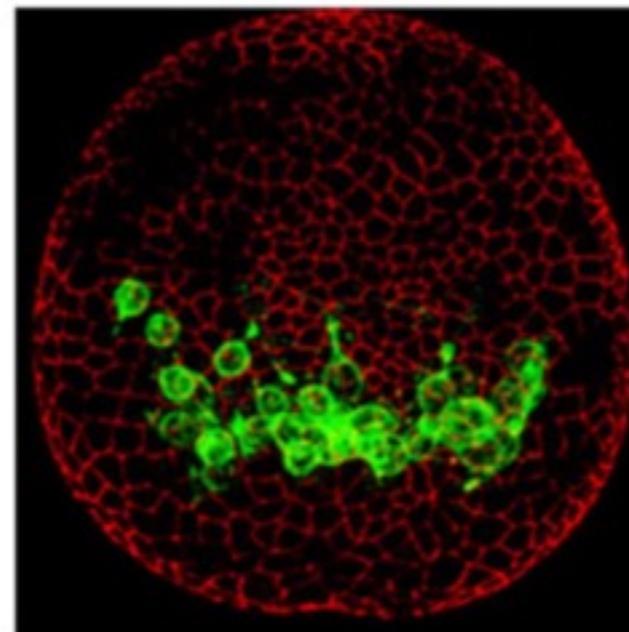
Average 1



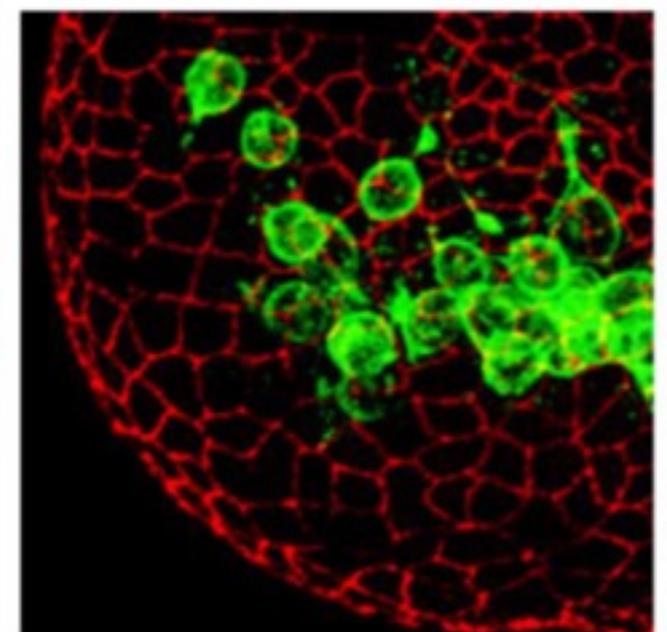
Line average 8



Zoom 1x



Zoom 2x



Zoom

# Microscope confocal

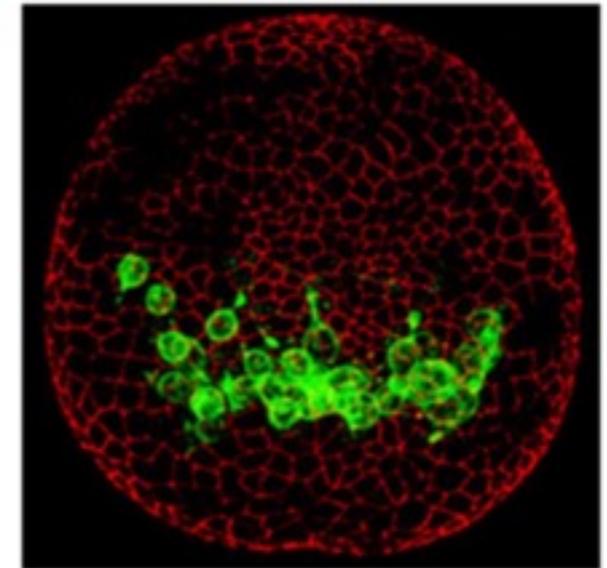
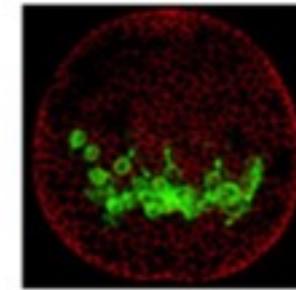
Nombre de pixel

$$\Delta xy$$

Same area, different  
number of pixels

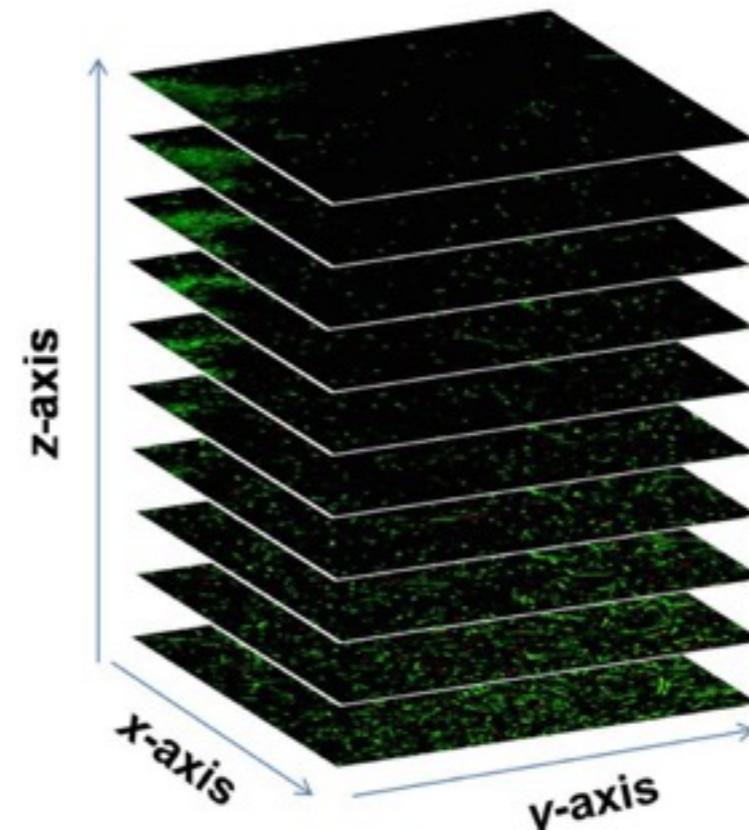
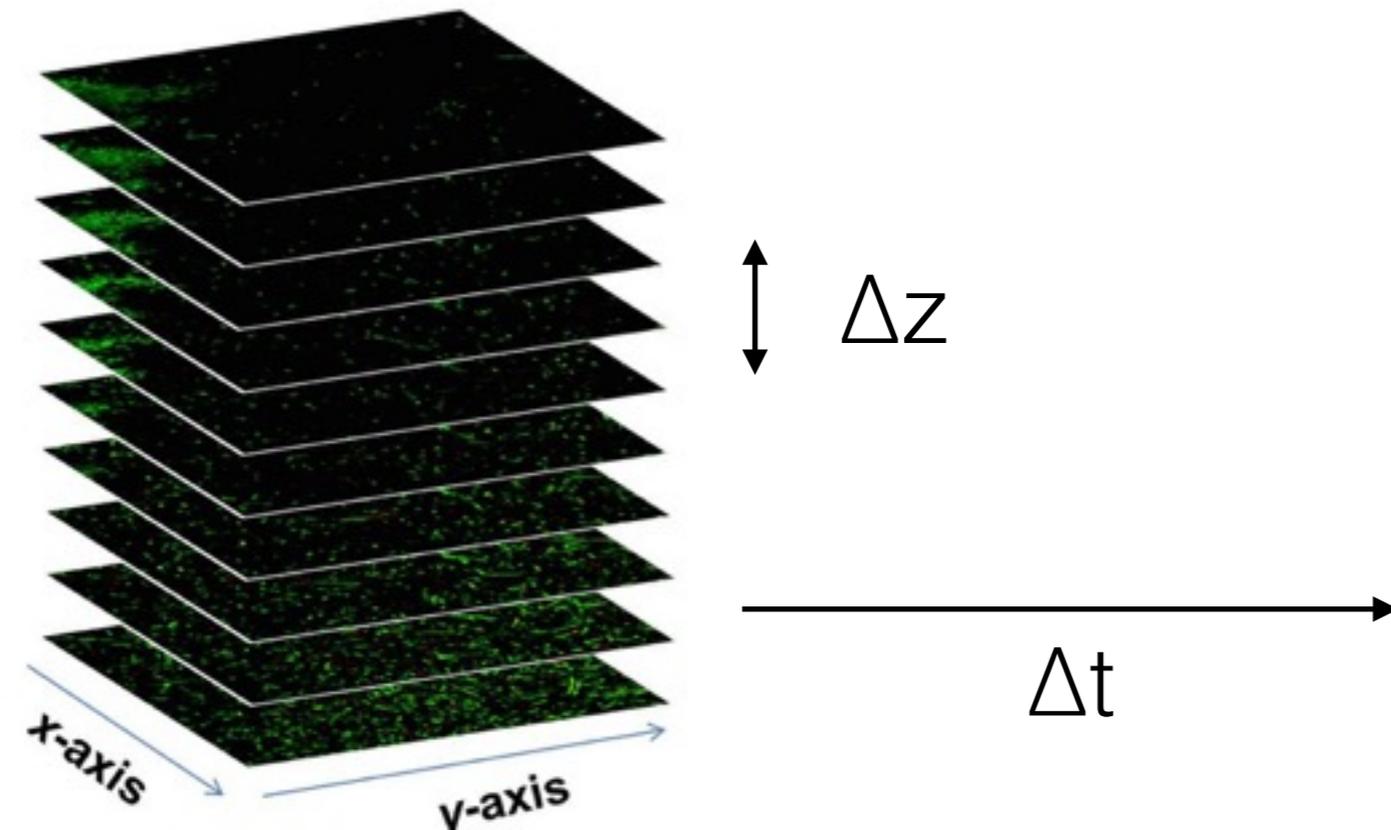
512 by 512

1024 by 1024



About 0.5 MB

About 2.0 MB



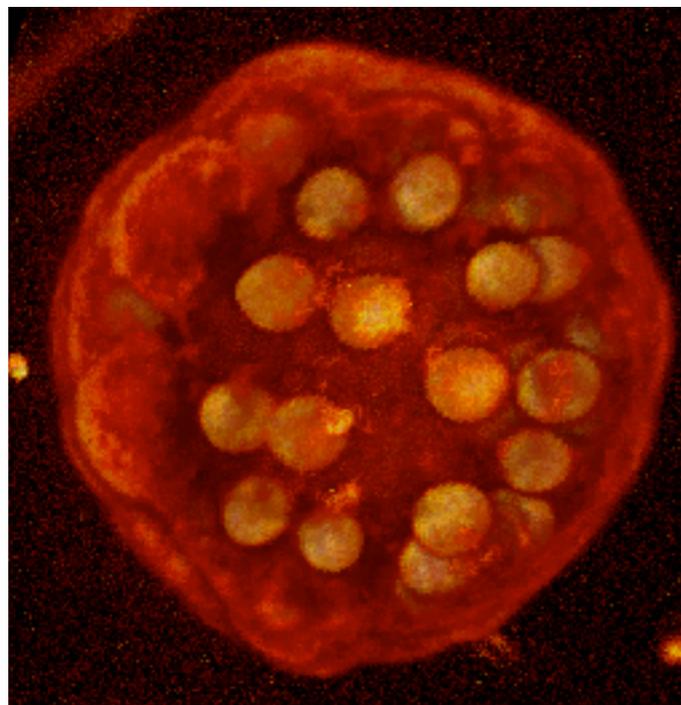
# Microscope confocal

Objectifs



Photoblanchiment  
Phototoxicité

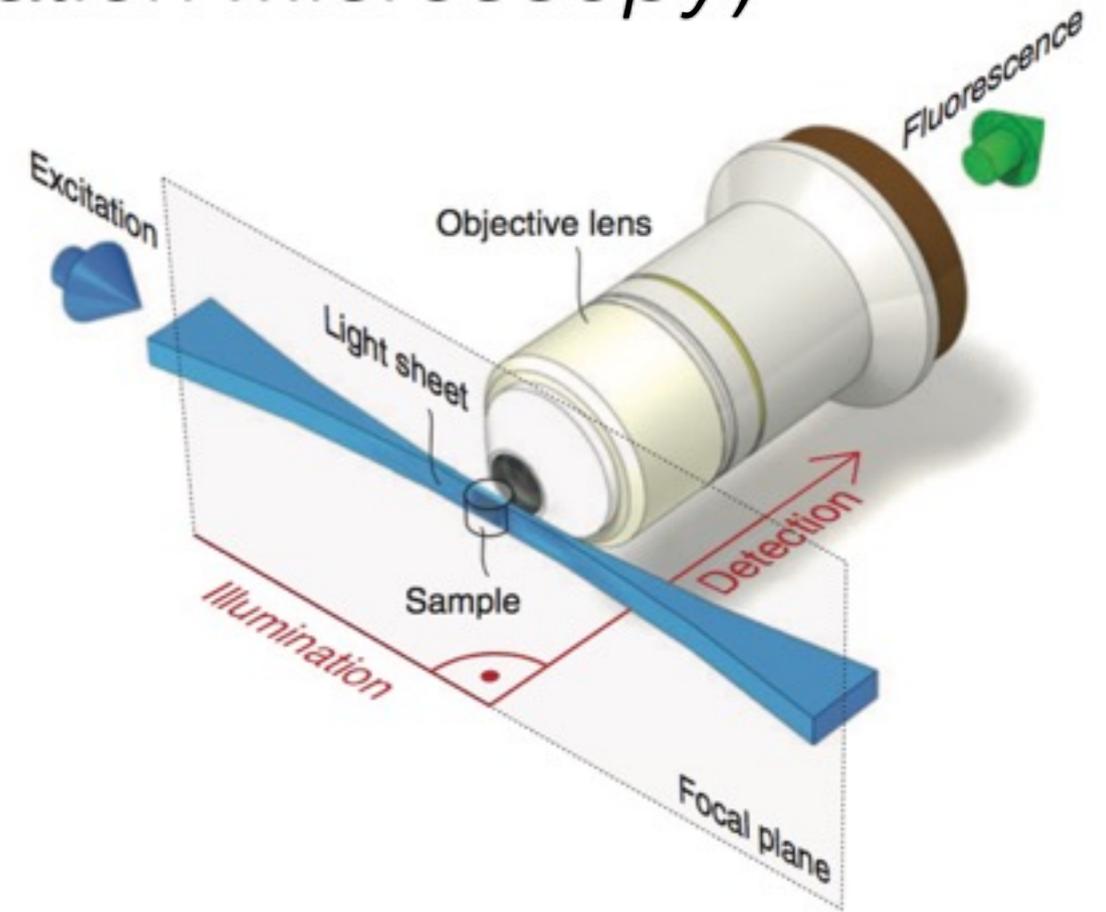
Les embryons bougent



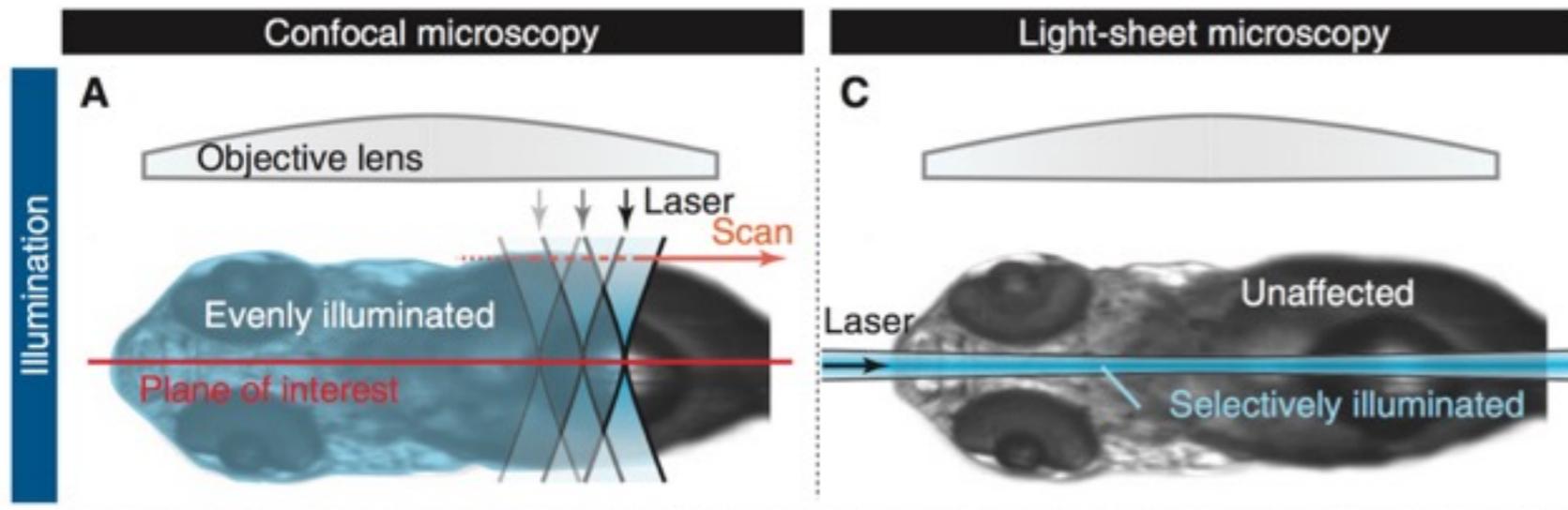
$$\# \text{ photons} \sim \text{density} \cdot \text{object size} \cdot \text{resolution} \cdot \text{signal} \cdot \text{time period} \cdot \text{repetitions}$$

# Microscope SPIM (*Selective Plane Illumination Microscopy*)

Beaucoup plus rapide



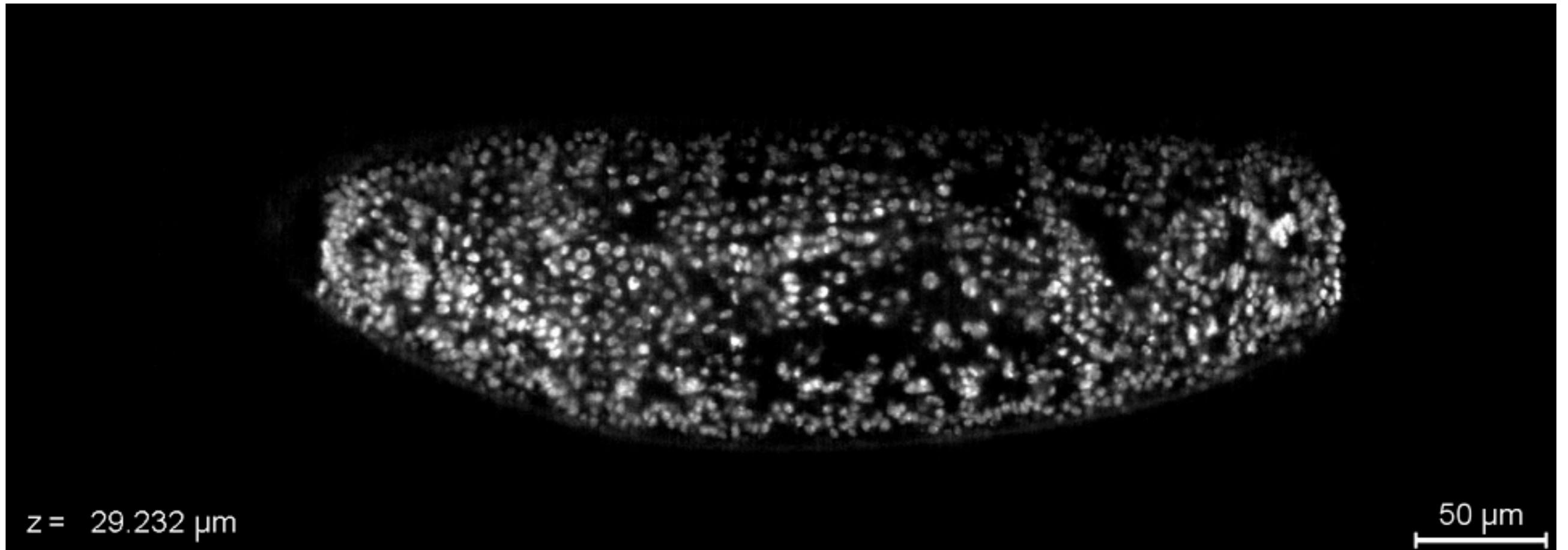
Huisken 2004



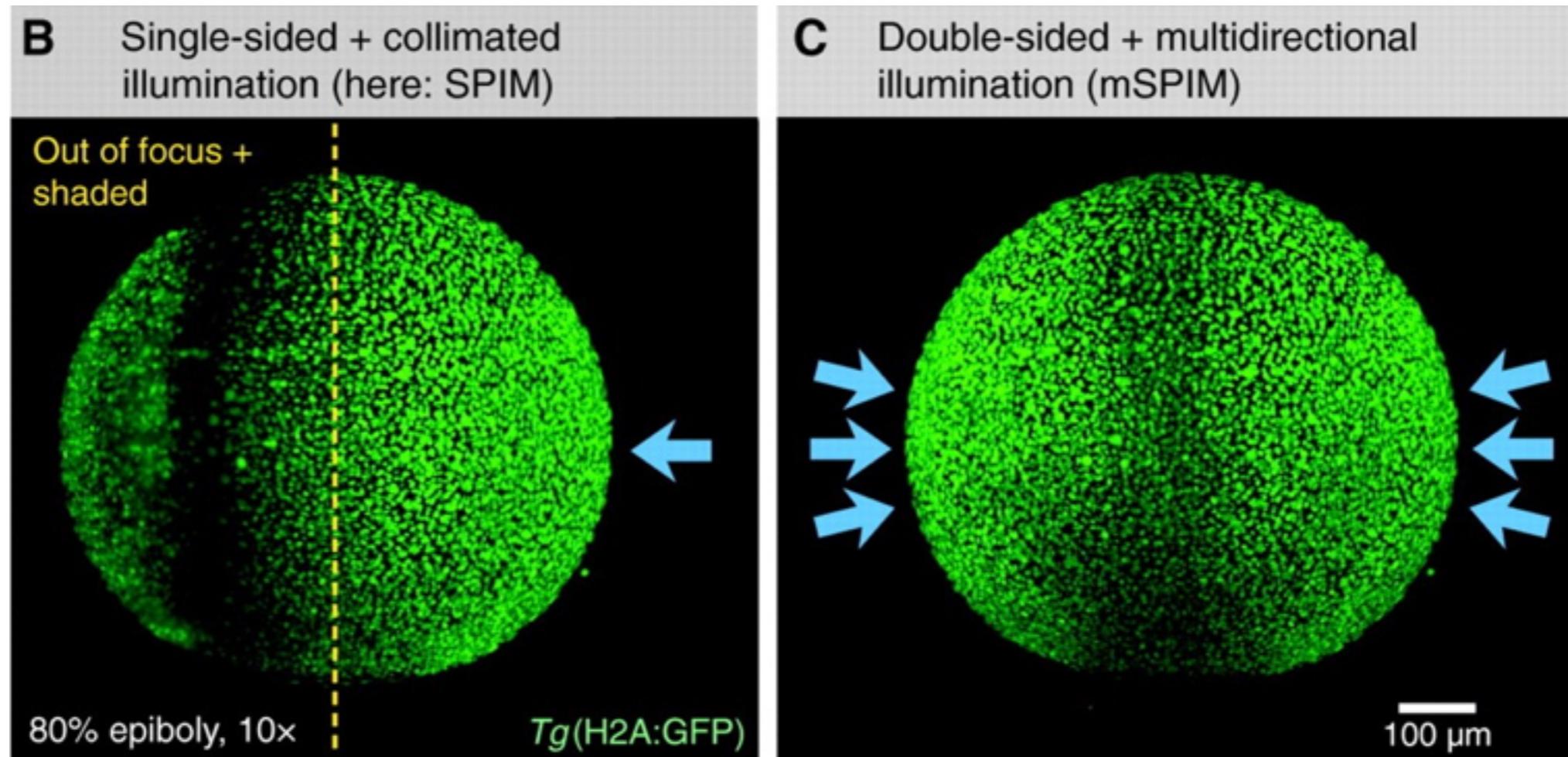
Moins d'illumination

Huisken 2009

Microscope SPIM  
(*Selective Plane Illumination Microscopy*)

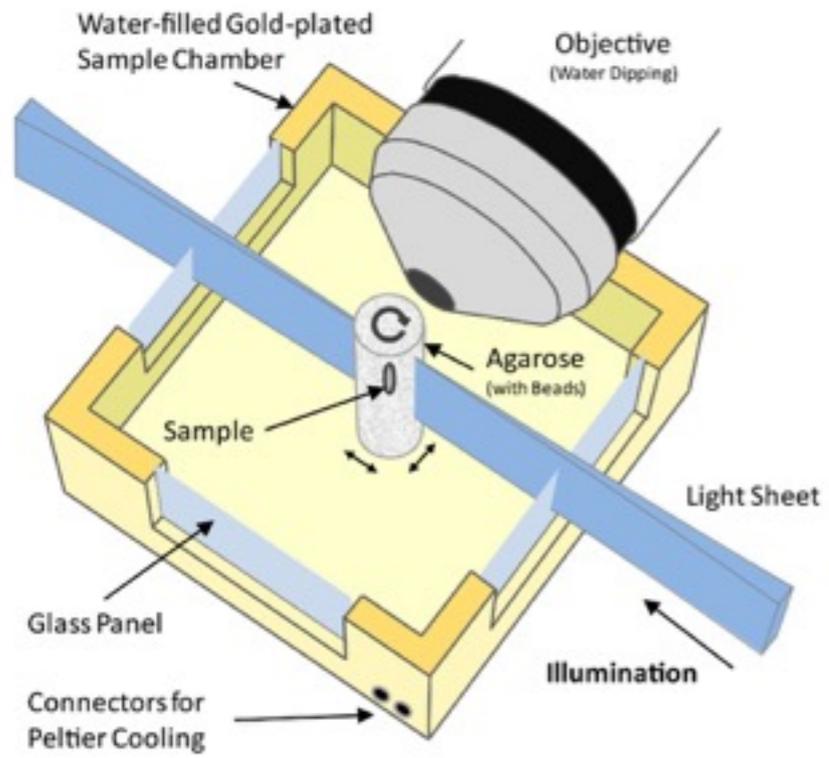


# SPIM : Multiples Illuminations

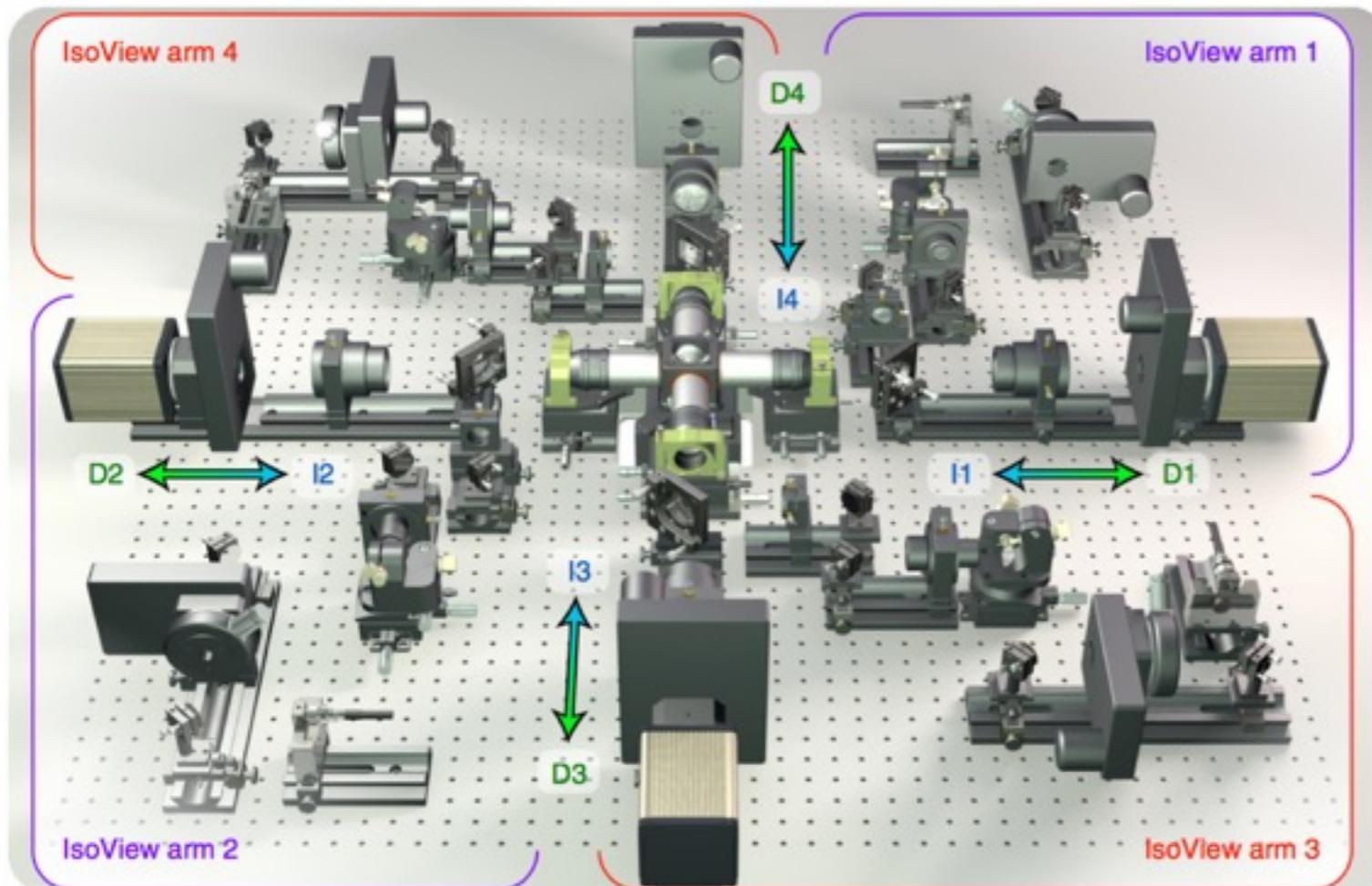
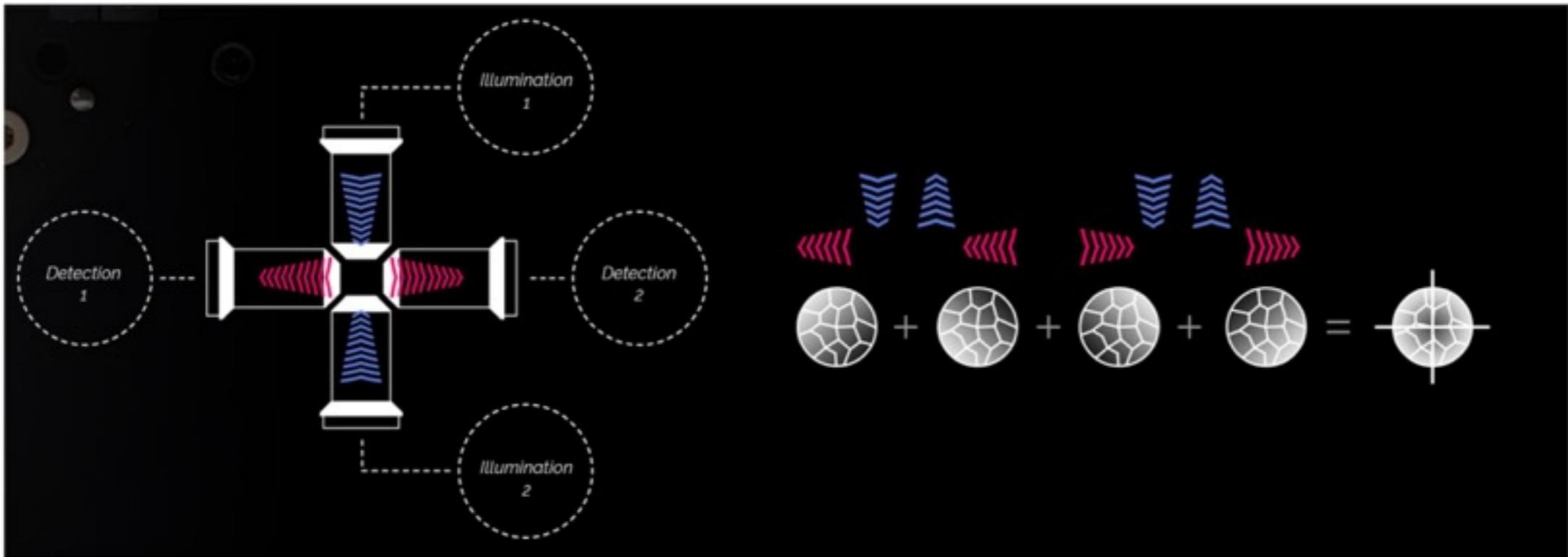


Huisken 2009

# SPIM : Multiples Angles

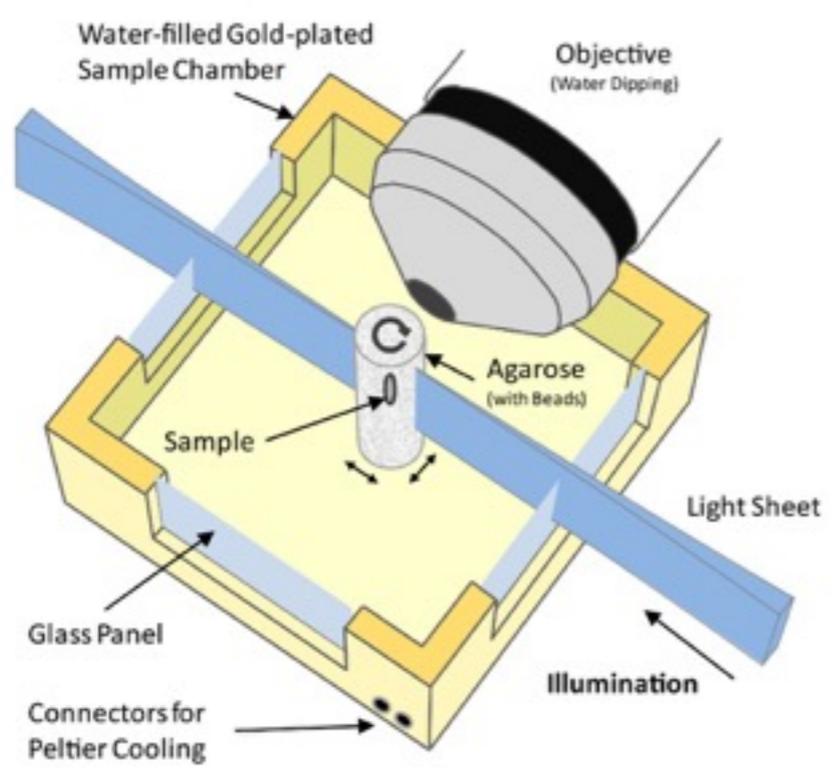
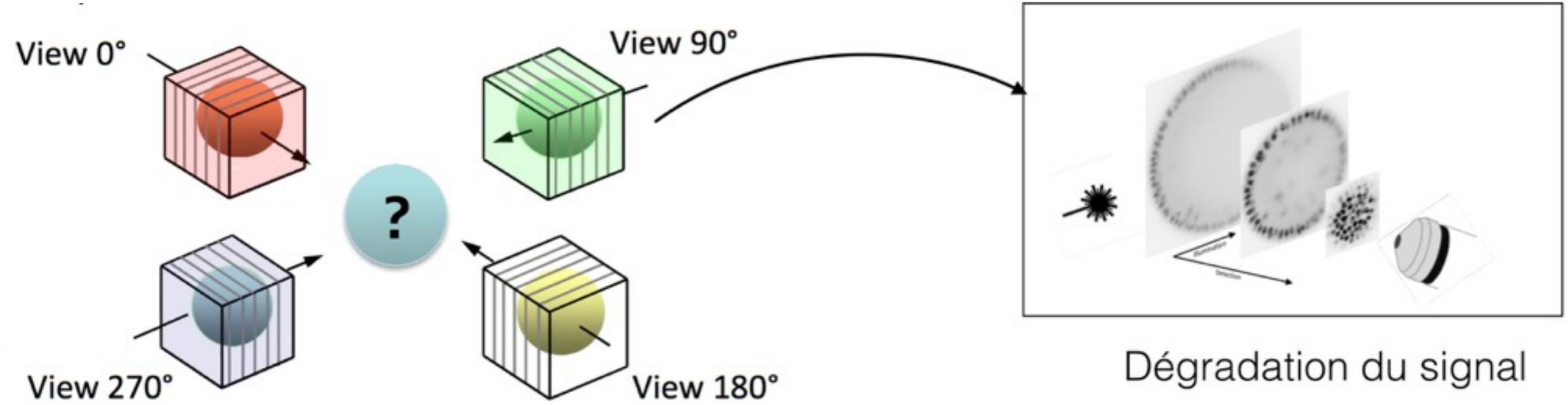


# SPIM : Multiples Vues



# SPIM : Recalage et Fusion

## **Spatial**



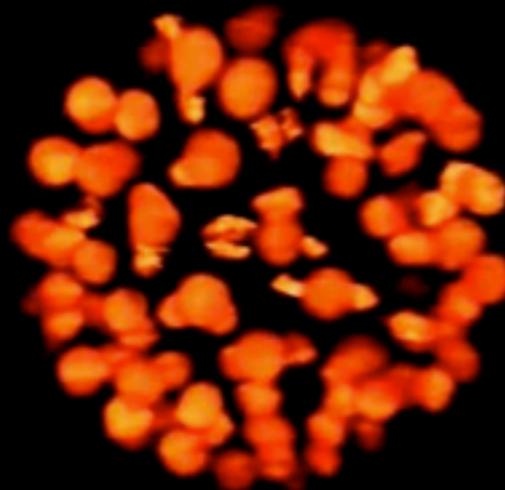
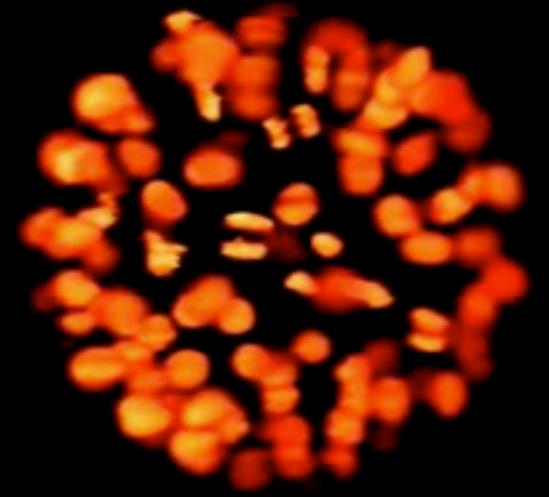
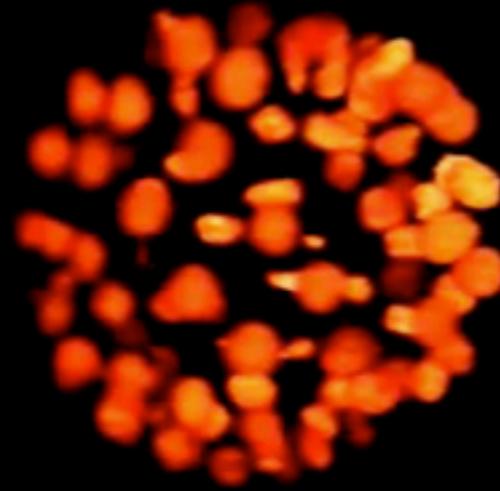
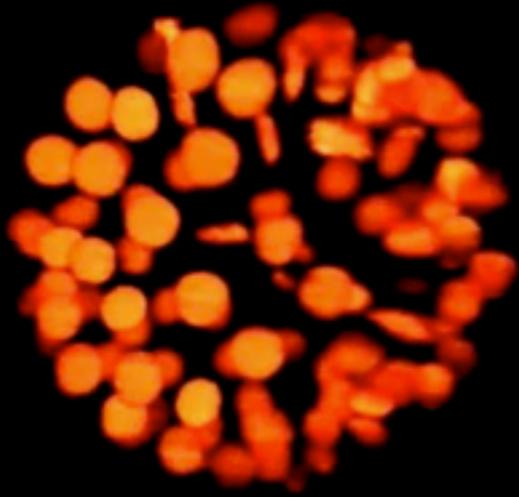
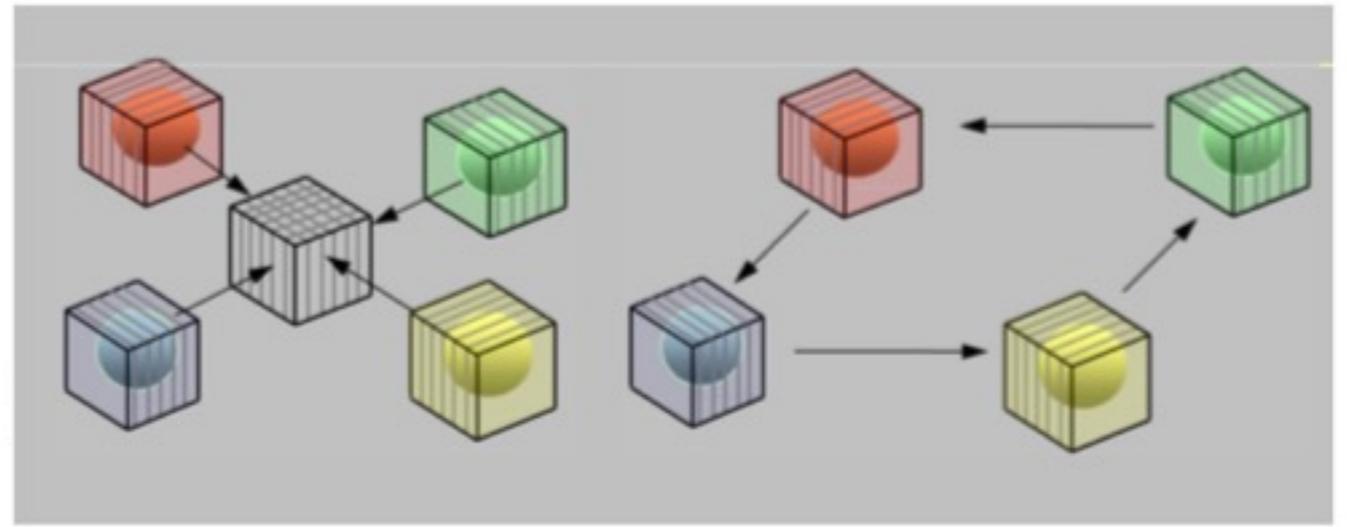
SPIM : Recalage et Fusion

**Temporel**

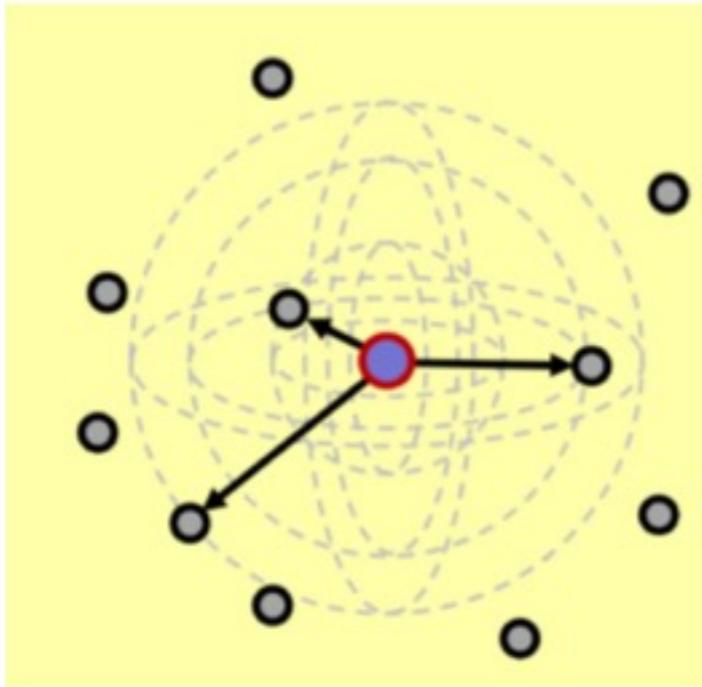
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# Recalage sur l'intensité

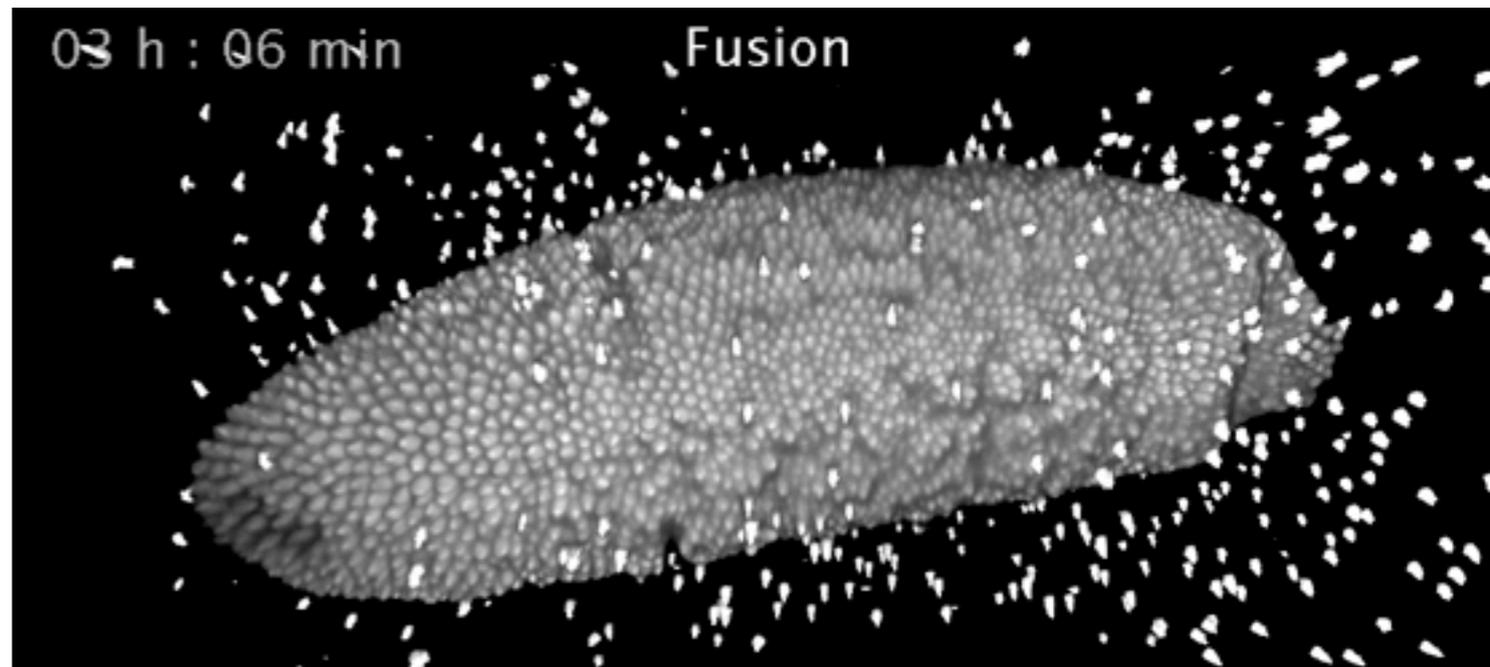
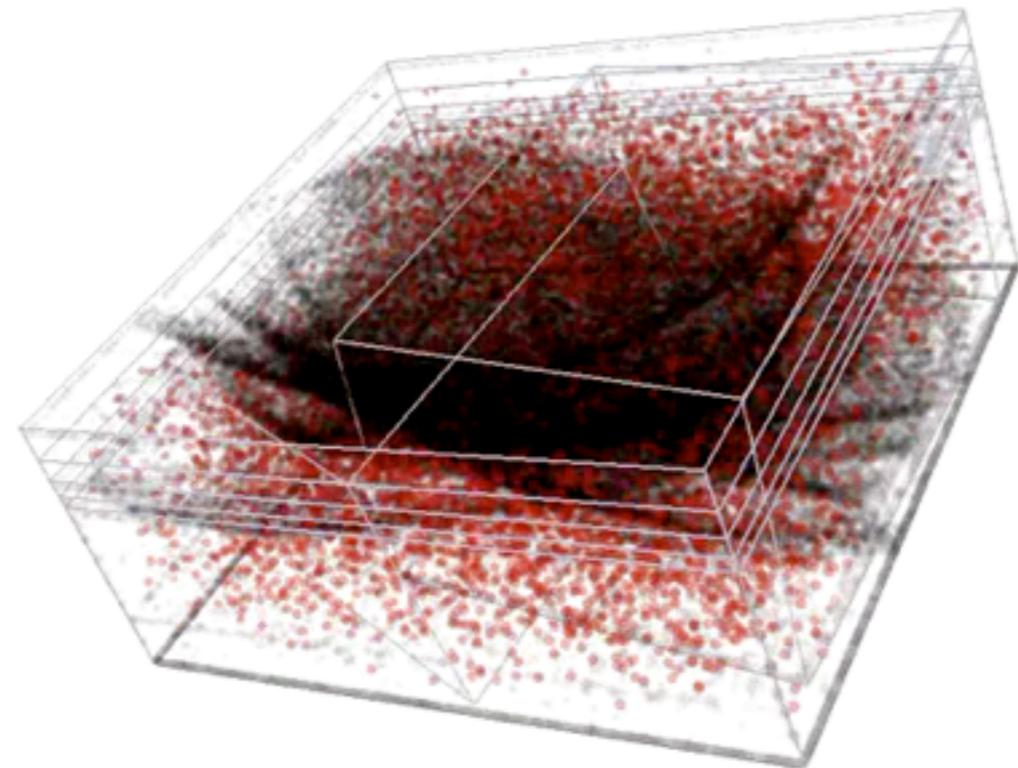
Très lent



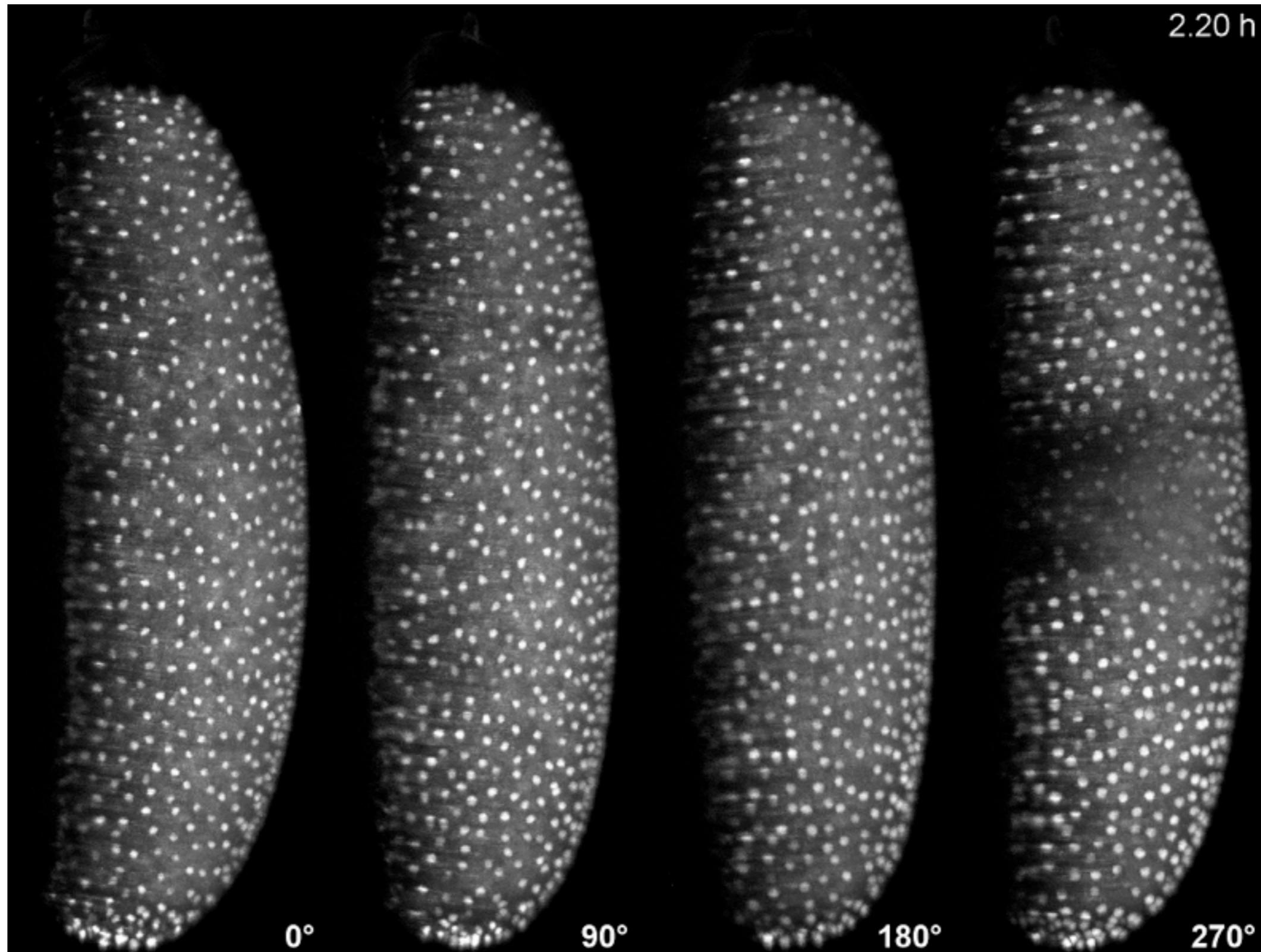
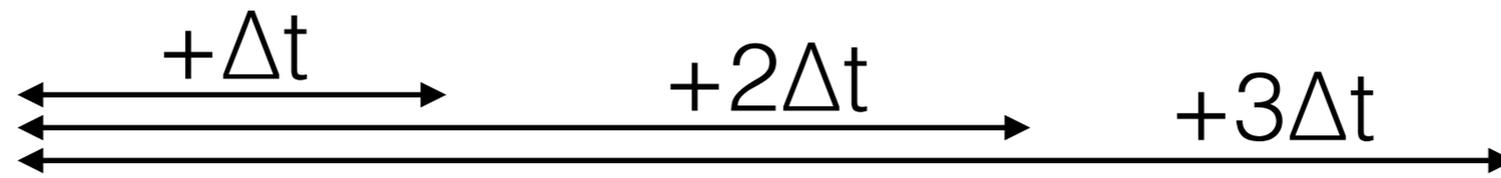
# Recalage à l'aide de repères



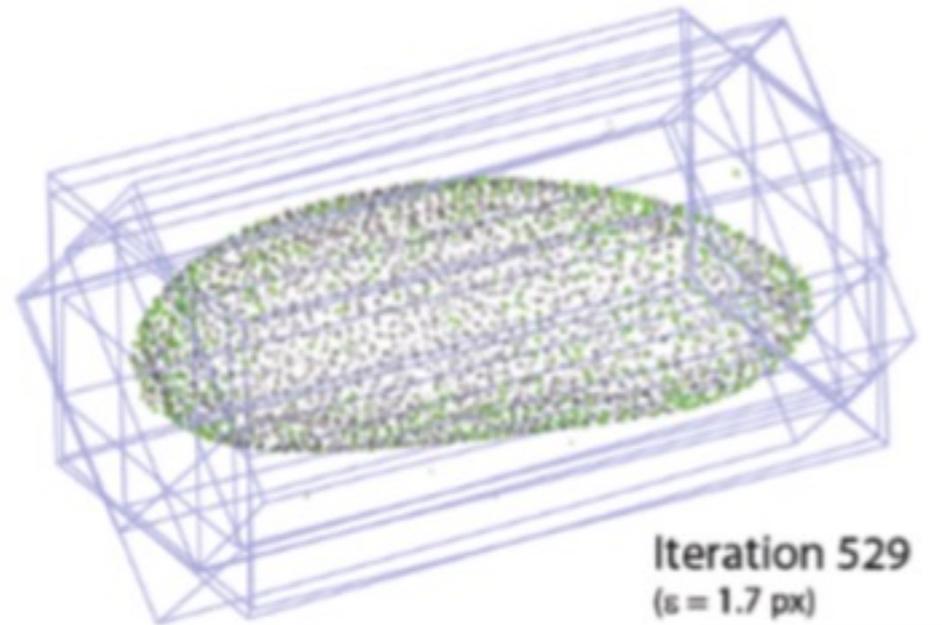
Très rapide, et très bon résultats mais nécessite d'injecter des micro-billes



# Problèmes du recalage à partir d'images



# Recalage à l'aide des segmentations

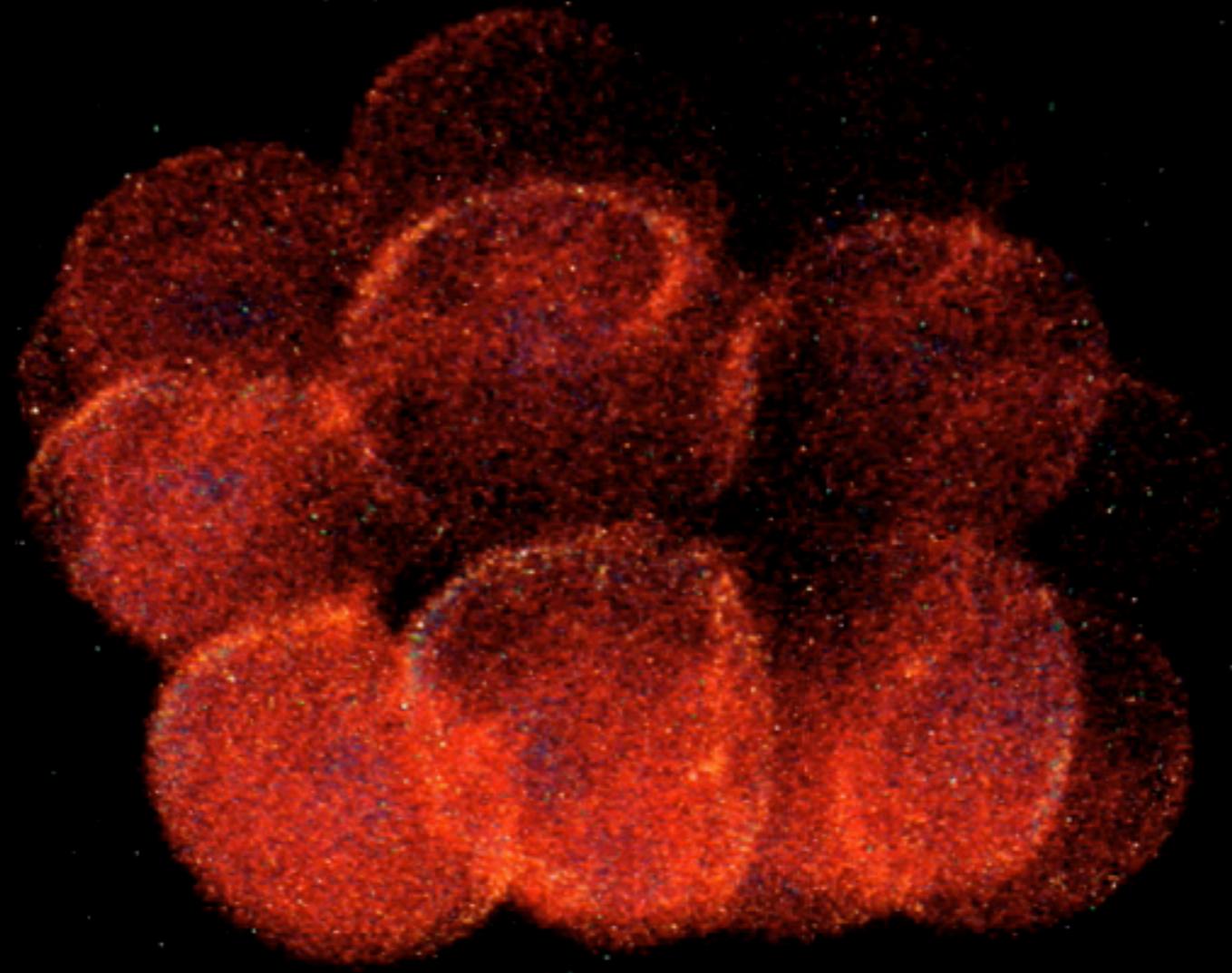


Rapide, mais dépend de la qualité des images et des algorithmes de segmentations

Qu'observe t'on ?

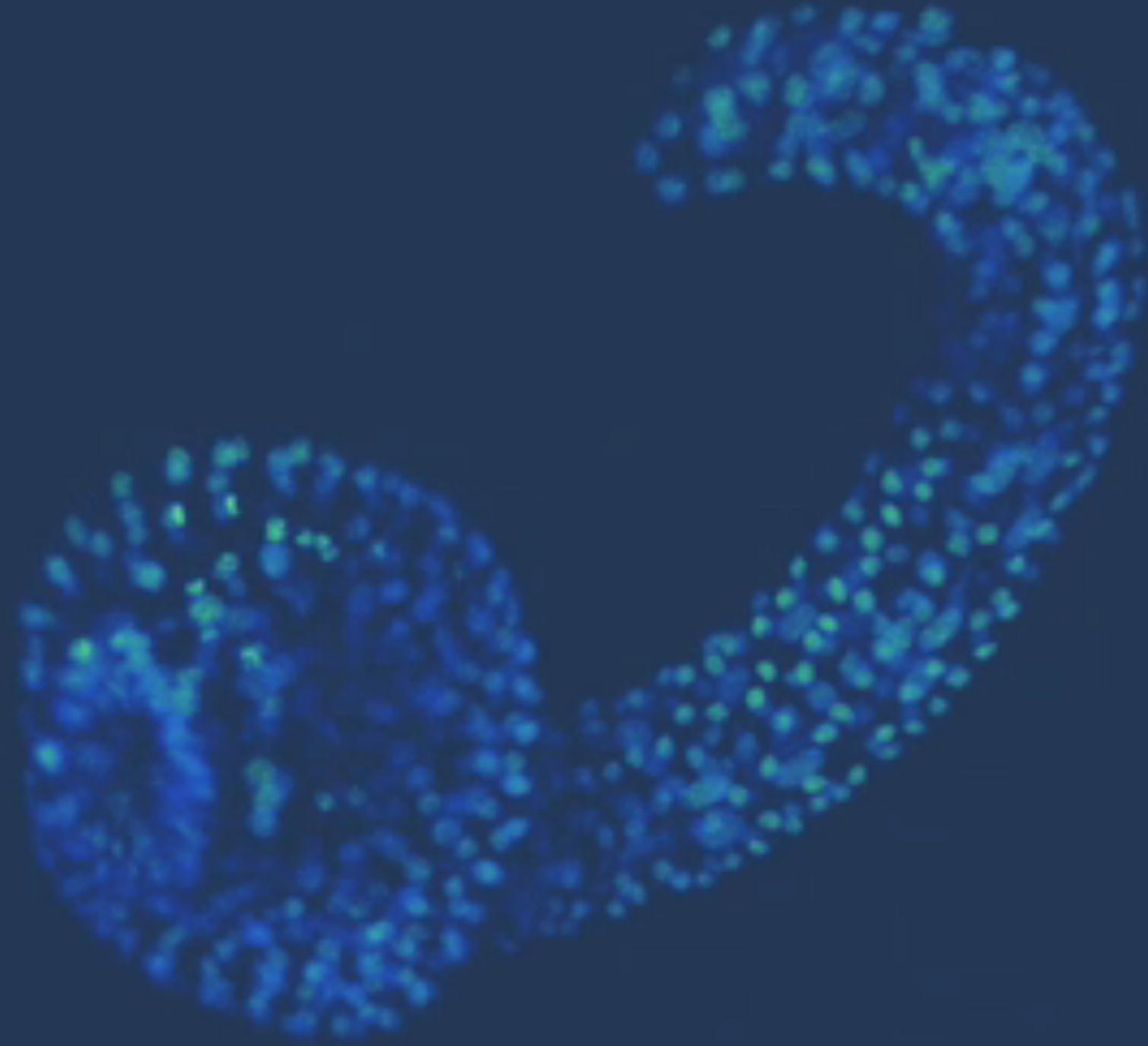


# Sea Urchin



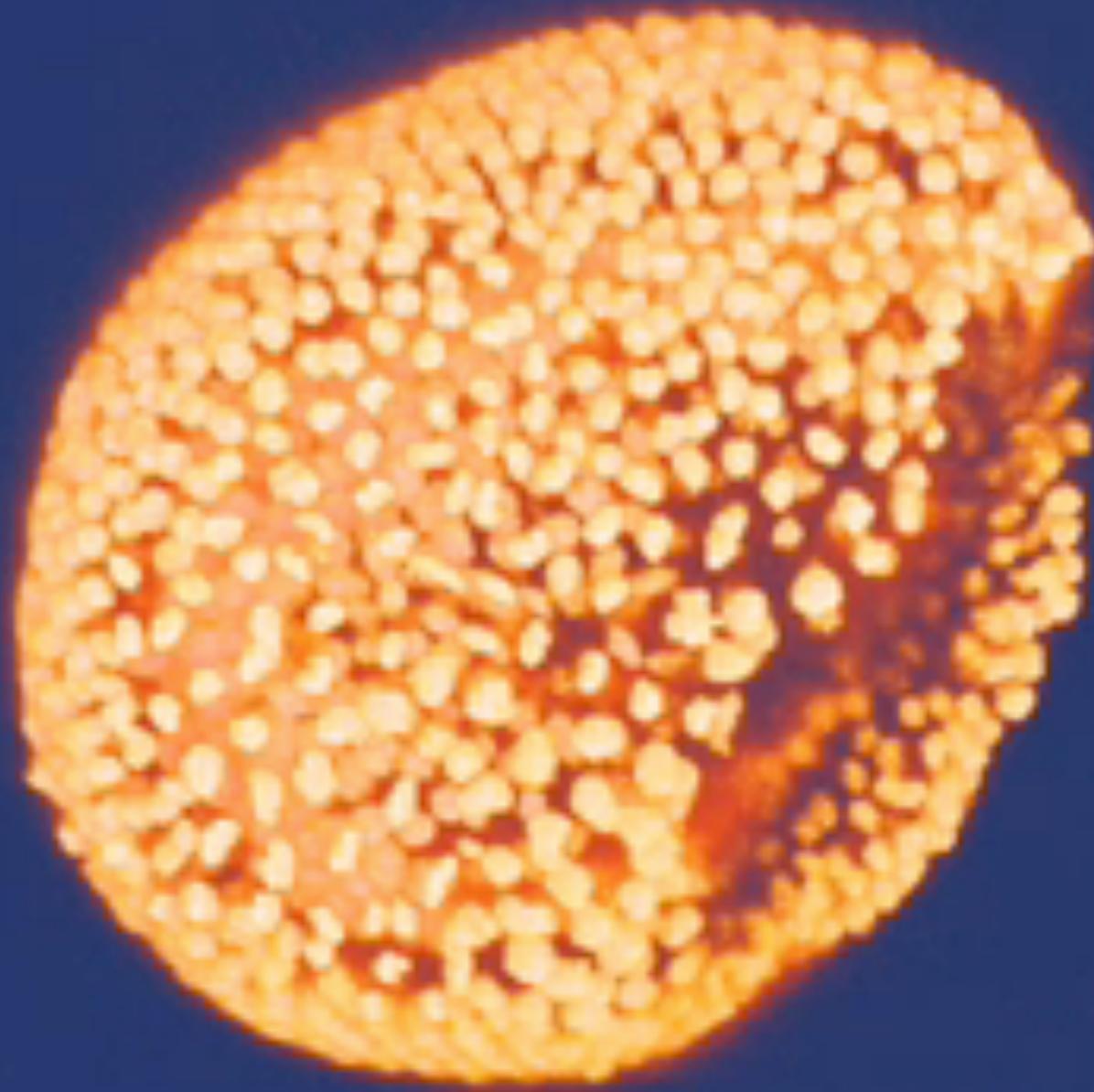


# Phallusia Mamillata



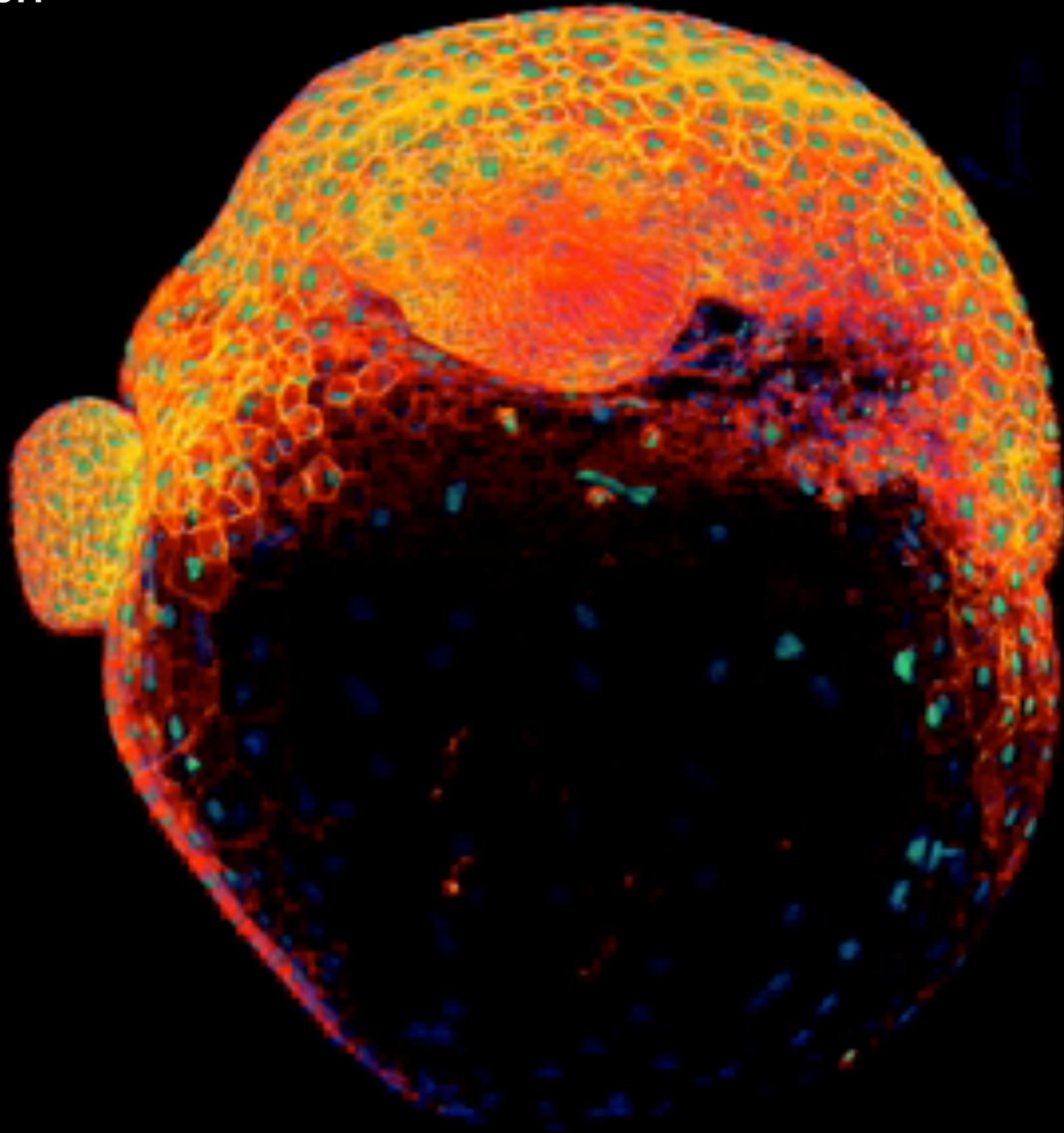


# Amphioxus



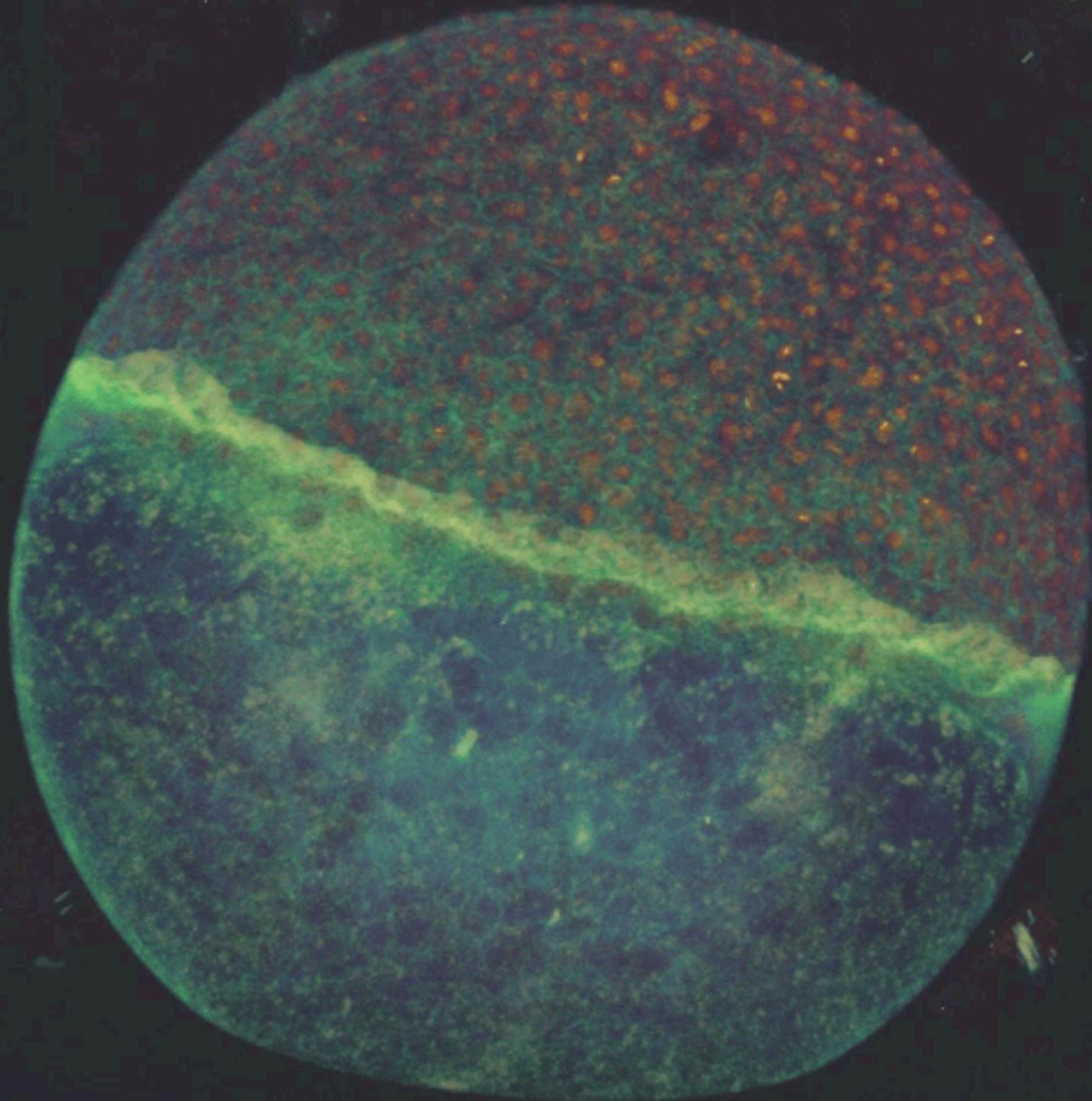


# Zebrafish



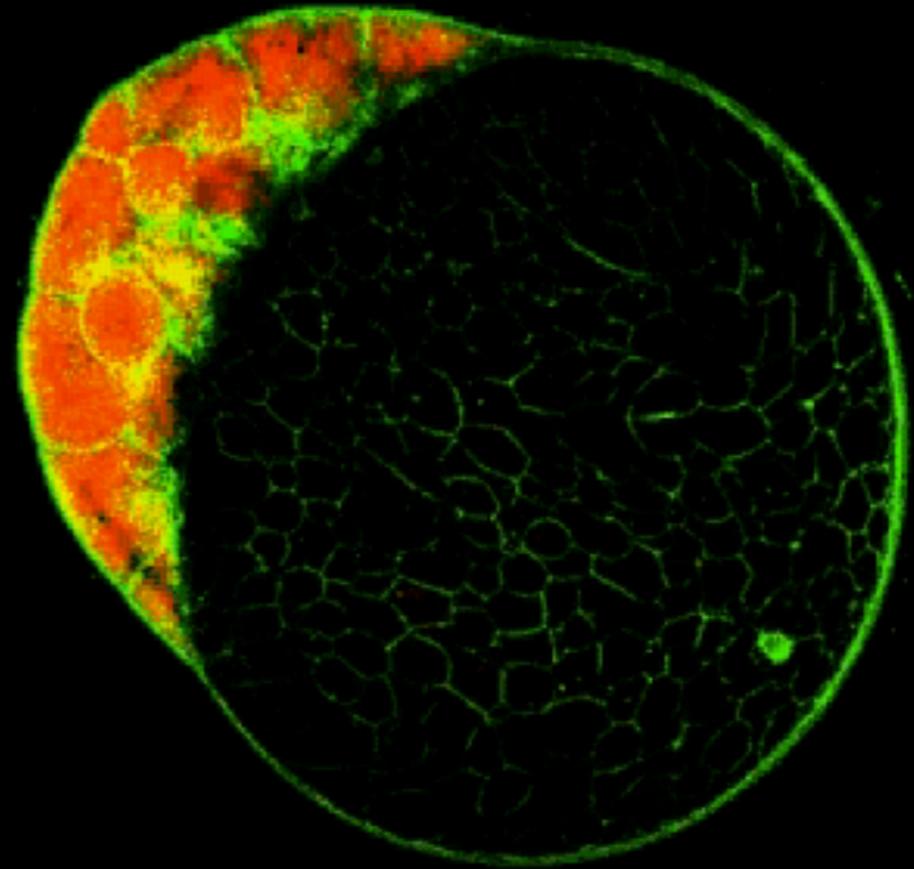
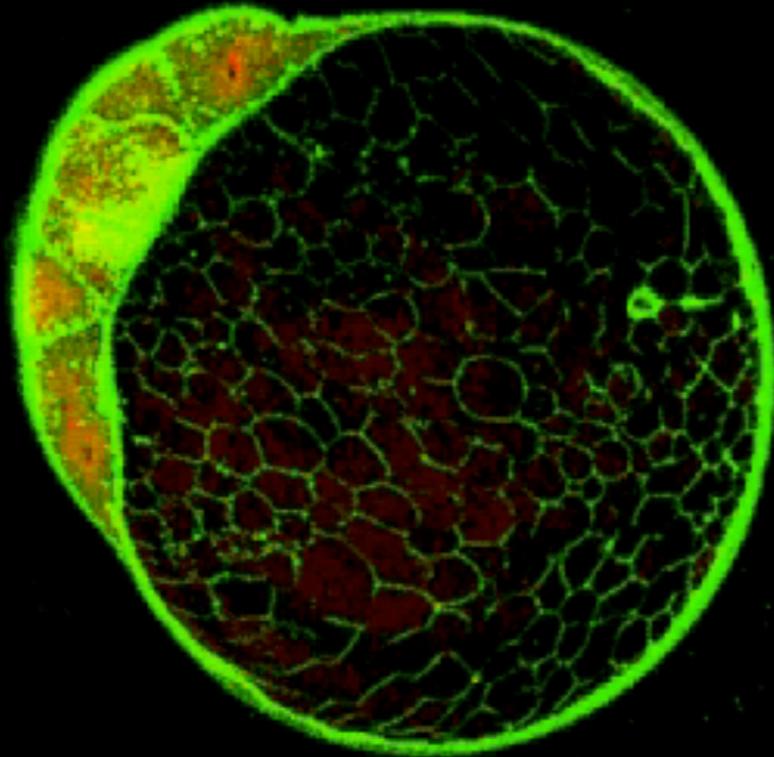
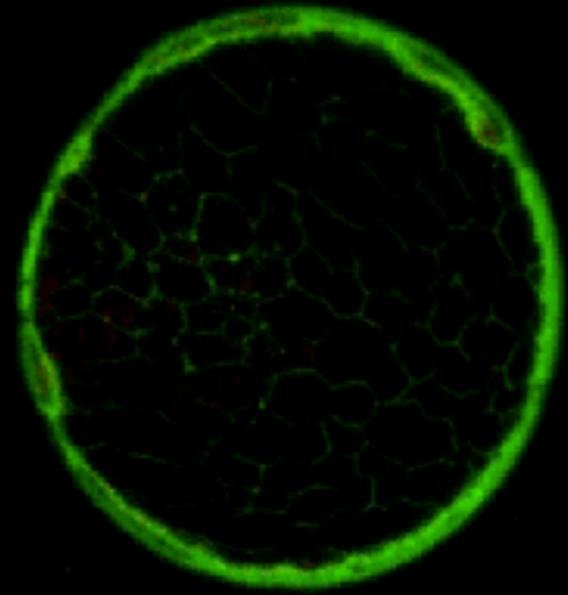
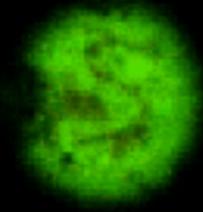


# Zebrafish



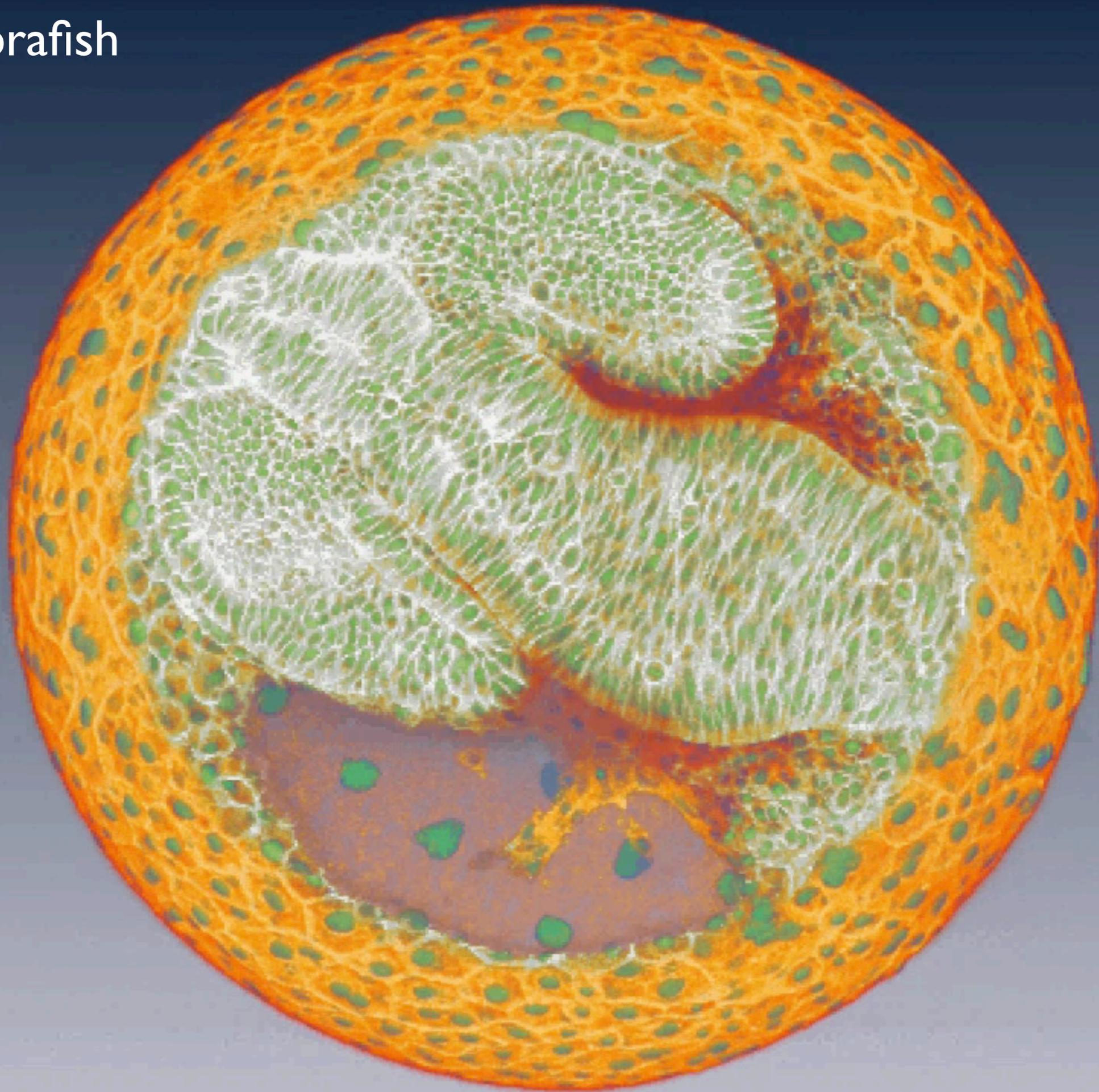


# Zebrafish



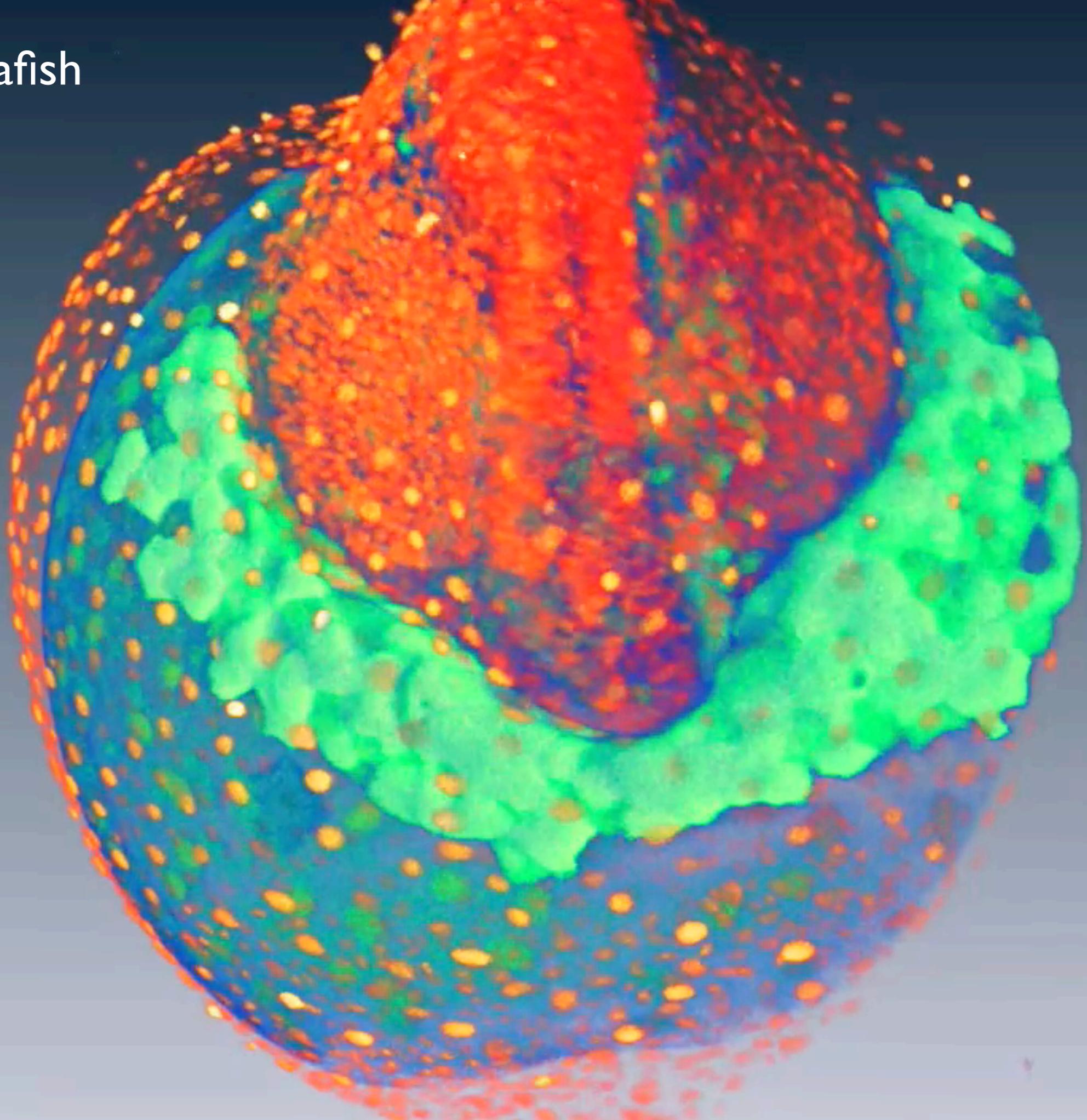


# Zebrafish



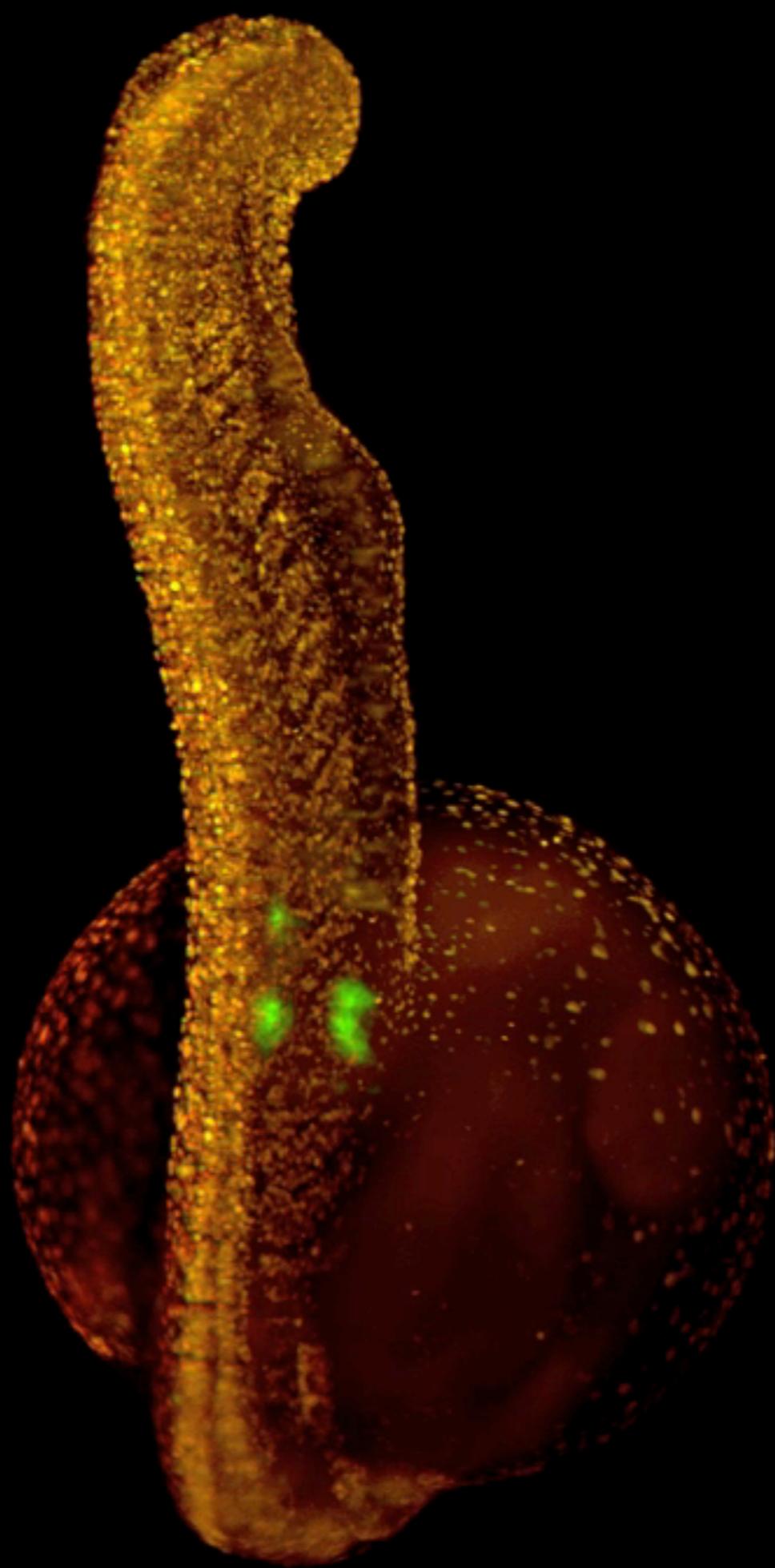


# Zebrafish





# Zebrafish



## Taille Image 4D

### Acquisition standard:

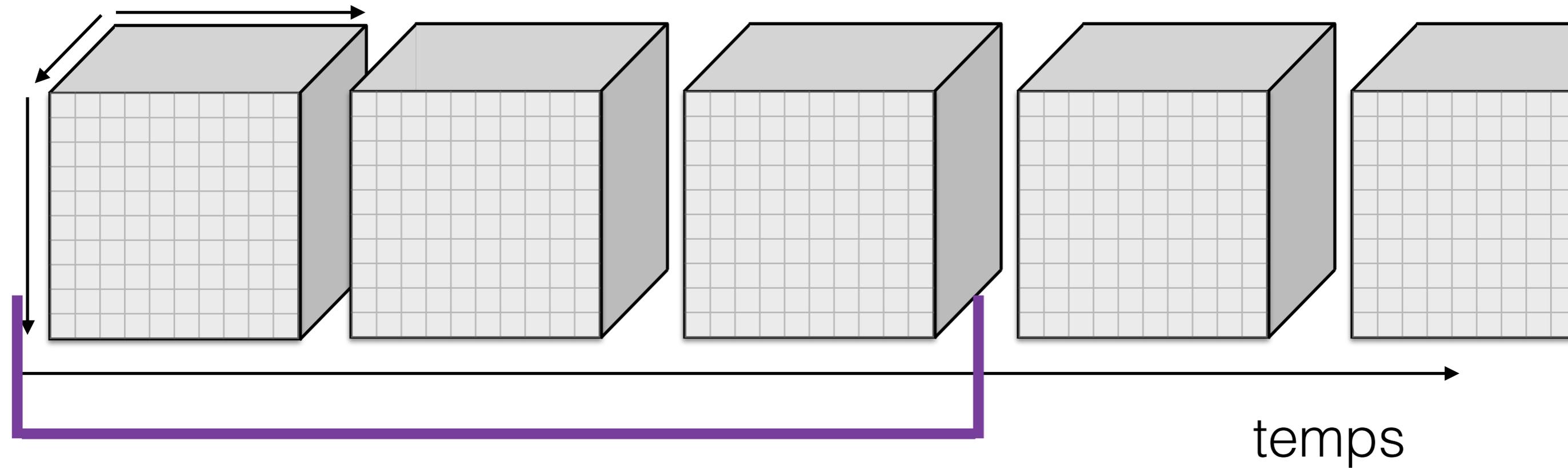
$X * Y * Z * \text{Temps} * \text{Canaux} * \text{Angle} : 1000 * 1000 * 200 * 1000 * 2 * 4$

→ + de mille milliards de voxels / embryons

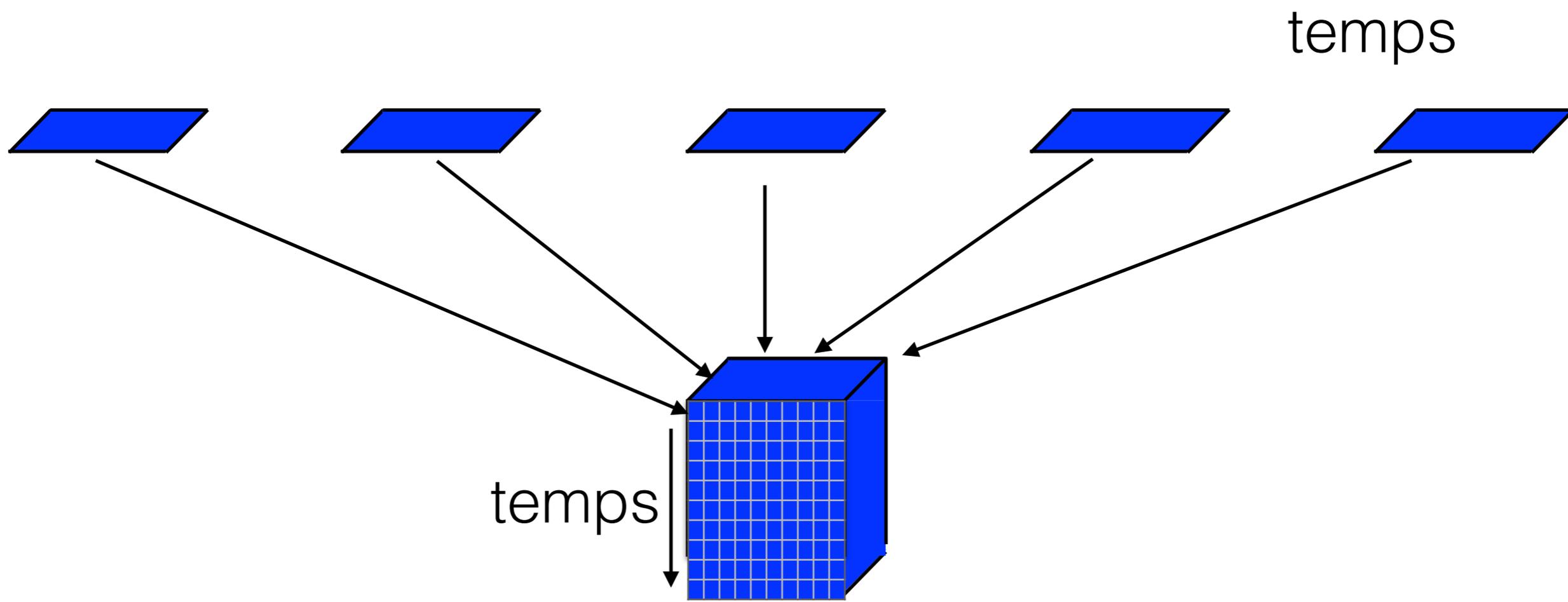
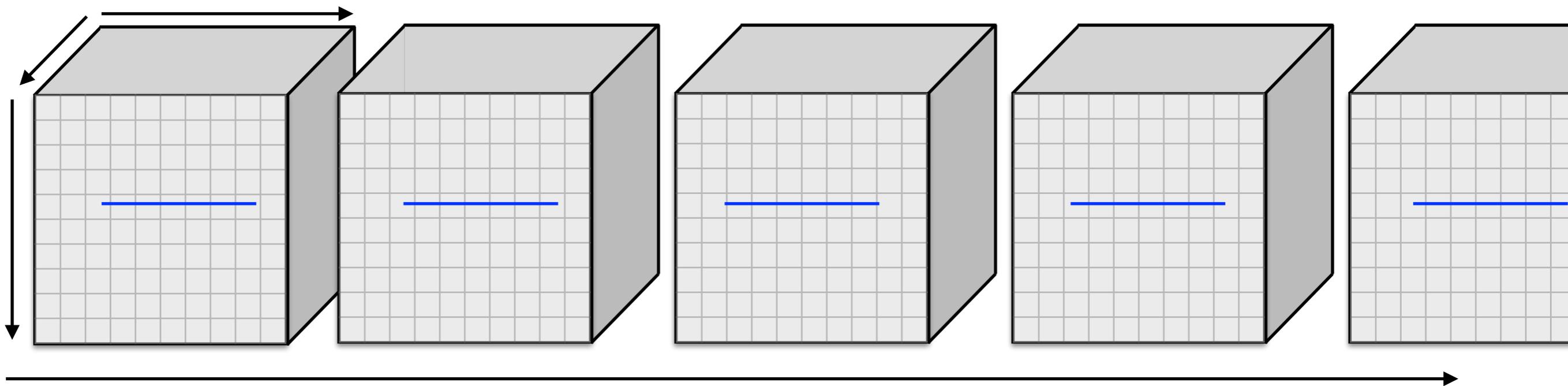
→ plusieurs terabytes / embryons

→ très bon taux de compression ~90%

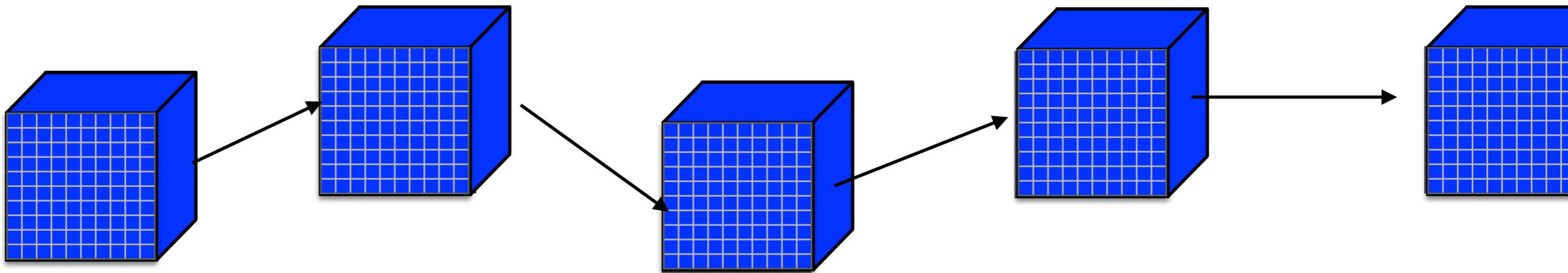
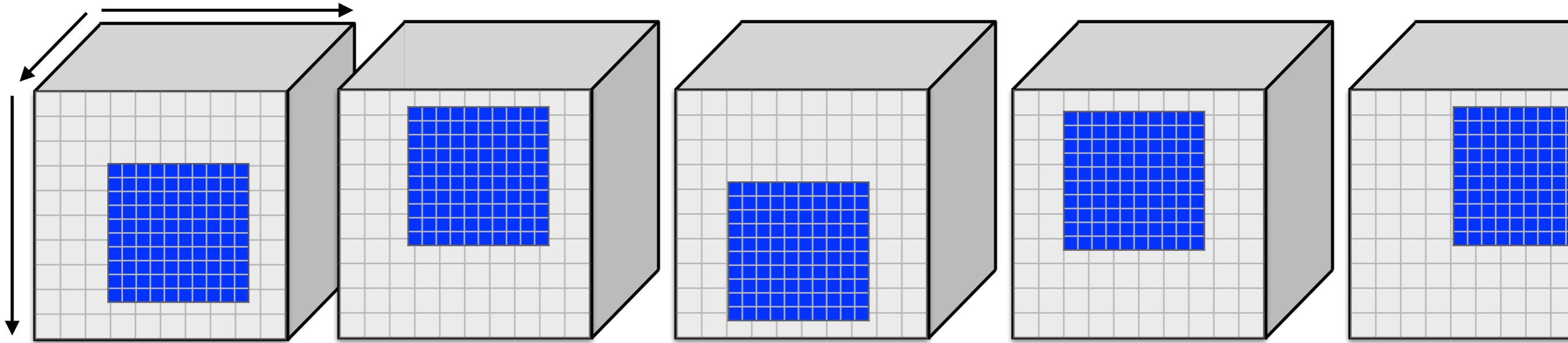
# Comment parcourir les images 4D ?



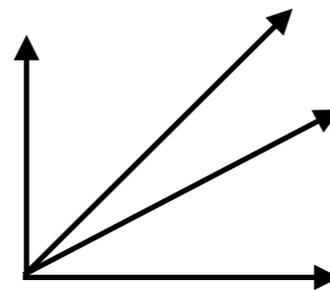
# Comment parcourir les images 4D ?



# Comment parcourir les images 4D ?

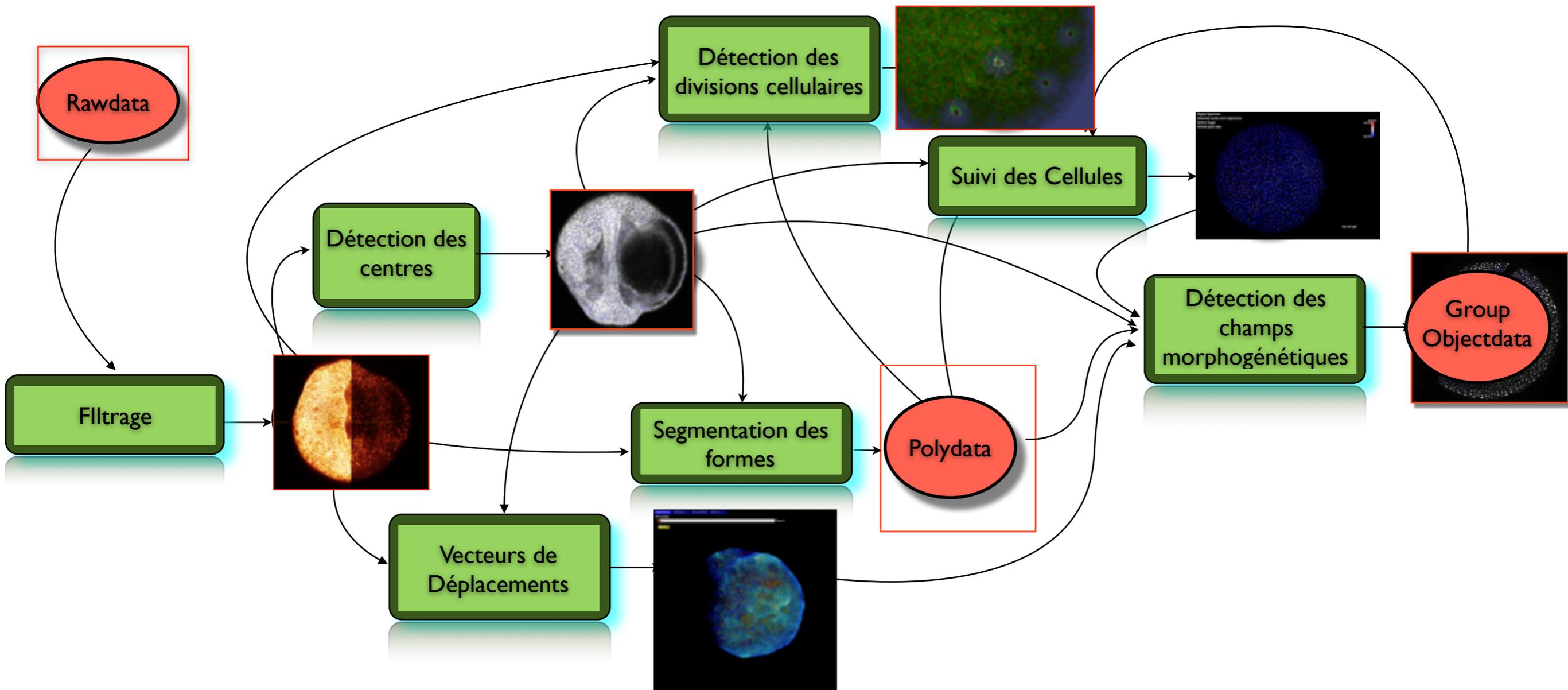


Espace 4D

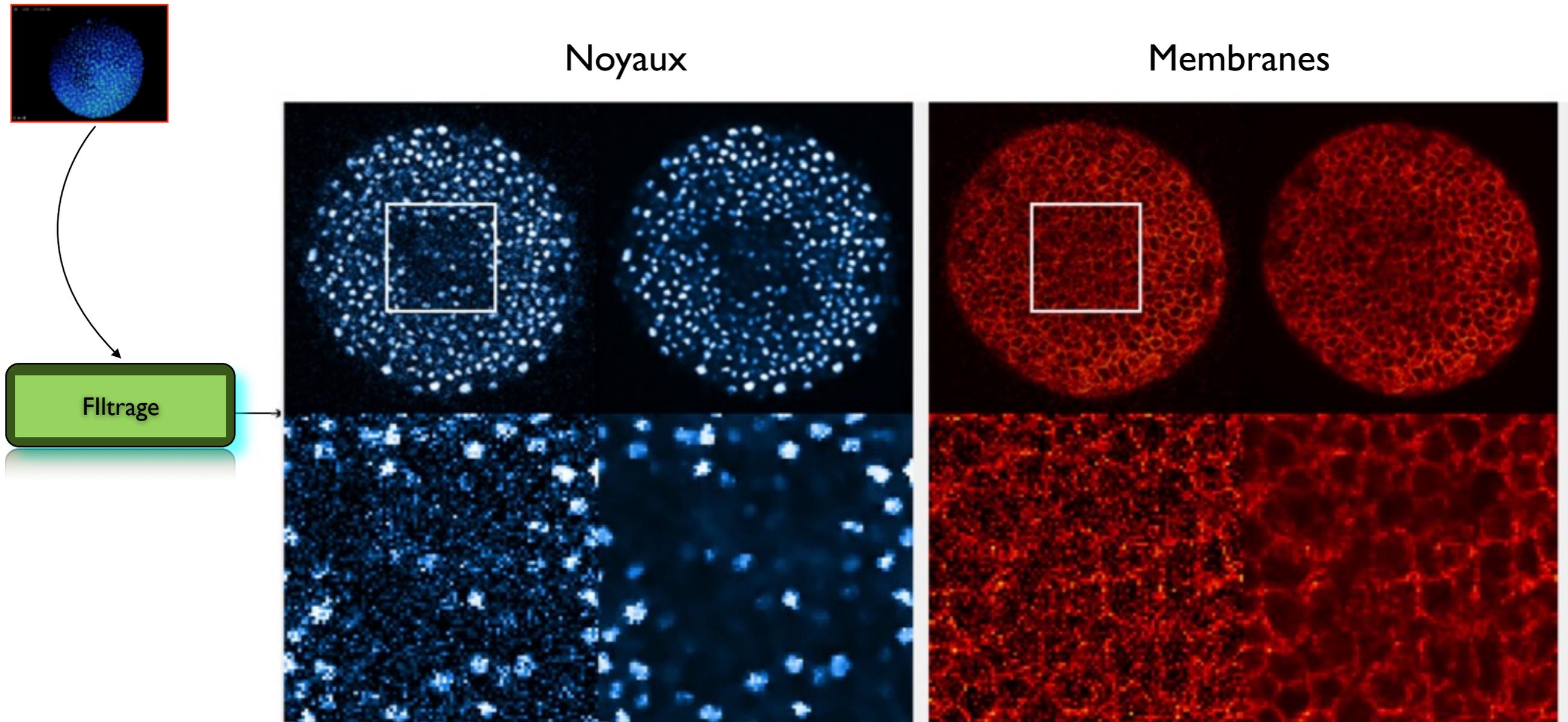


# Reconstruction d'images 4D

# Reconstruction Image 4D

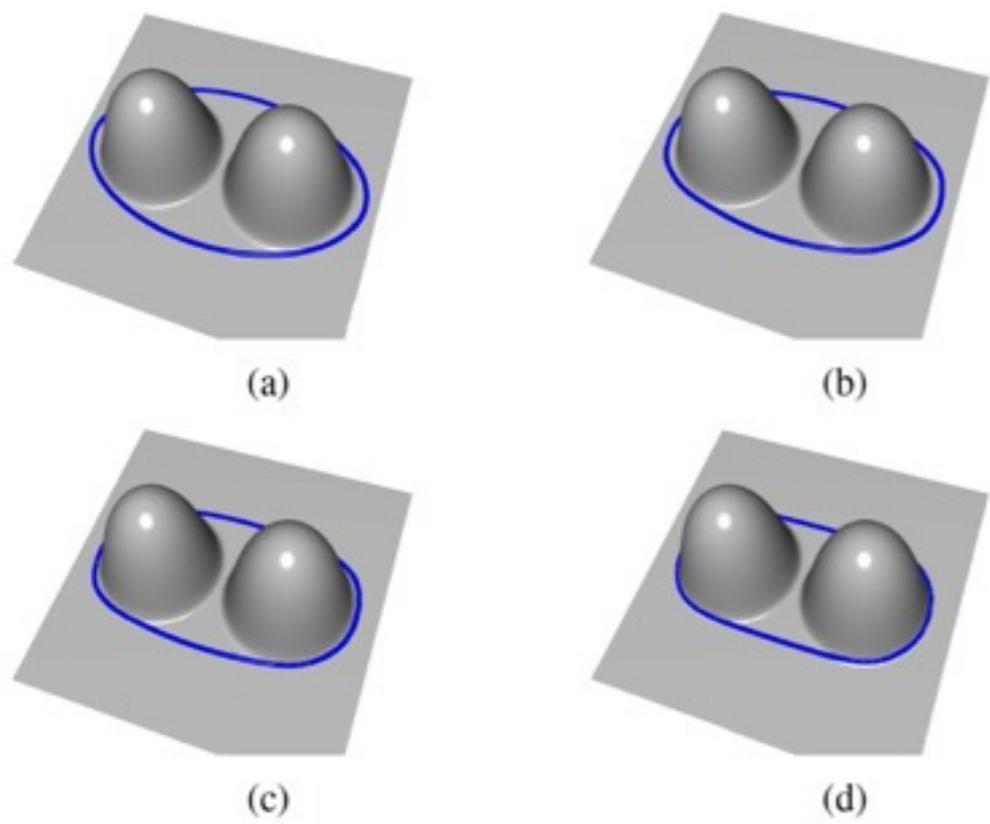
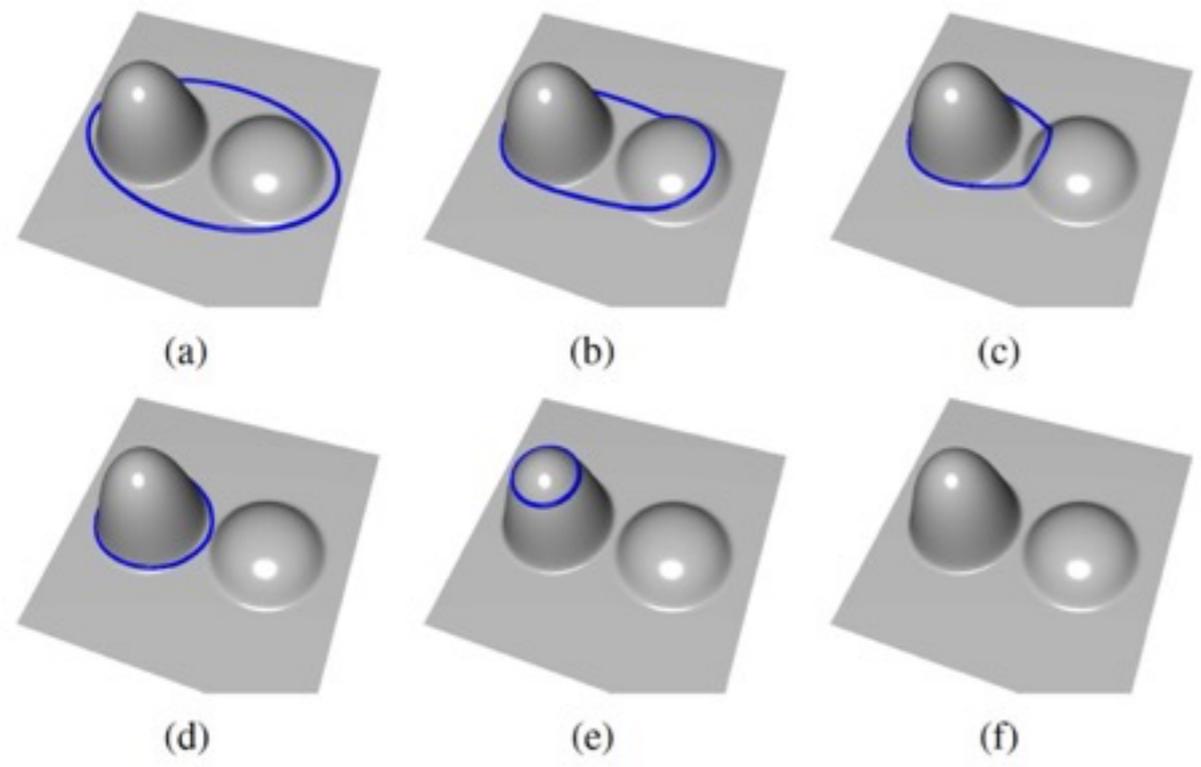


# Filtres non linéaire

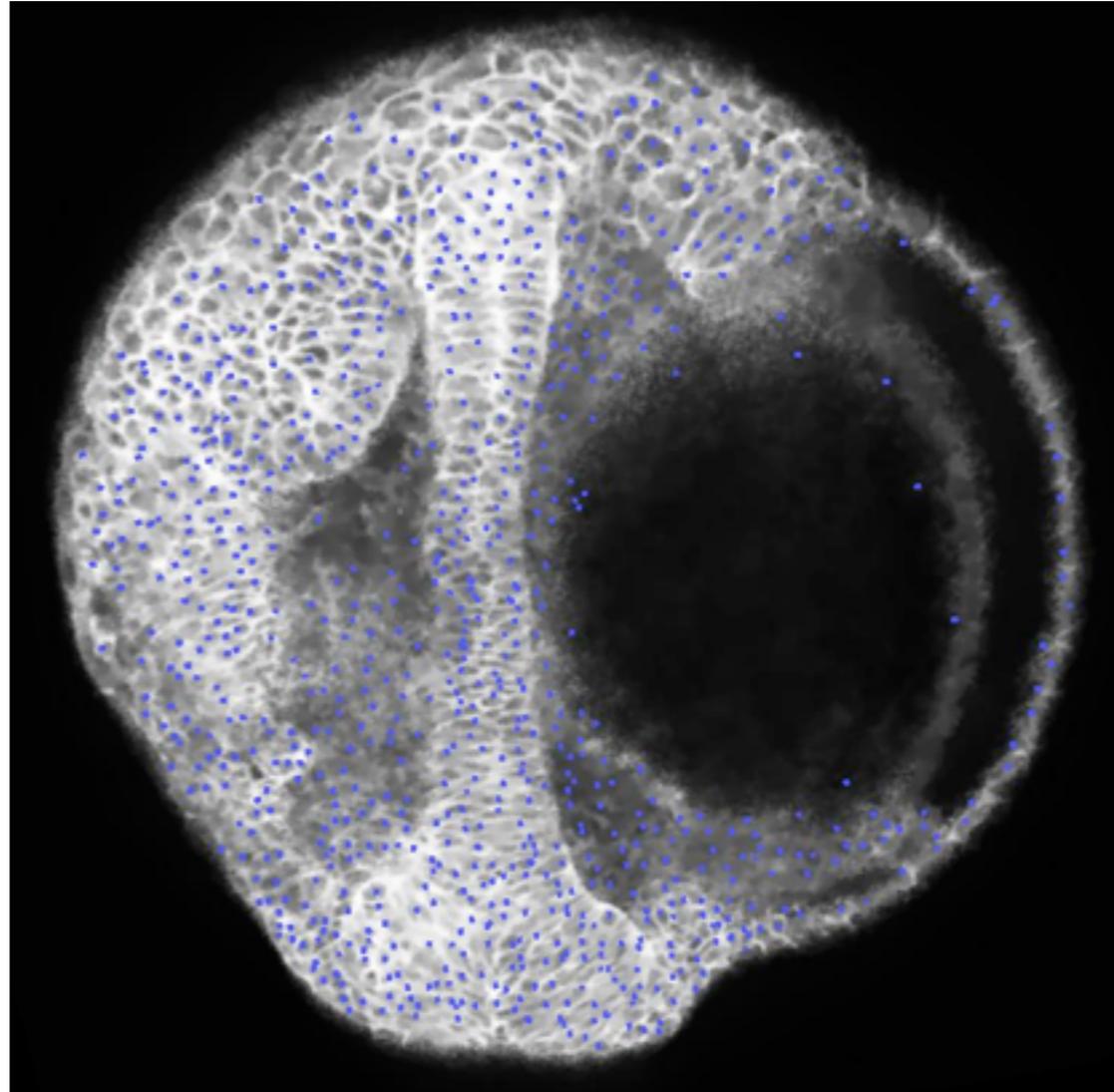
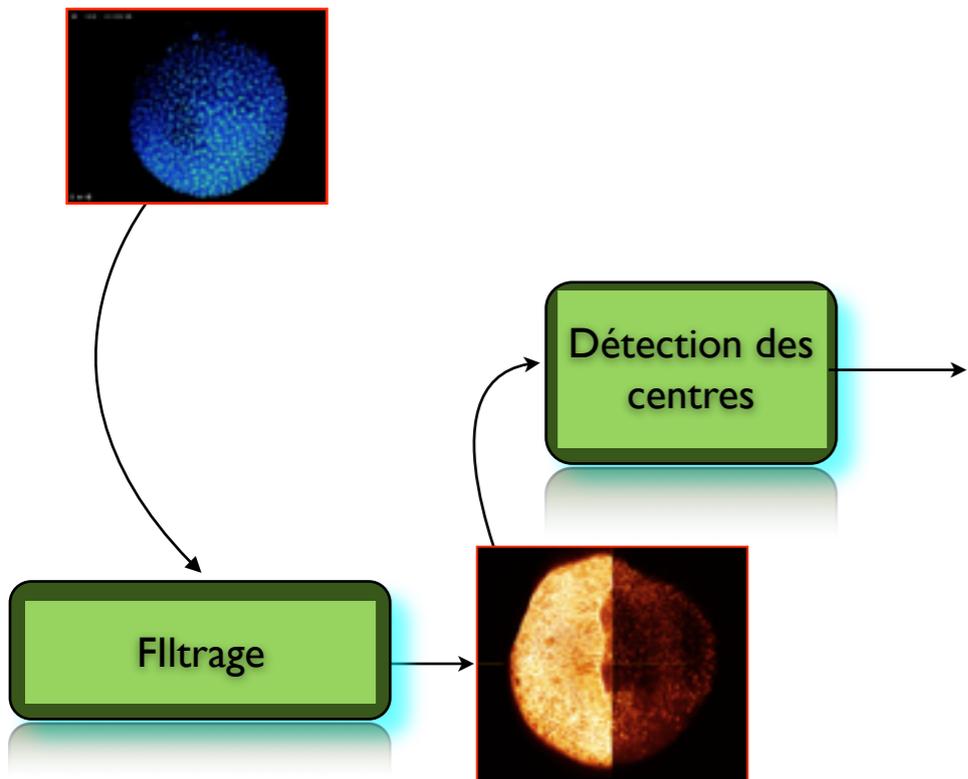


*Intégrité des contours* : on souhaiterait éliminer le bruit sans rendre flous les frontières des objets.

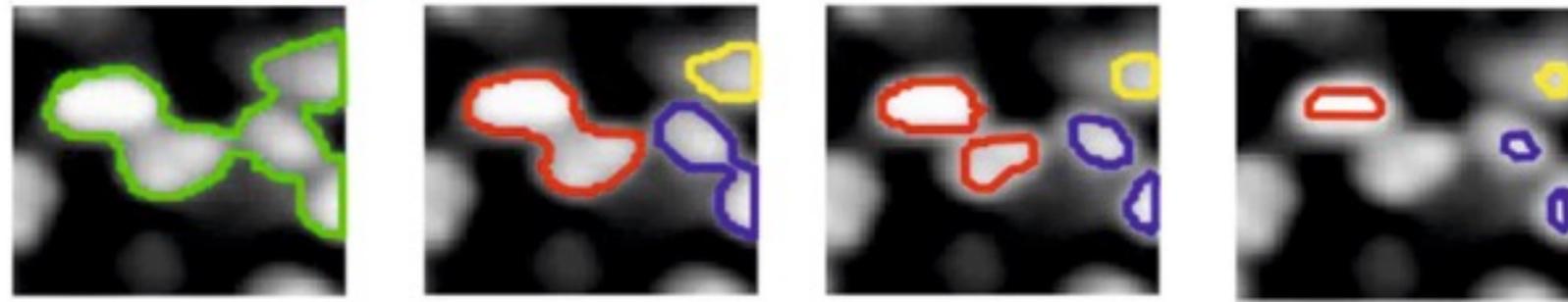
# Geodesic Mean curvature Flow



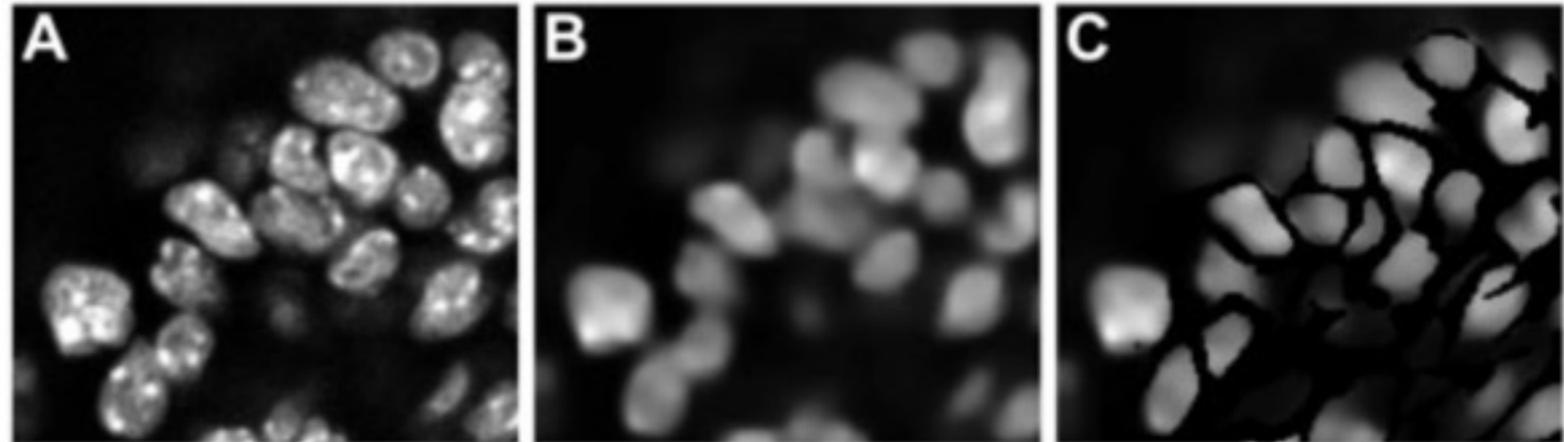
# Identification des cellules



# Morphologie Mathématique

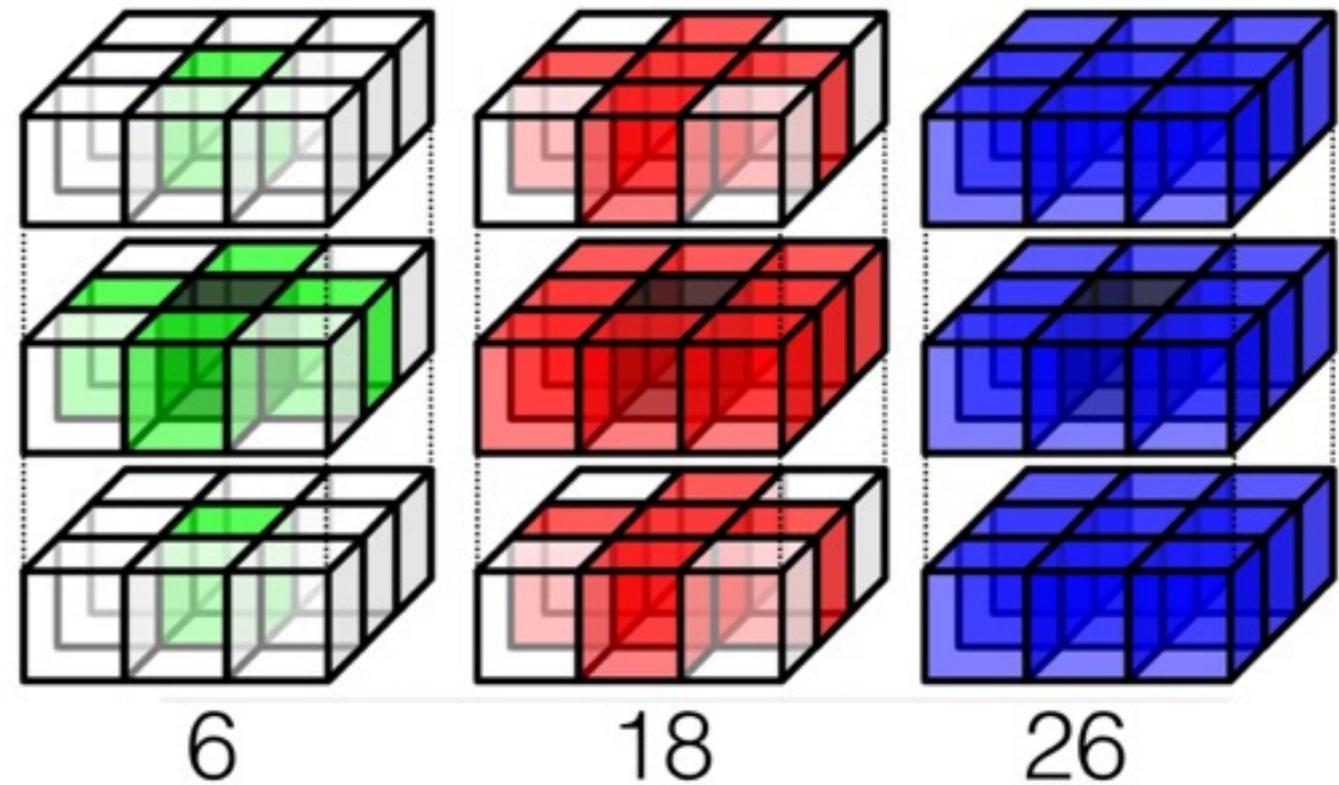


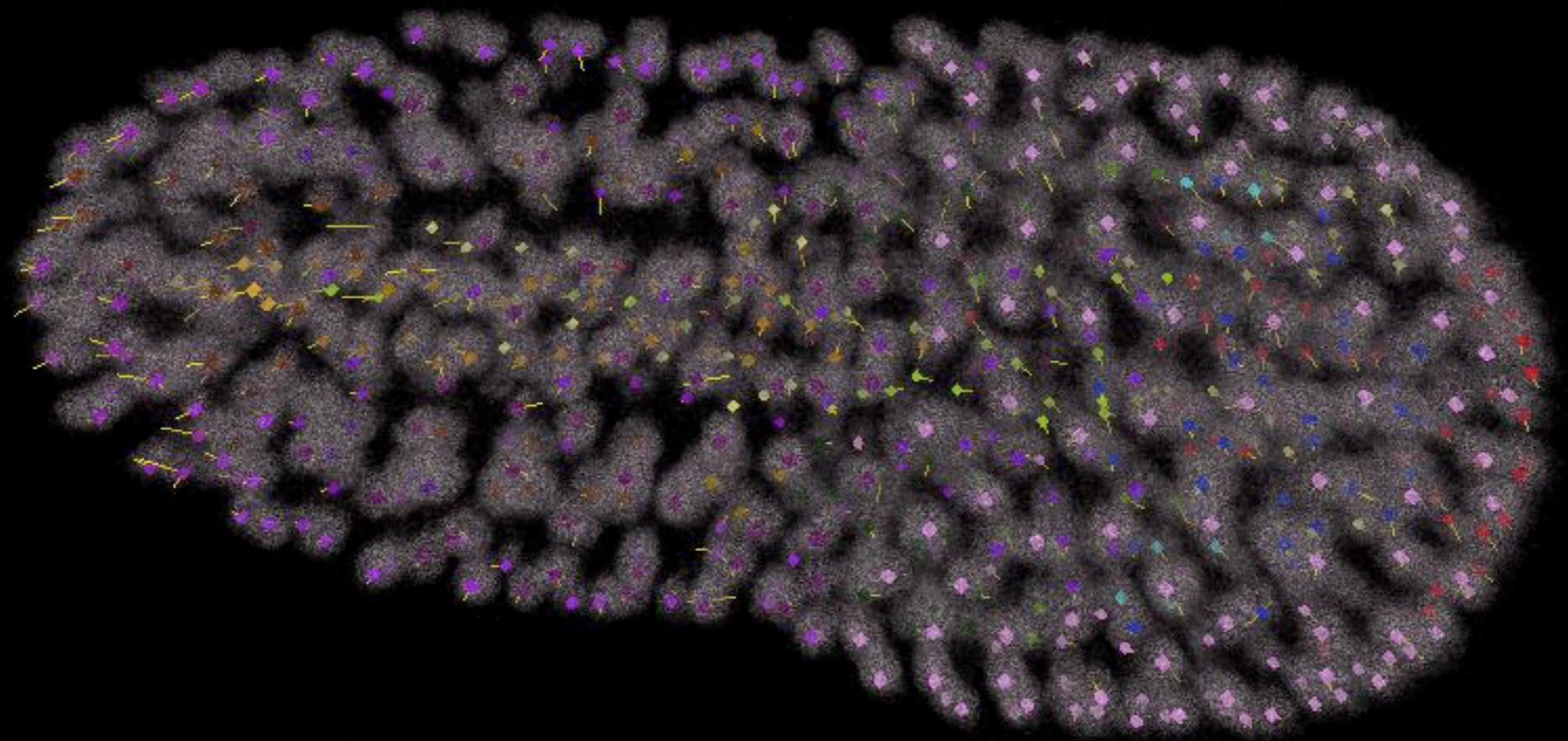
## Soustraction de l'image des membranes



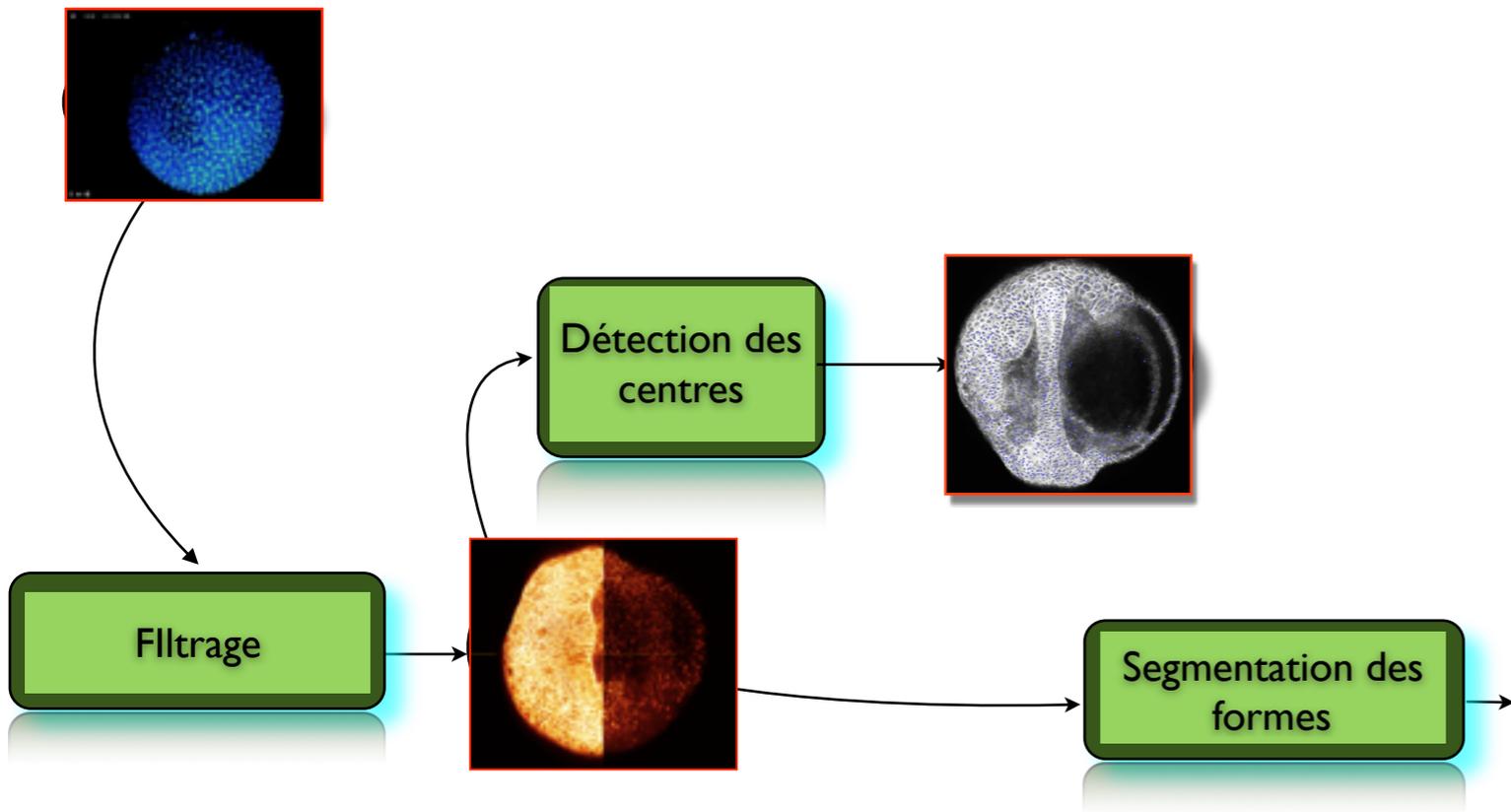
Pop 2013

## Labels par composantes connexes





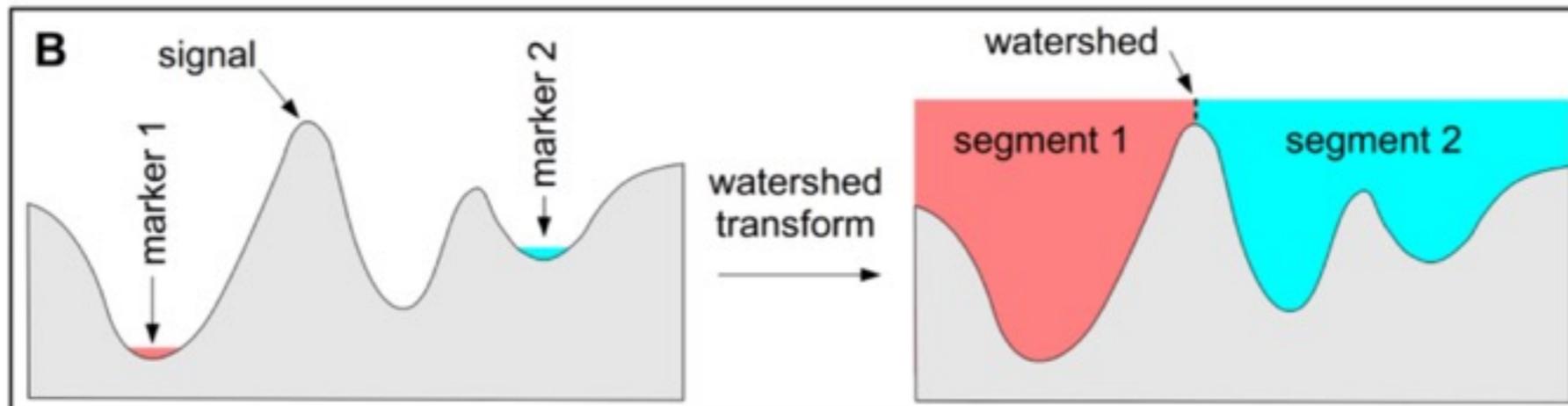
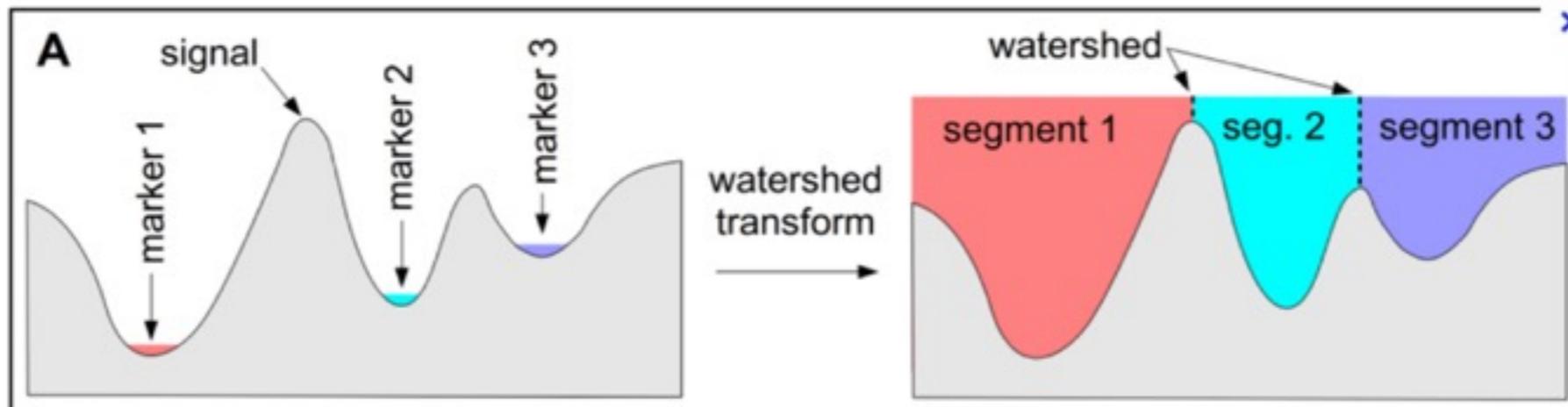
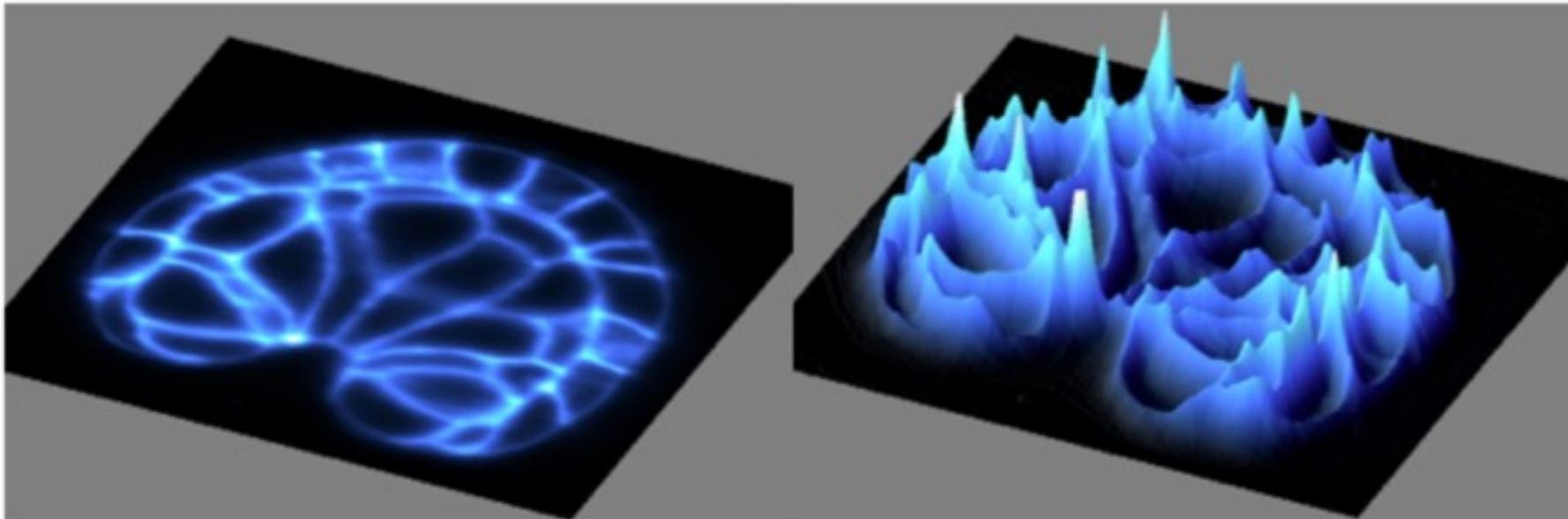
# Segmentation 3D



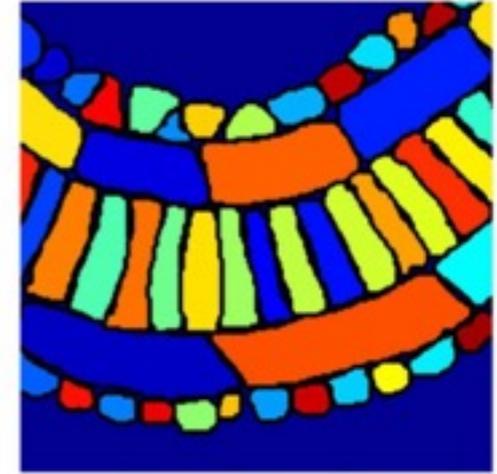
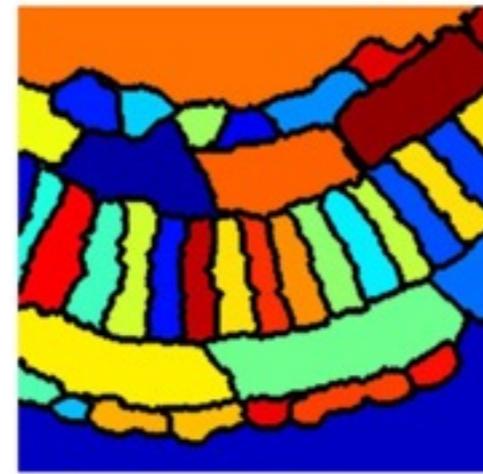
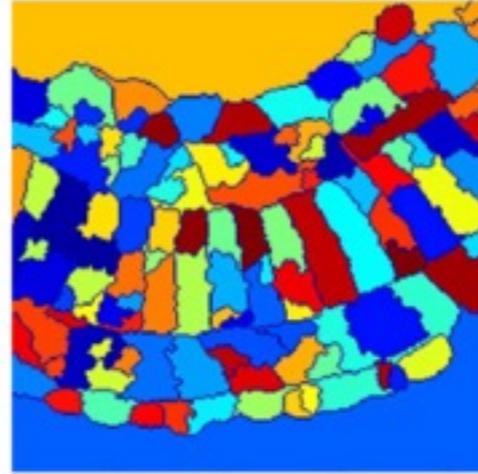
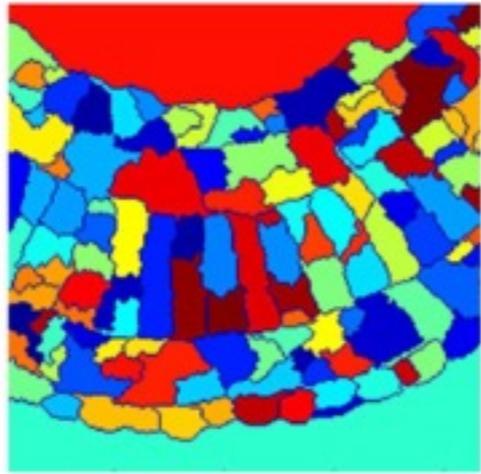
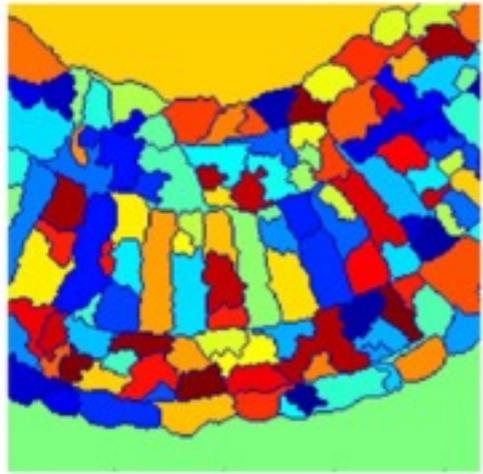
# Watershed

Intensity image

Intensity image as a landscape



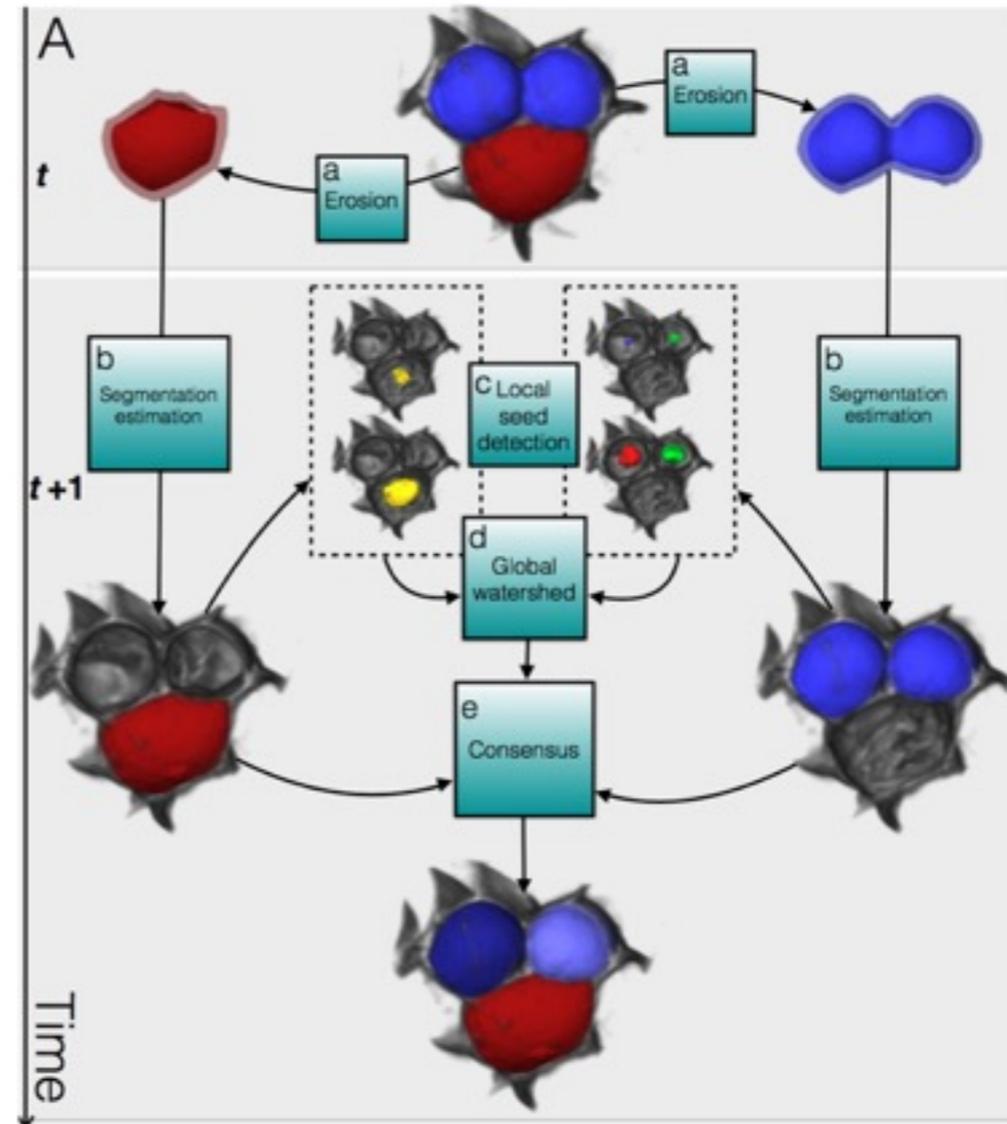
# Watershed



Random seed

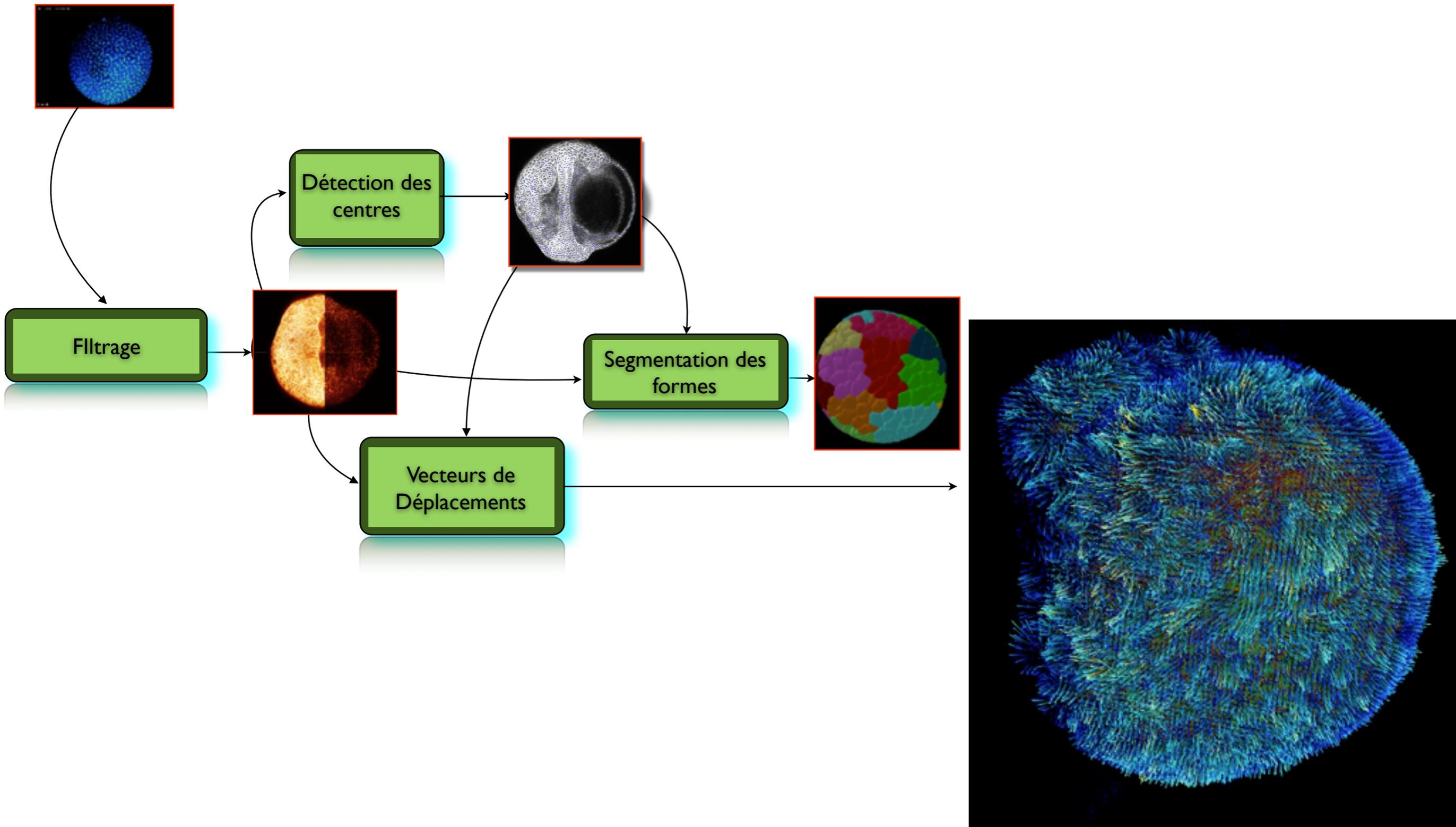
*Delibaltov 2013*

## Watershed 4D



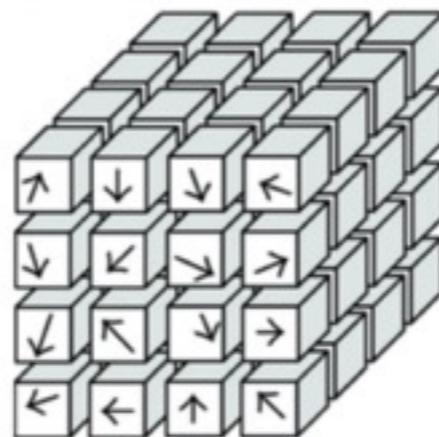
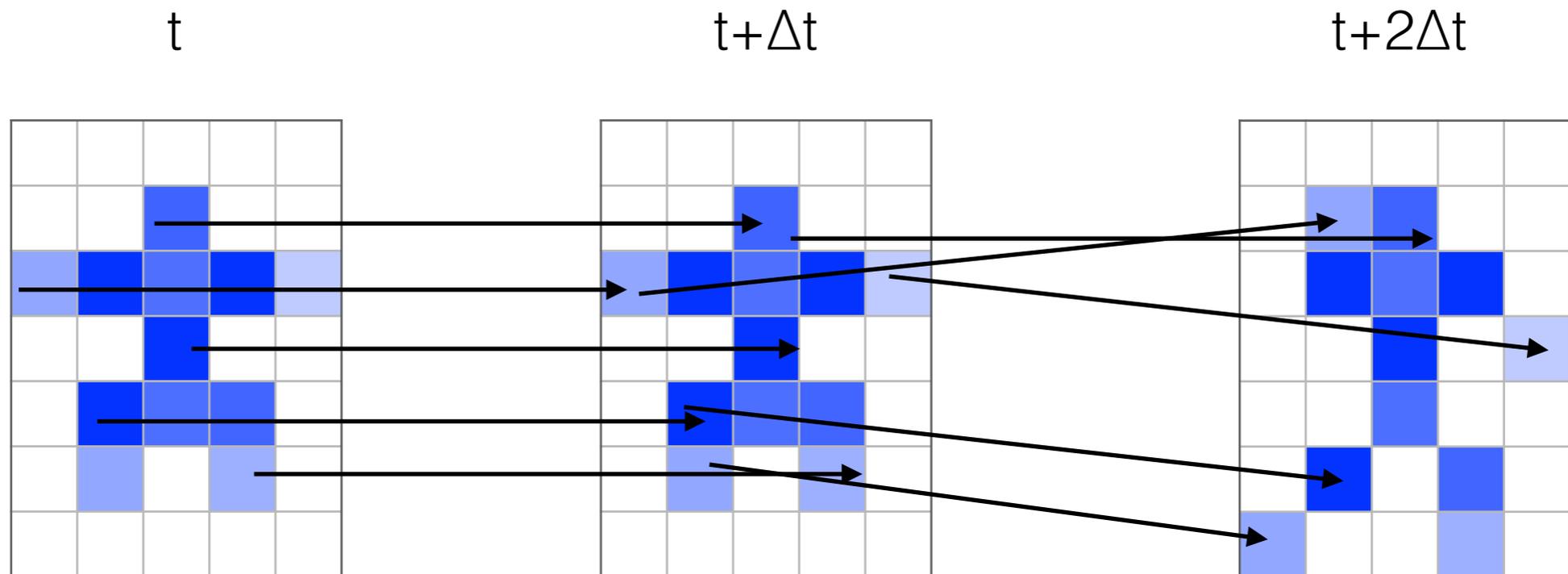
*Guignard 2015*

# Reconstruction Image 4D

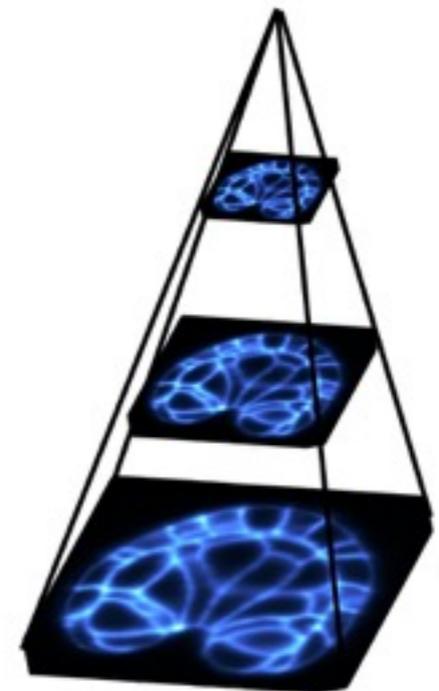


# Vecteurs de déplacements (Flot optique)

## Recalage non rigide



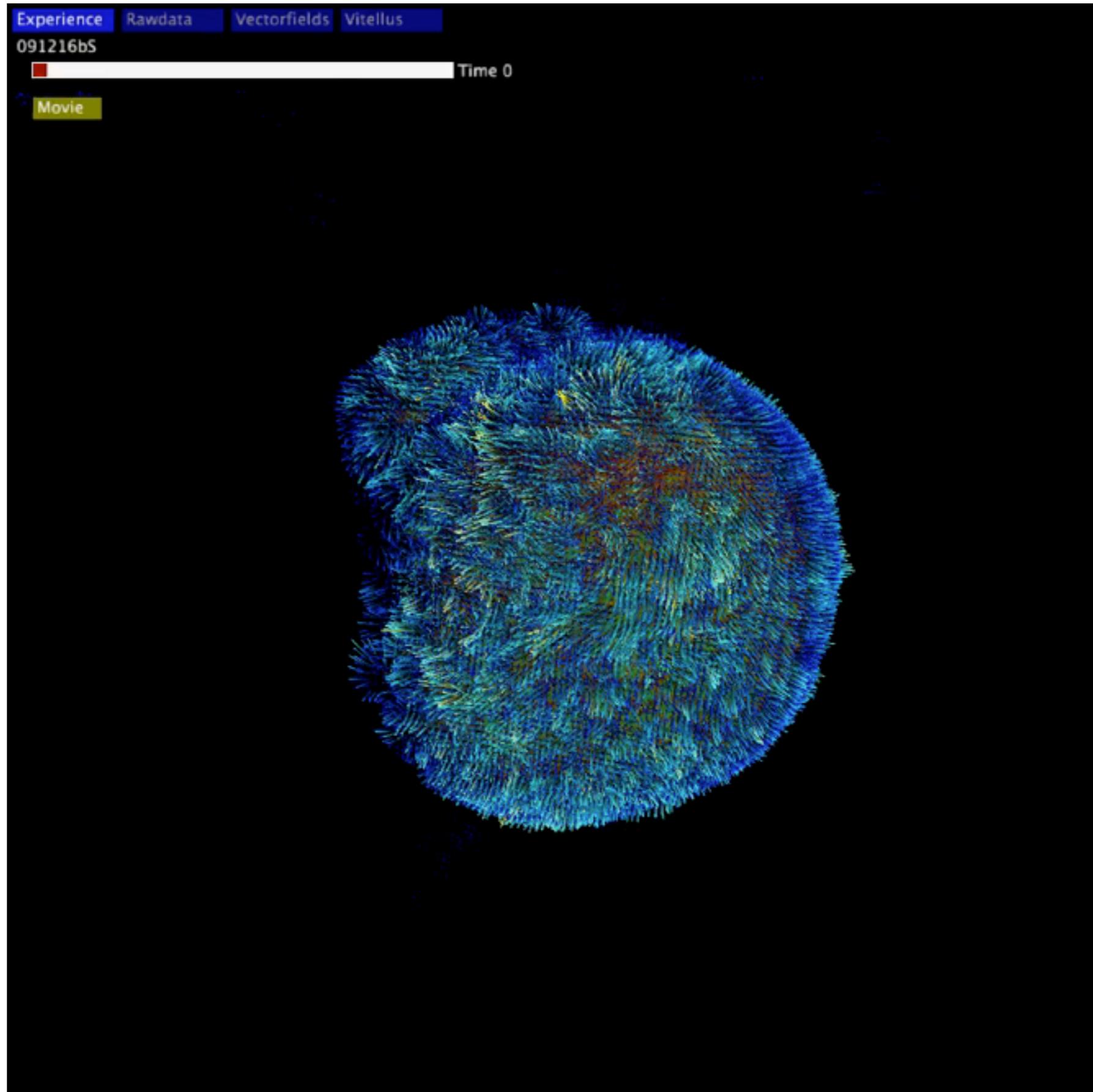
4D

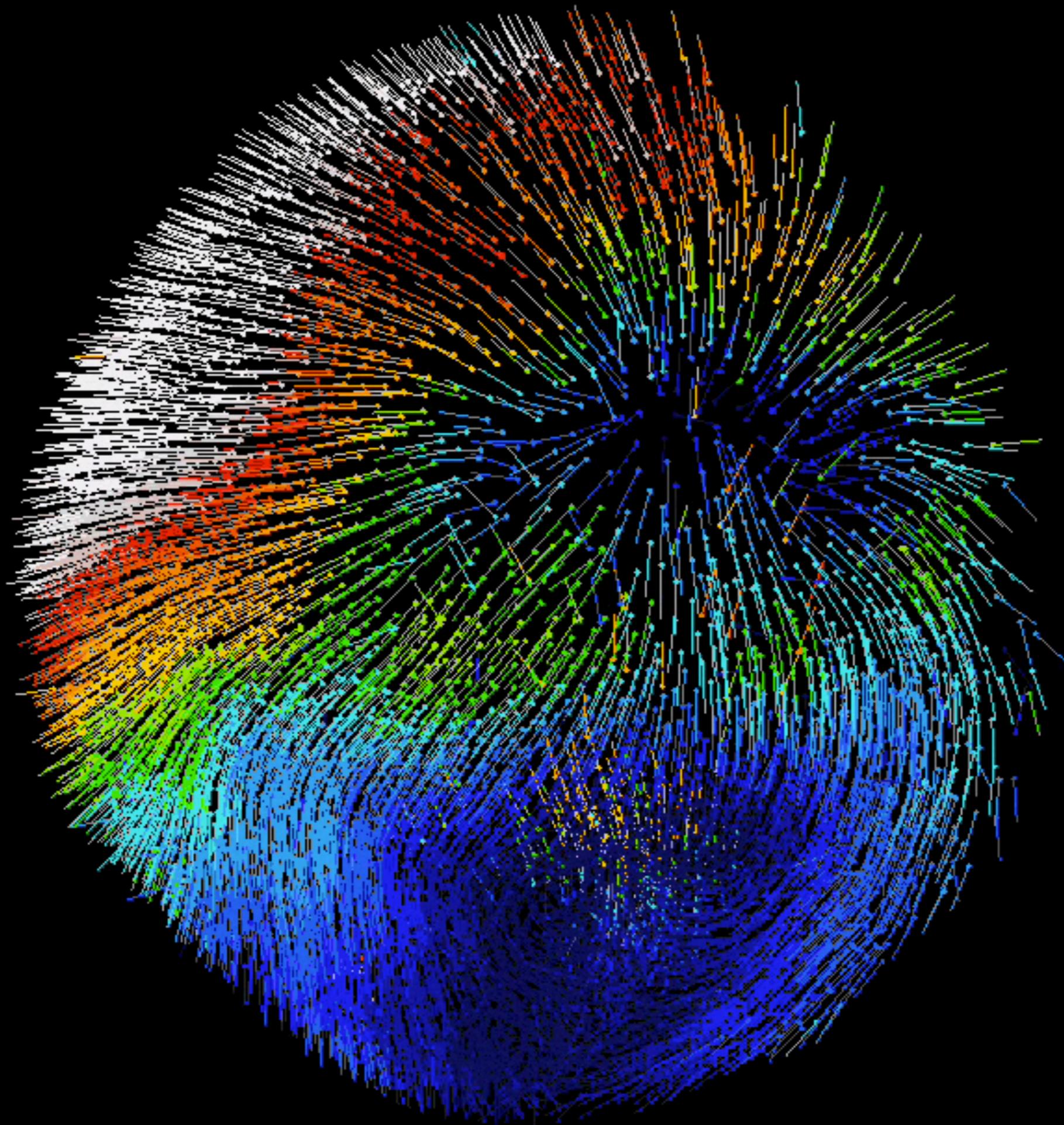


*Hypothèse* : invariance de l'intensité de chaque voxel entre 2 images

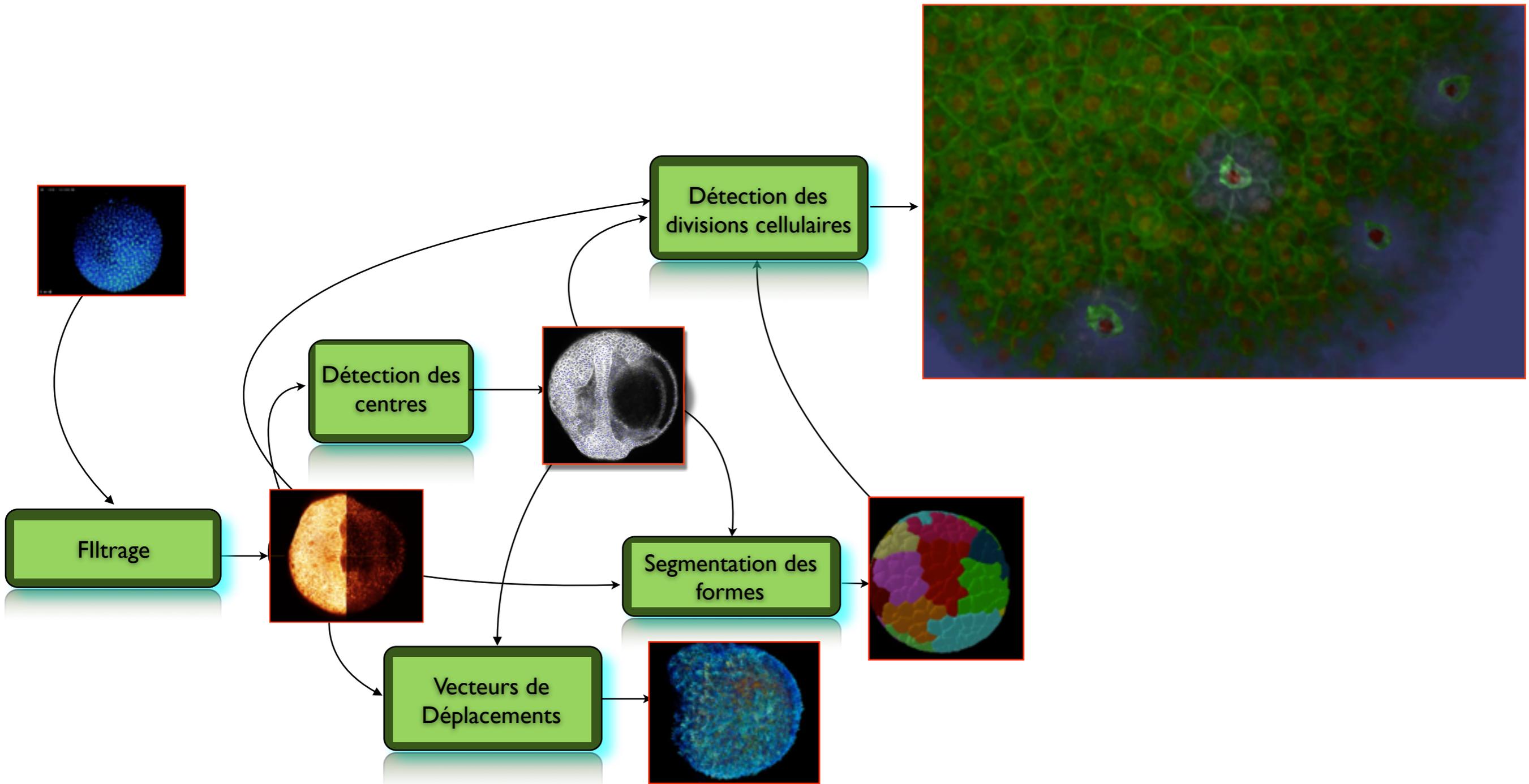
Approche multi-résolution

# Moyenne spatiale et temporel

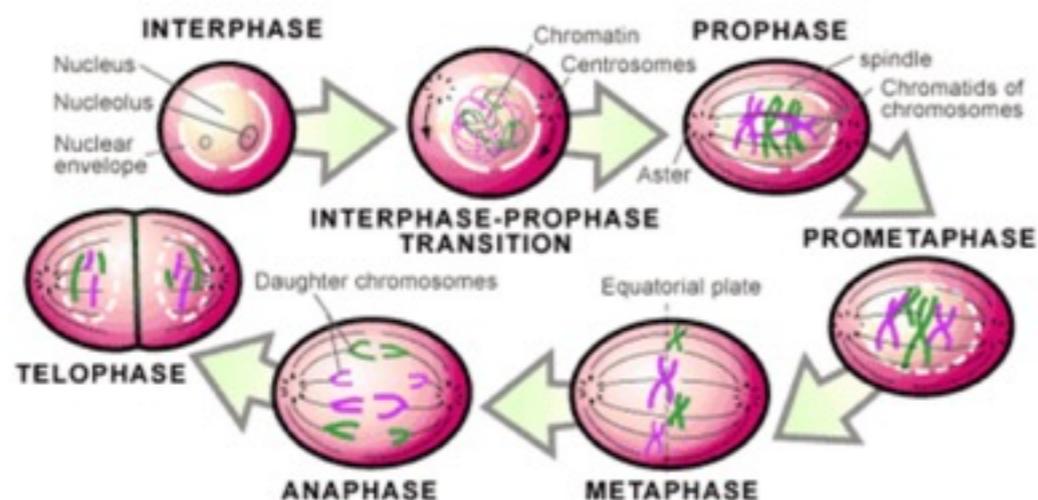
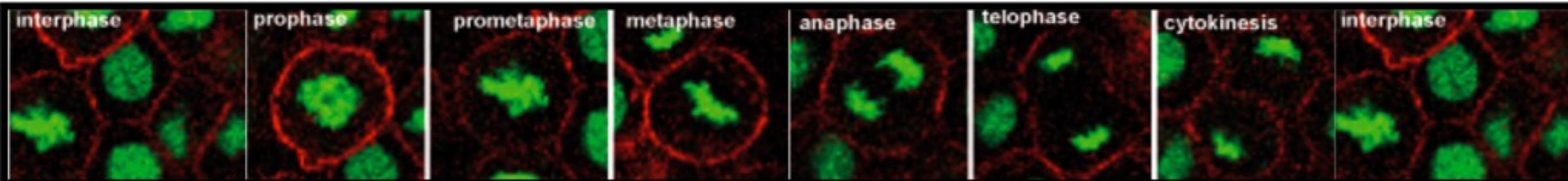




# Reconstruction Image 4D

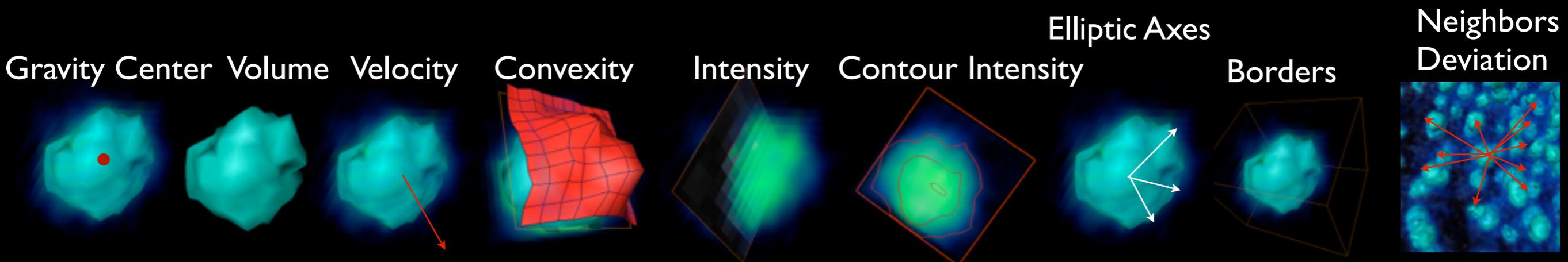


# Détection des divisions cellulaires

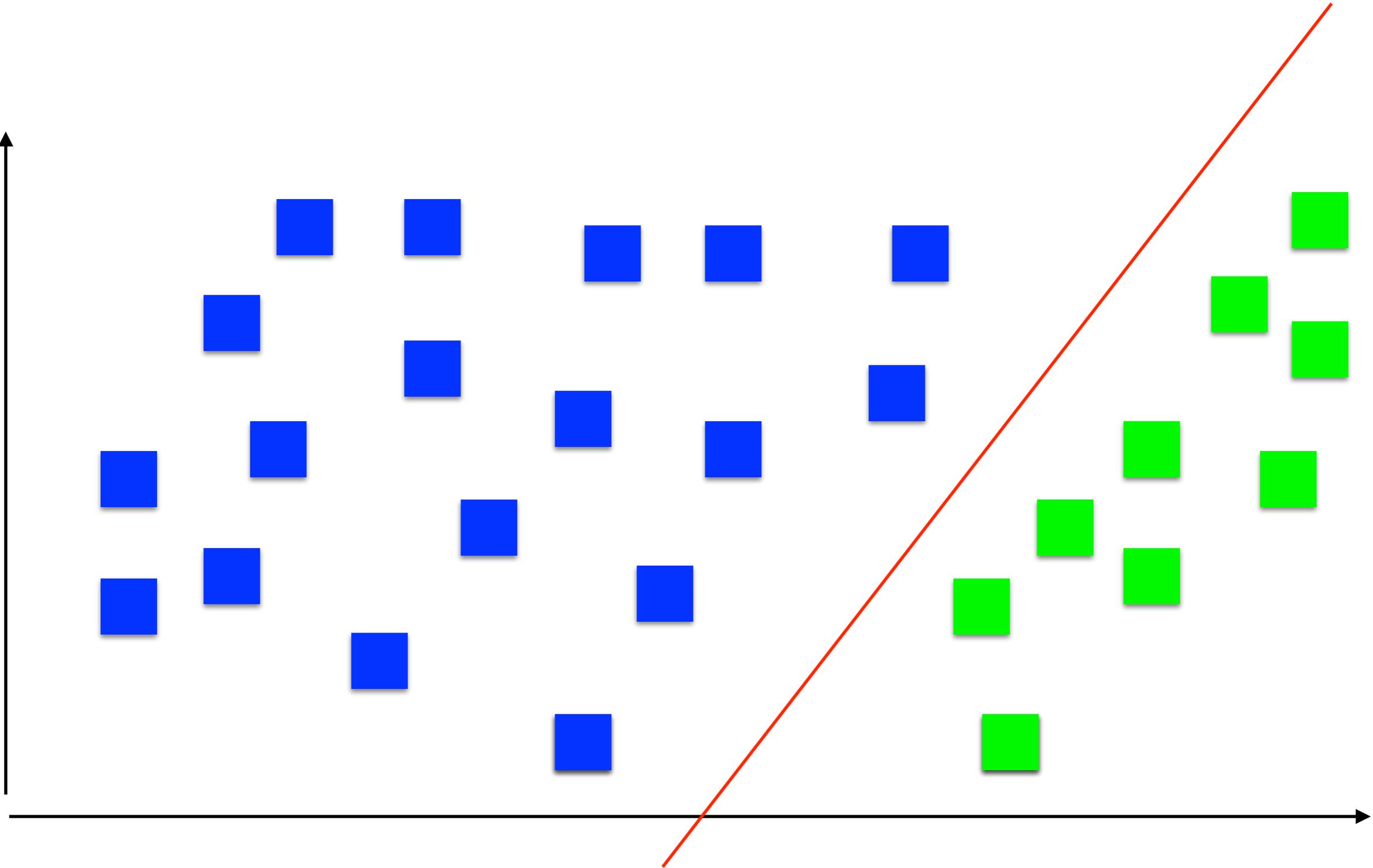


- **Prophase** : Membrane : shape + convex
- **Prometaphase** : Nucleus : volume down
- **Metaphase** : Nucleus : higher intensity
- **Anaphase** : 2 closest nuclei , 1 Membrane.
- **Telophase** : Membrane : shape change
- **Cytokinesis** : 2 Membranes

+ Interphase → 7 classes

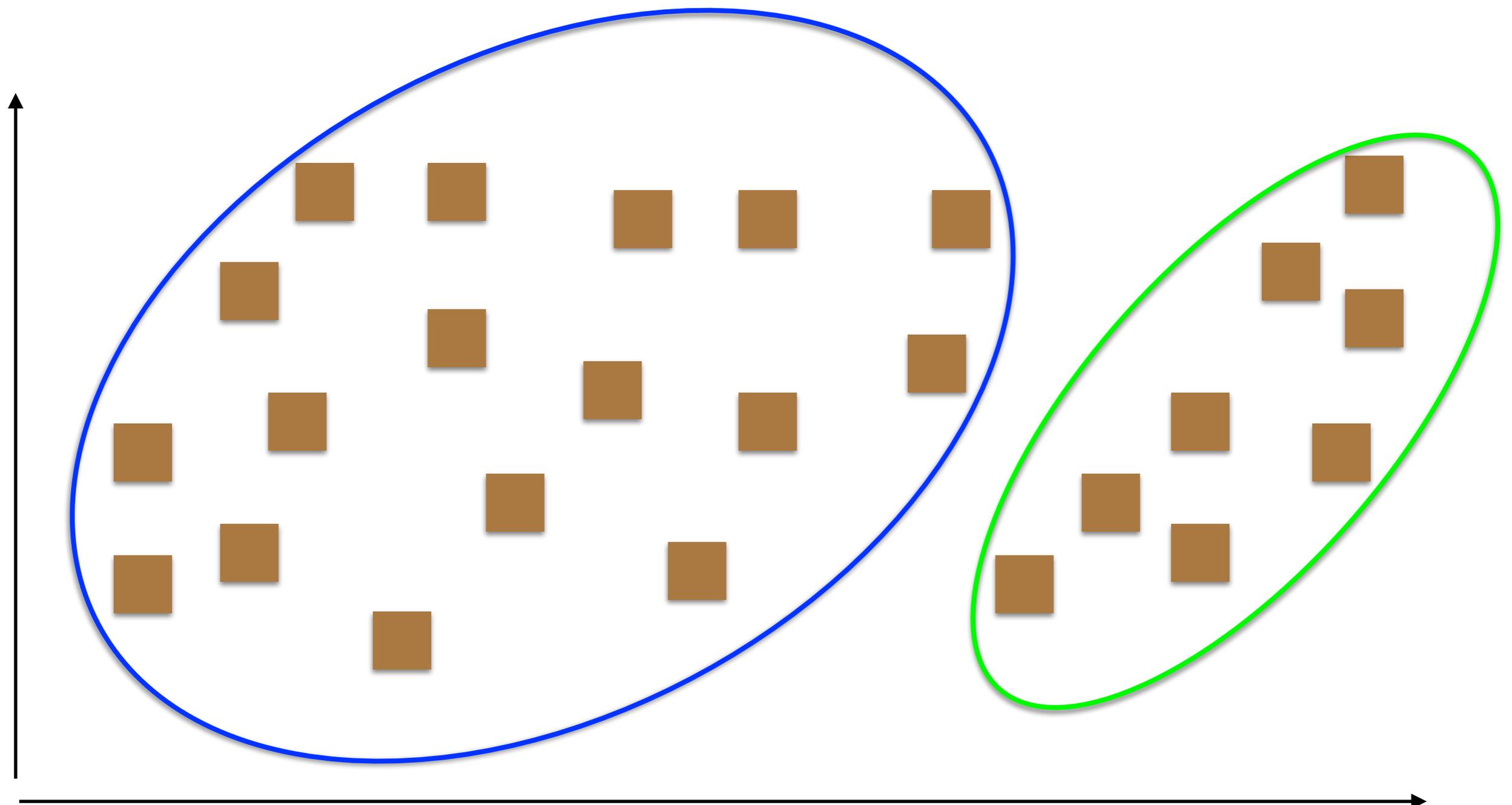


Très courte introduction à la classification



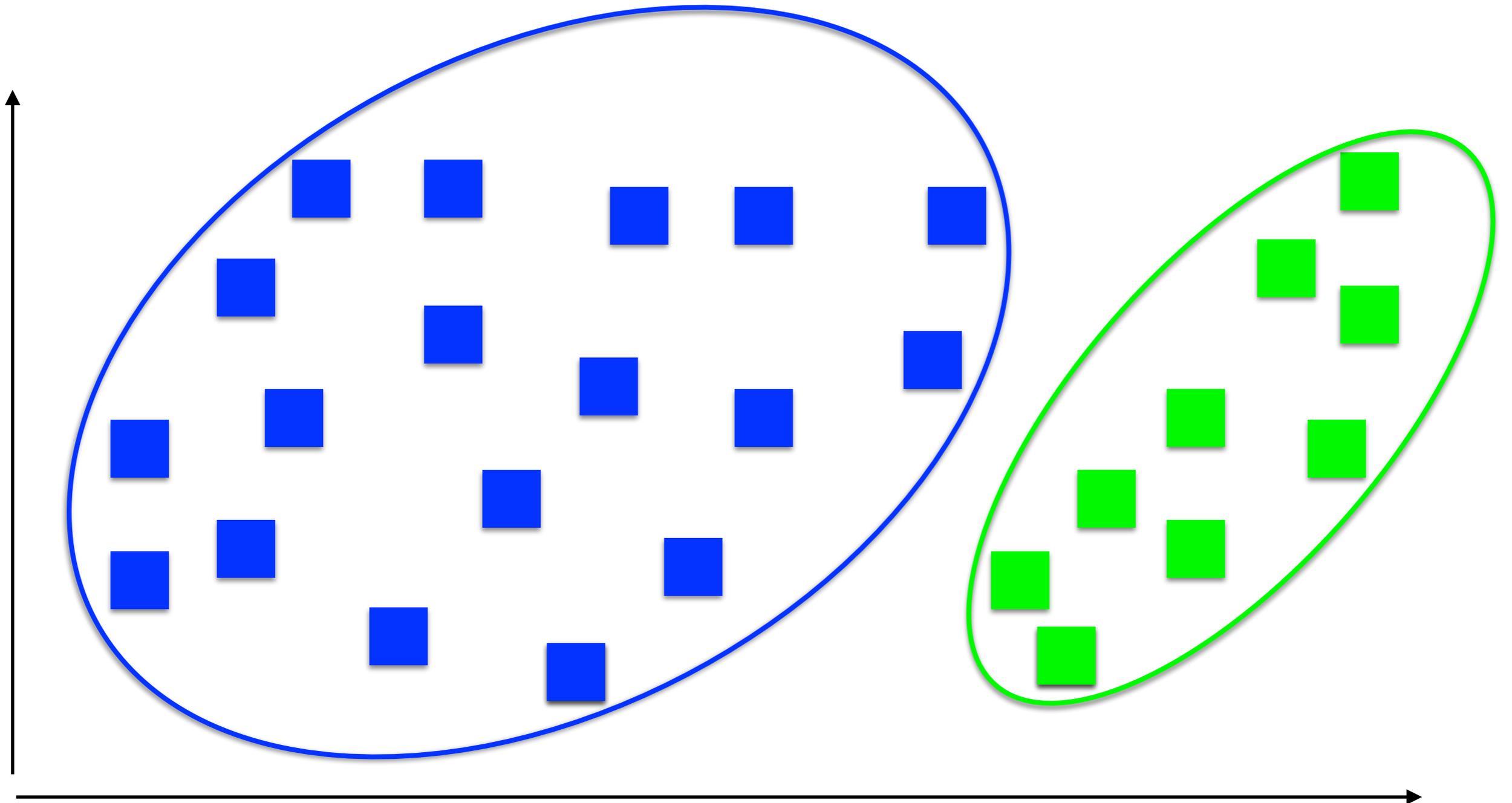
Classification supervisée

Très courte introduction à la classification



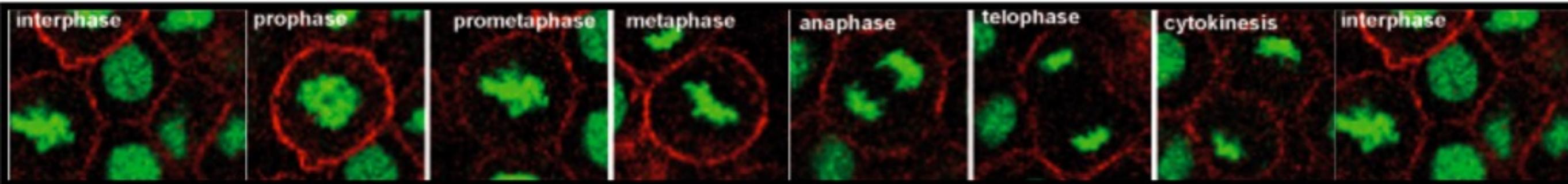
Classification non-supervisée

# Très courte introduction à la classification



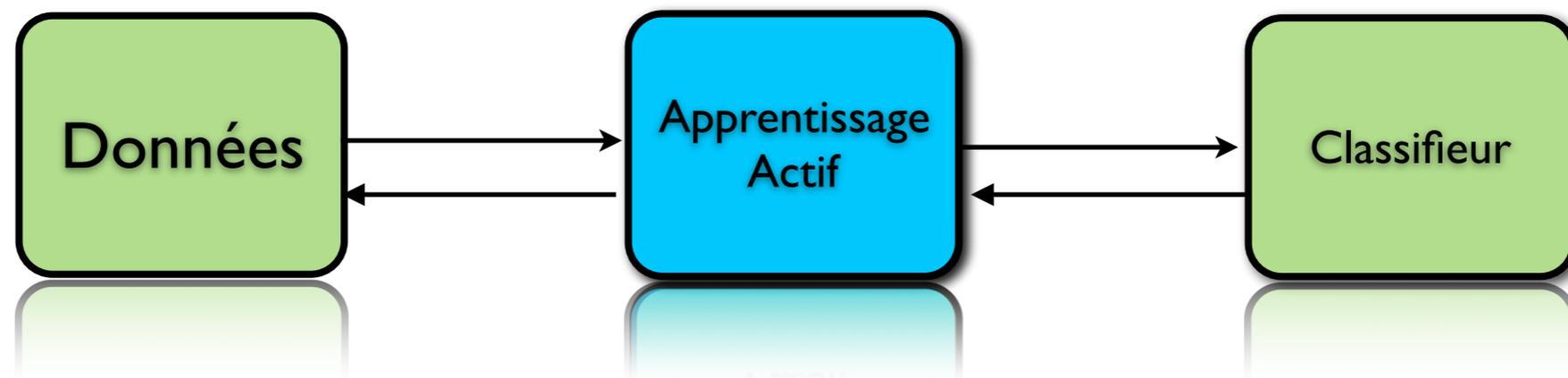
Classification non-supervisée

# Détection des divisions cellulaires



**Apprentissage supervisé**  
annotation experte très longue

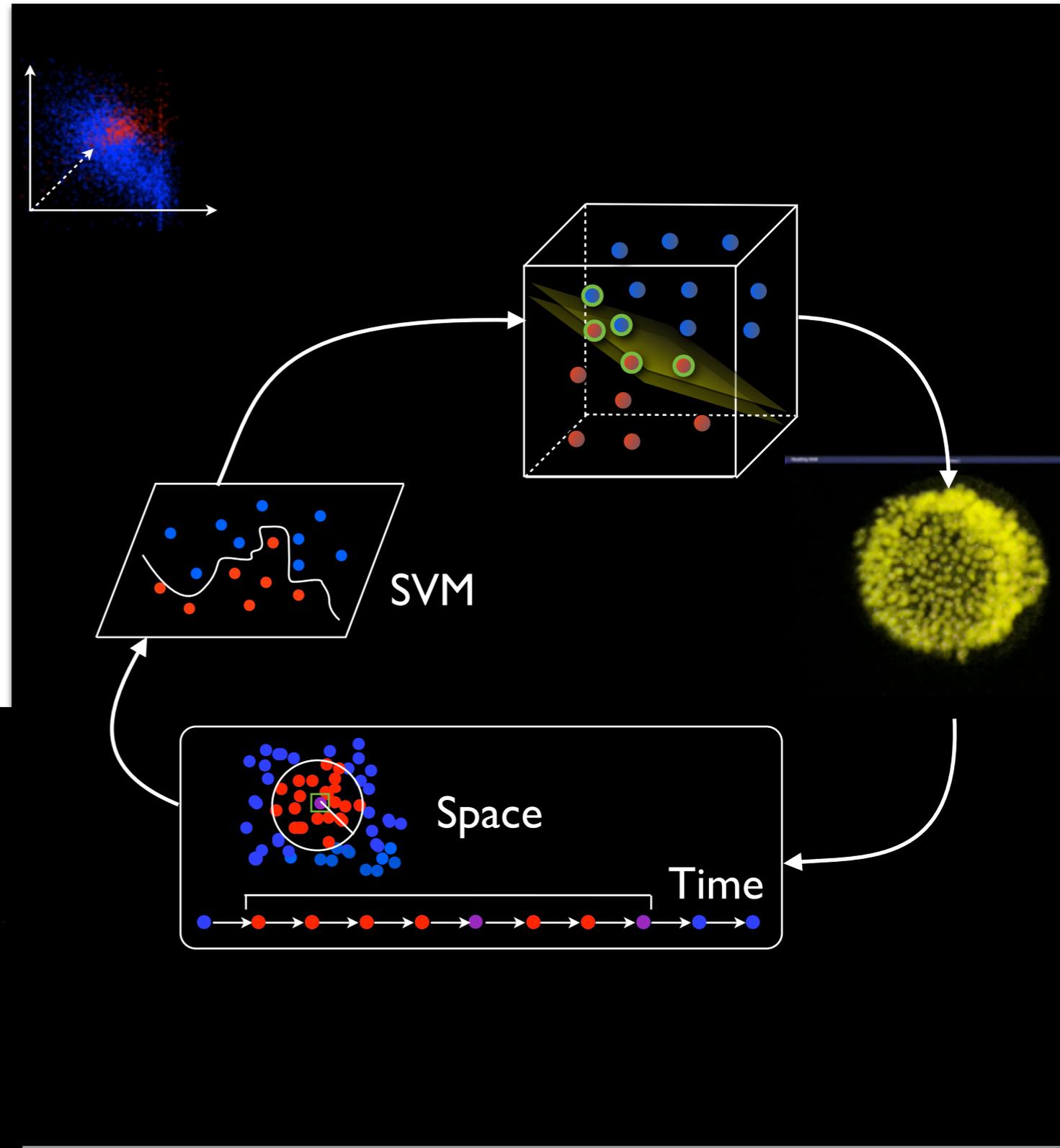
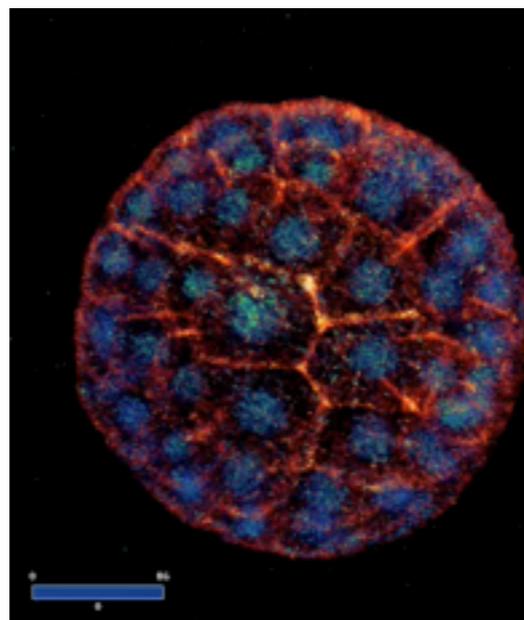
**Apprentissage non supervisé**  
~0.5% cellules en division

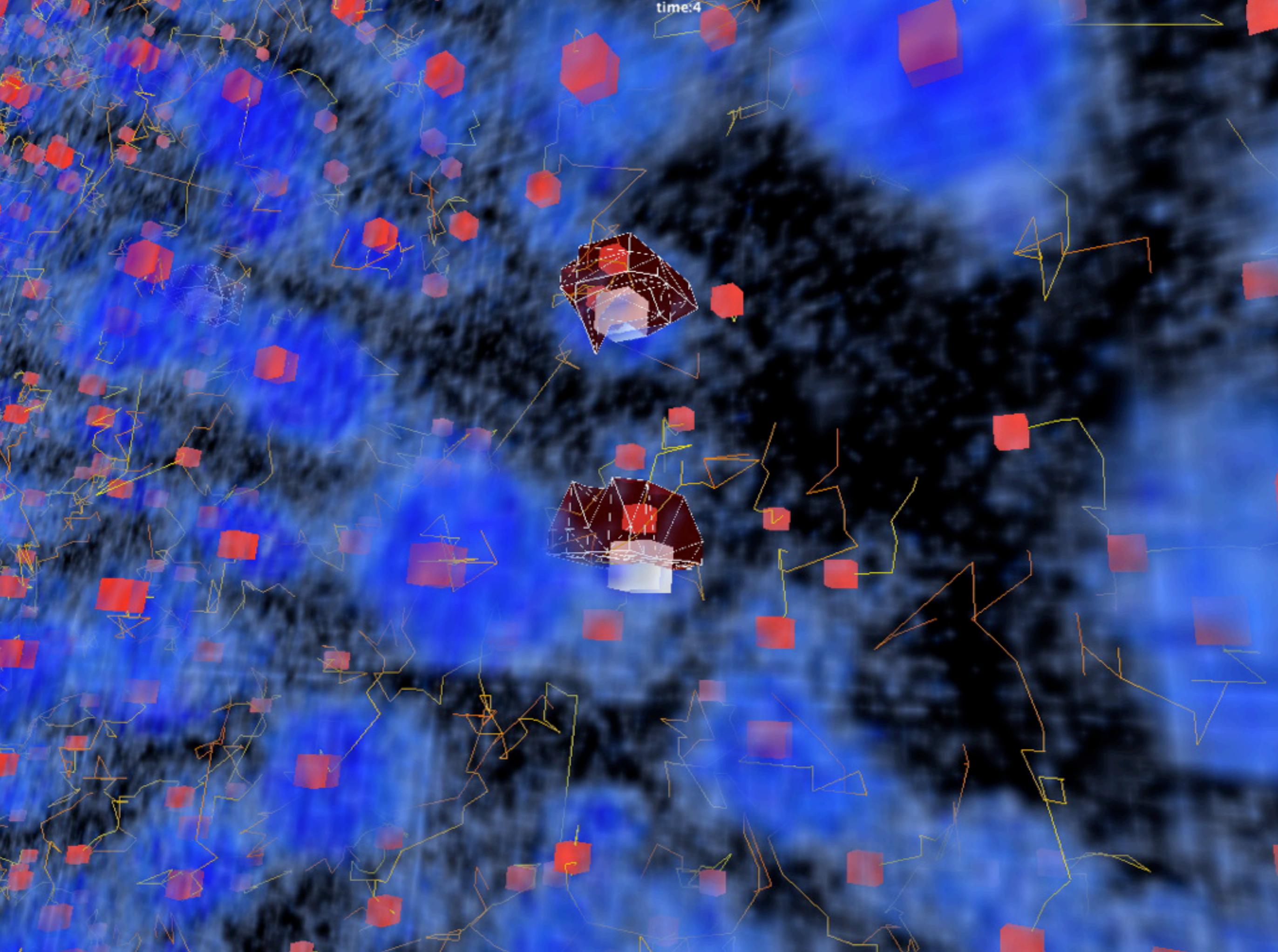


# Détection des divisions cellulaires

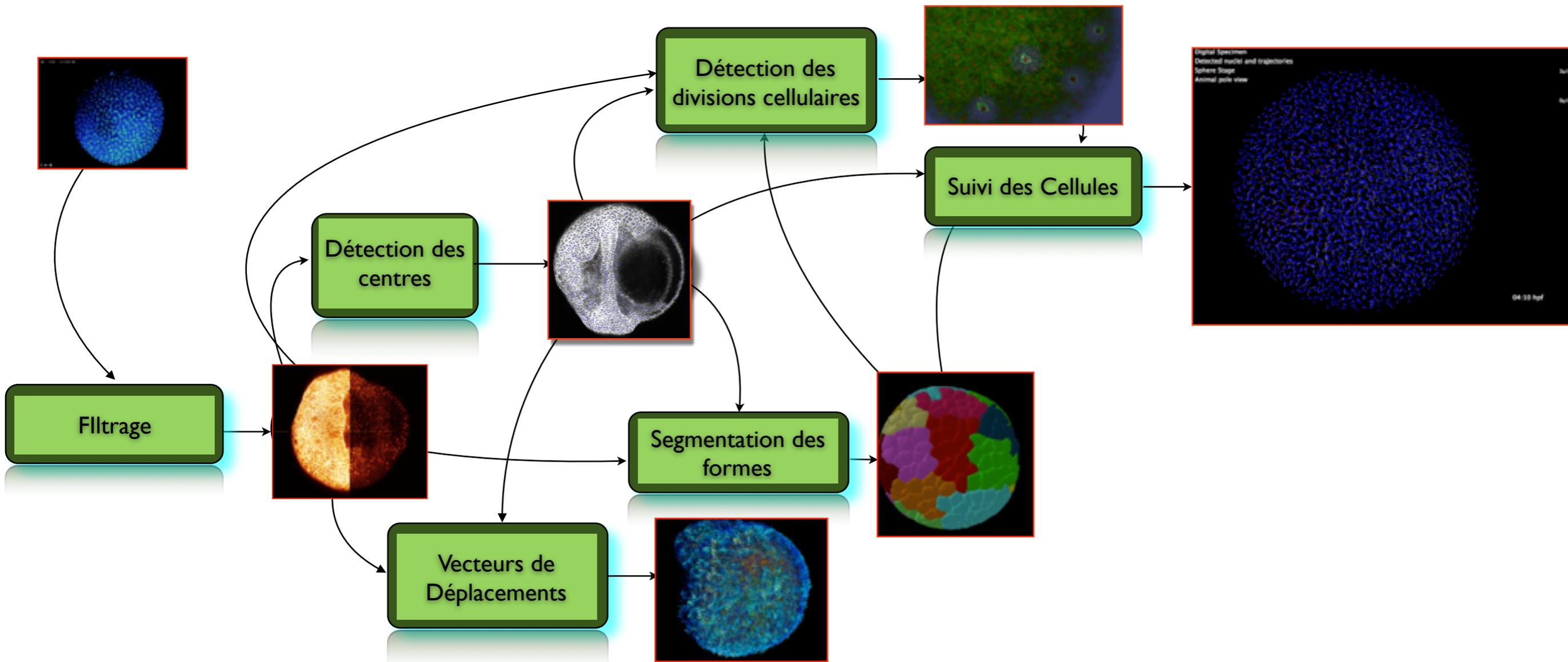
## Apprentissage actif

- Pré - classification définie par la connaissance
- Estimation de densité
- 1er embryon
  - a) Annotation manuelle à la frontière des clusters
  - b) Extension des labels dans l'espace et le temps
  - c) Redéfinition des frontières
- 2ème Embryon

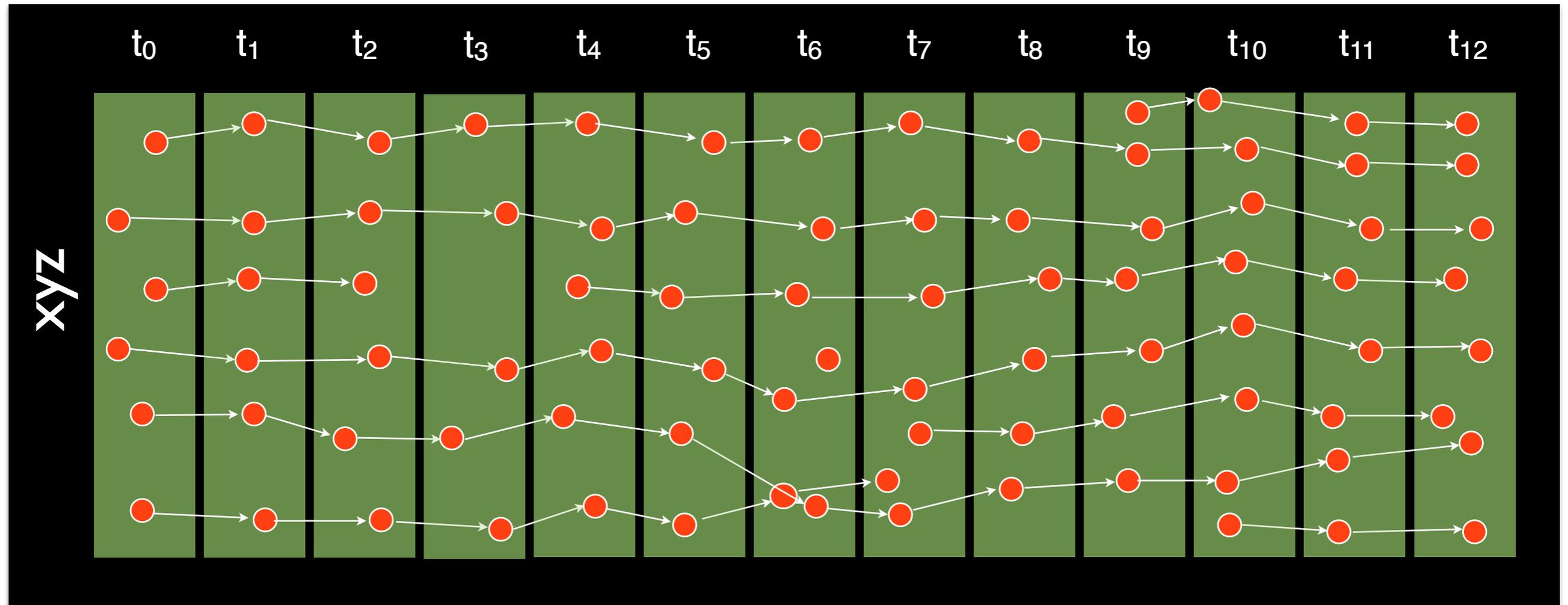




# Reconstruction Image 4D

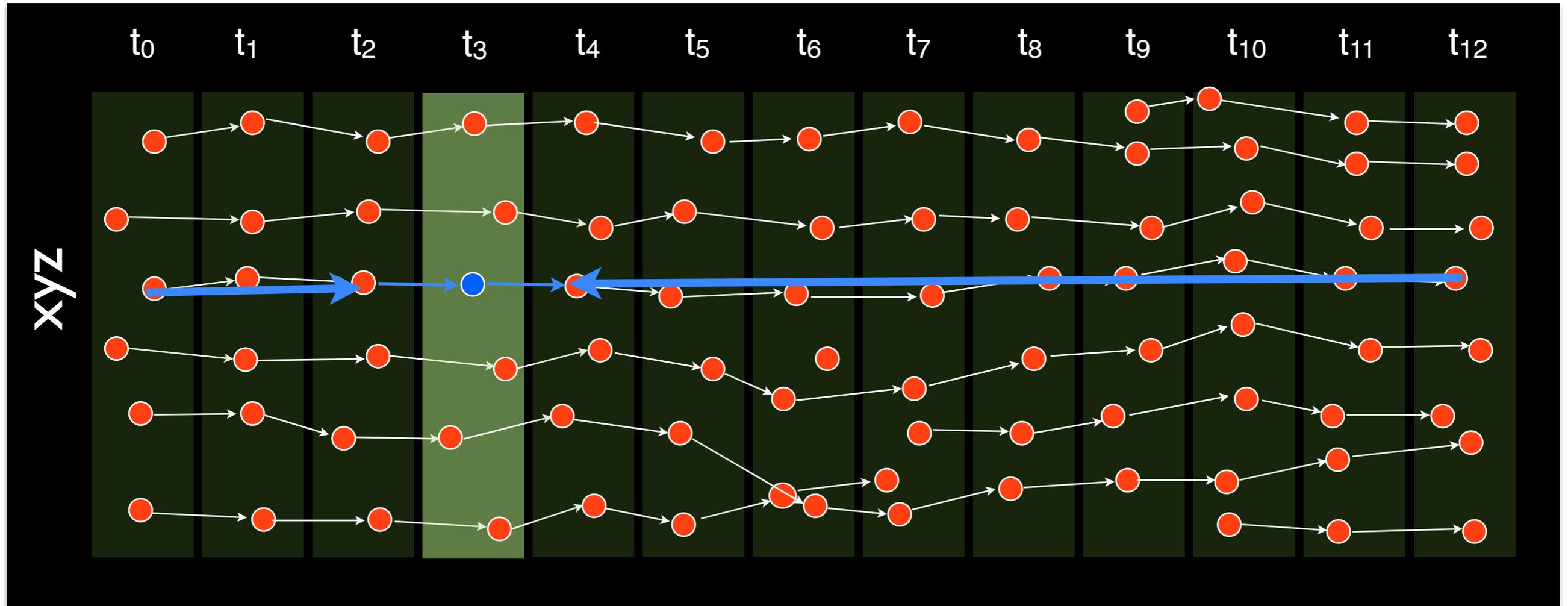


# Suivi cellulaire par plus proche voisins



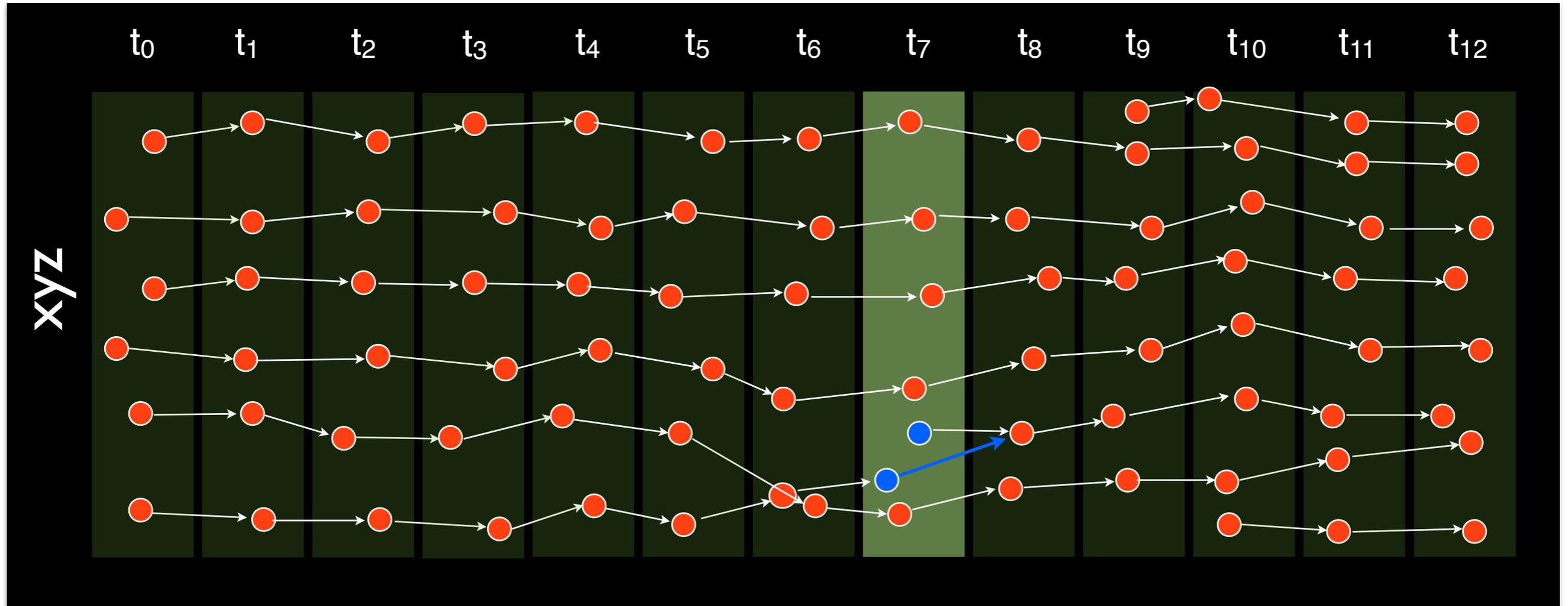
~ 90% de bons liens

# Erreurs ....



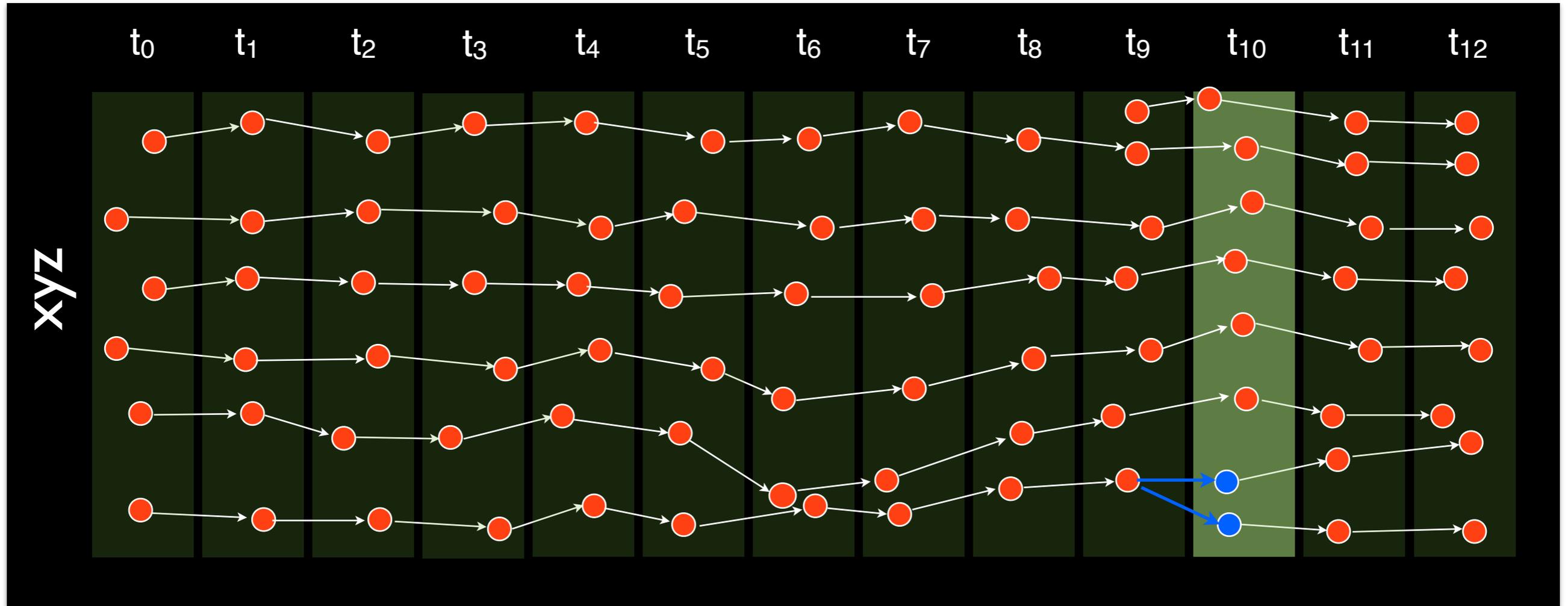
Faux Négatif

# Erreurs ....



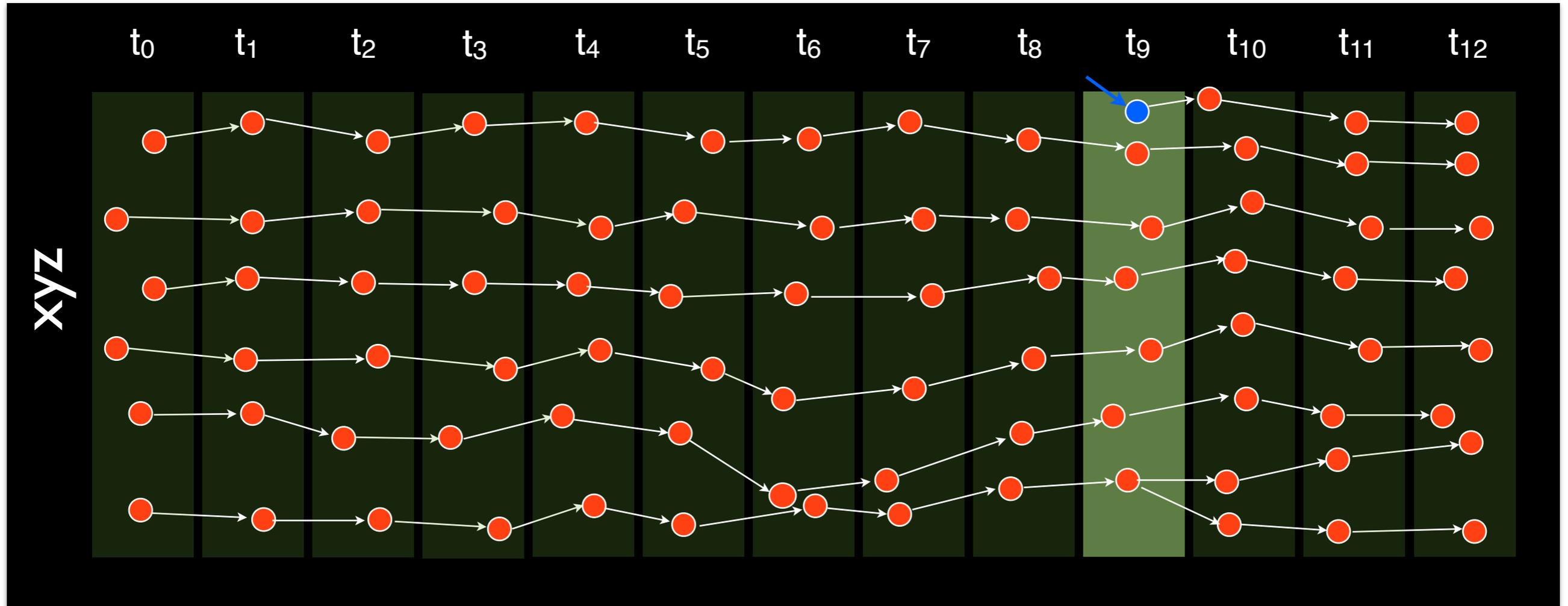
Faux positifs

# Erreurs ....



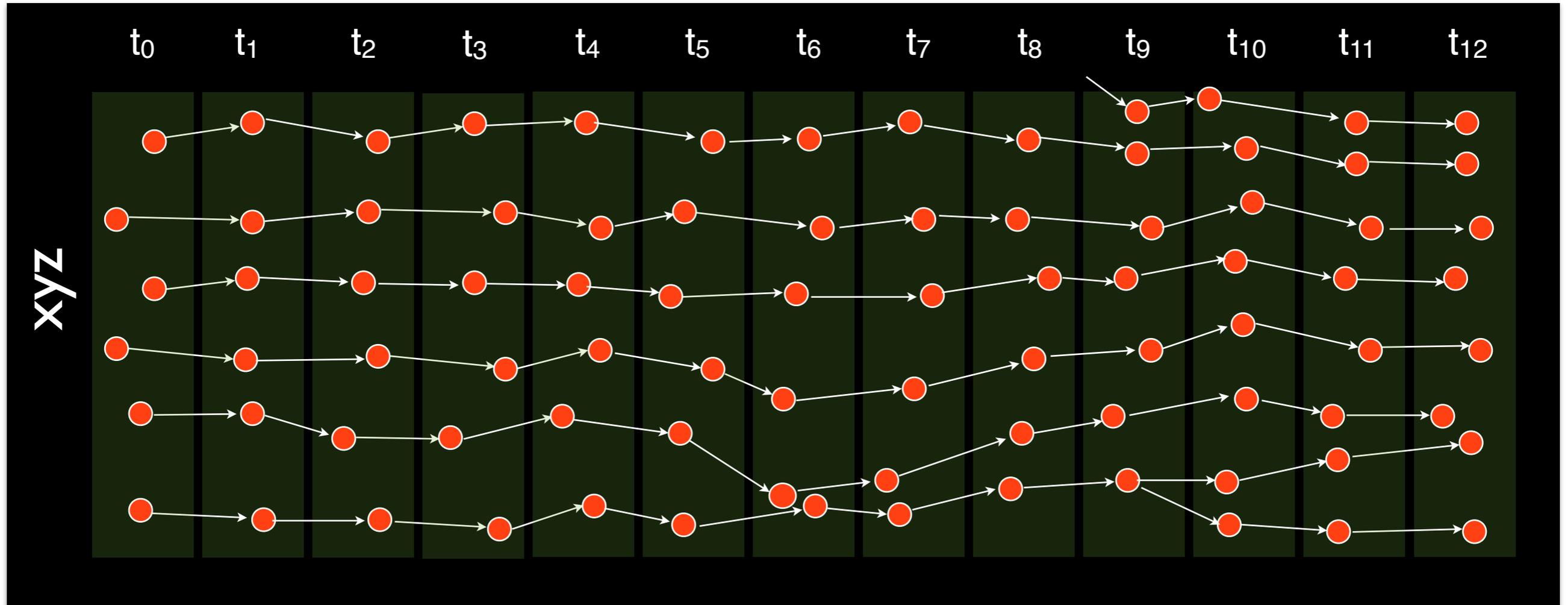
Cellule en division

# Erreurs ....



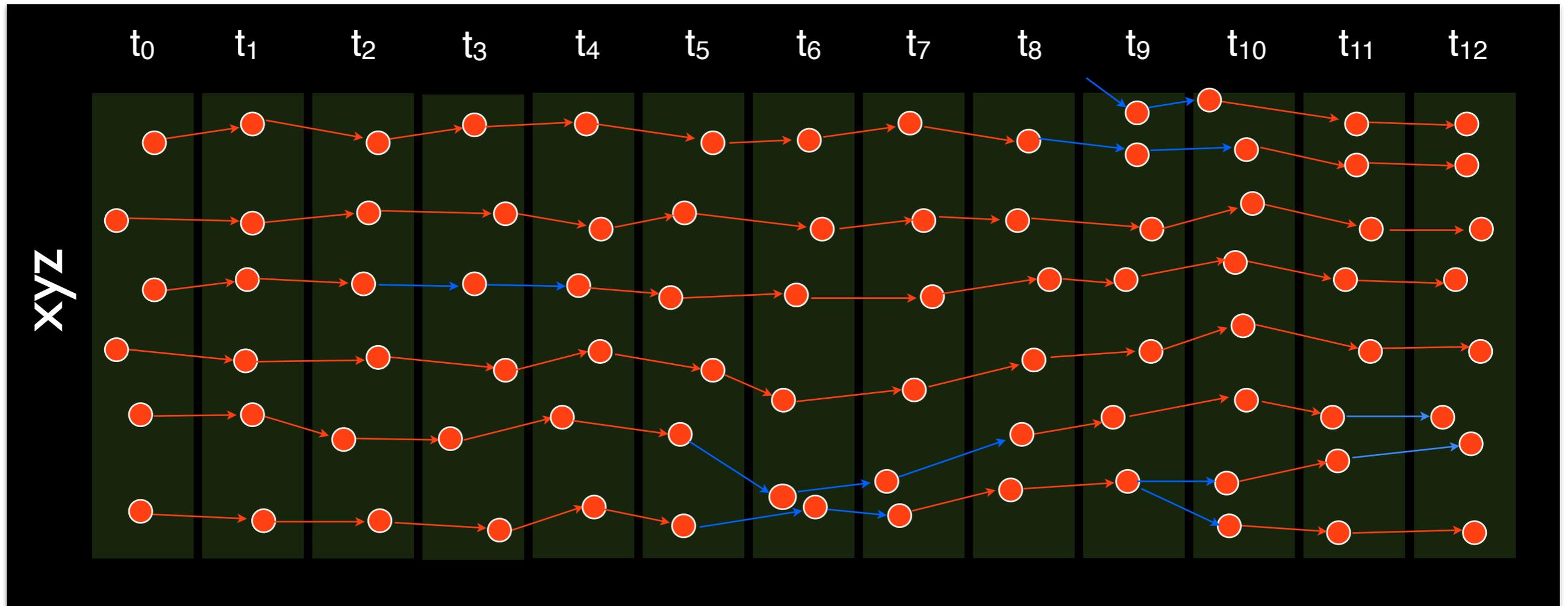
Limite de l'image

# Erreurs ....



100% bon liens ?

# Reconstruction probabiliste

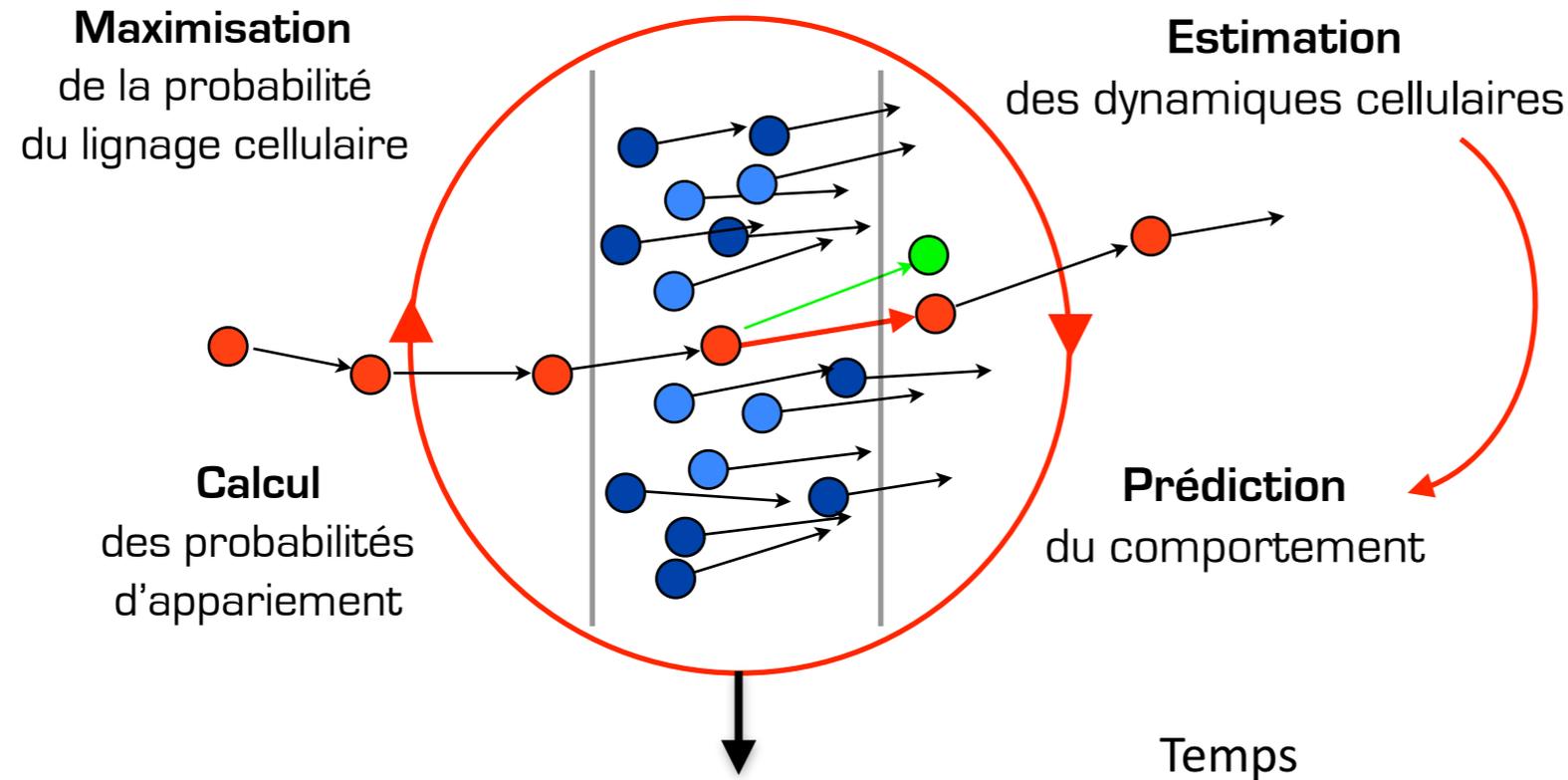


→  $P(\text{link})=1$

→  $P(\text{link})<1$

# Suivi cellulaire probabiliste par maximisation de la vraisemblance

## Propagation des certitudes par itération (Algorithme EM)



**Assimilation par les données**

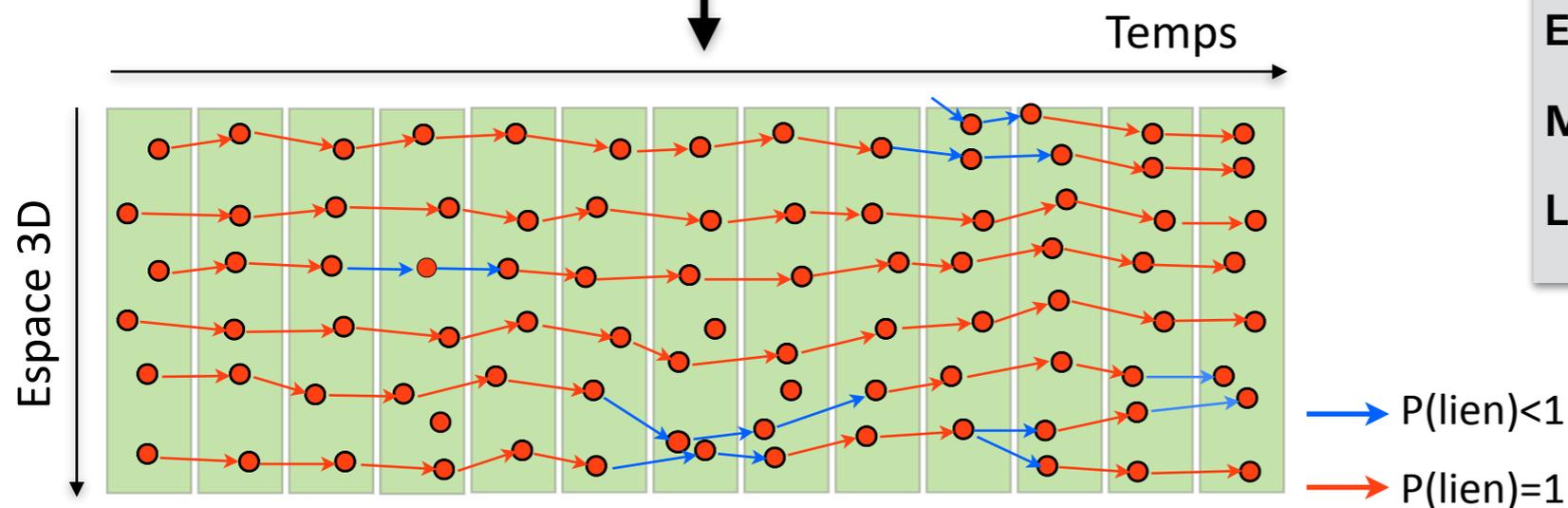
### Résultats :

**Test :** plusieurs organismes

**Evaluation :** % d'erreurs  $\leq$  autres méthodes

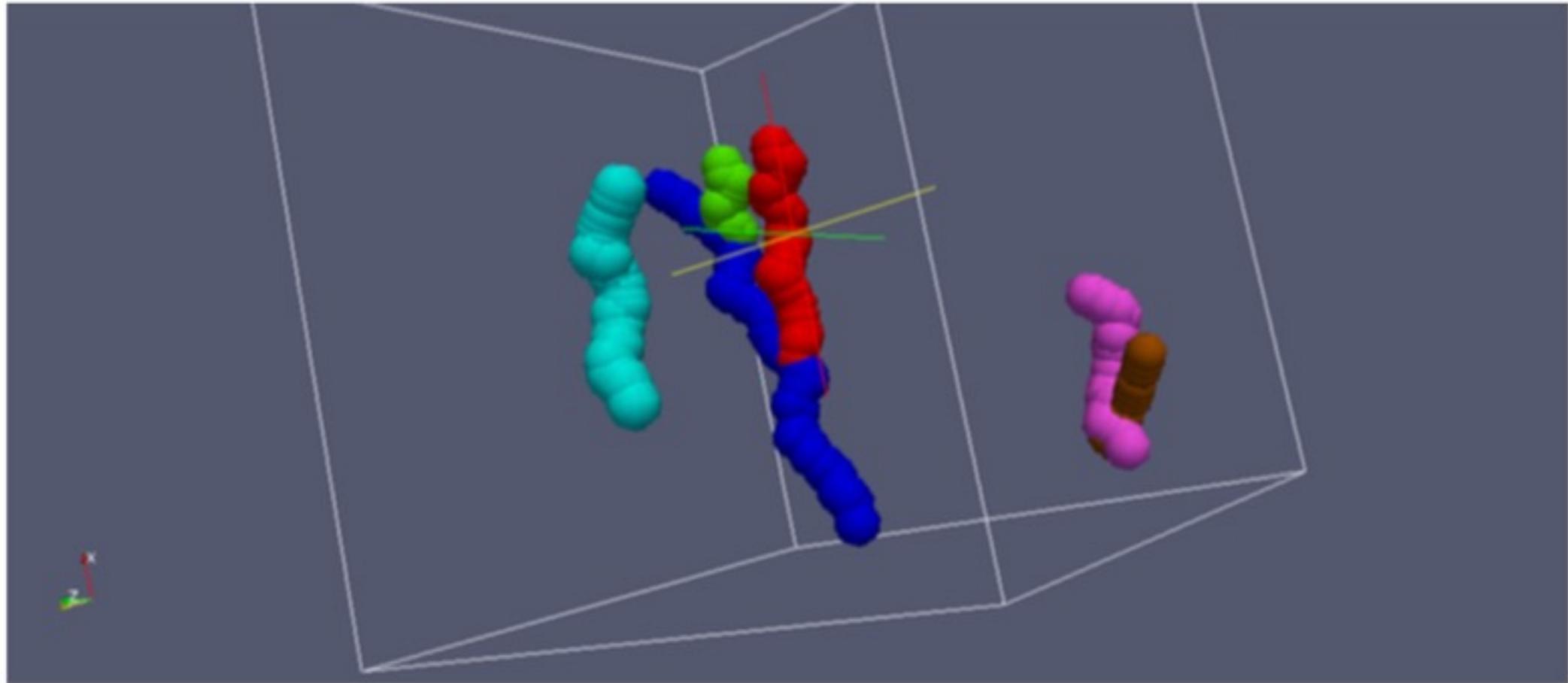
**Méthode sans paramètres**

**Lignage cellulaire facilement corrigible**



**Embryon numérique avec une probabilité associée à chaque lien**

# Segmentation de trajectoires 4D : 3D projection

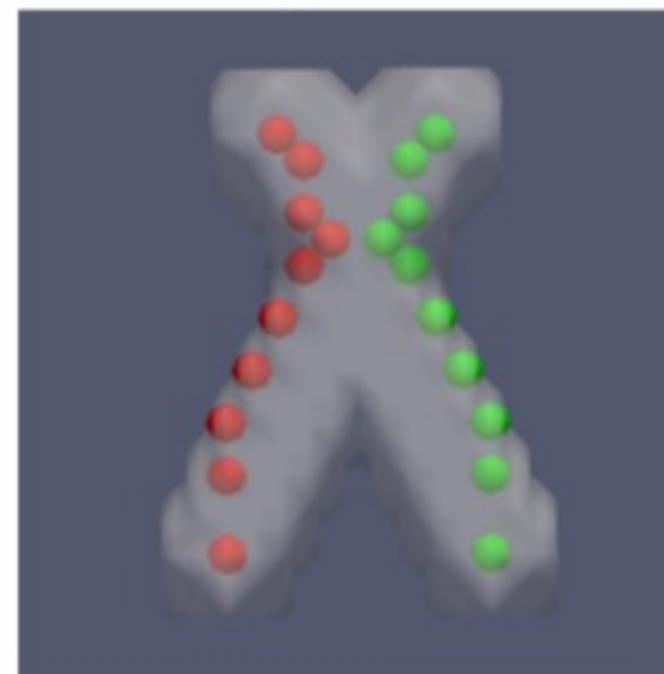


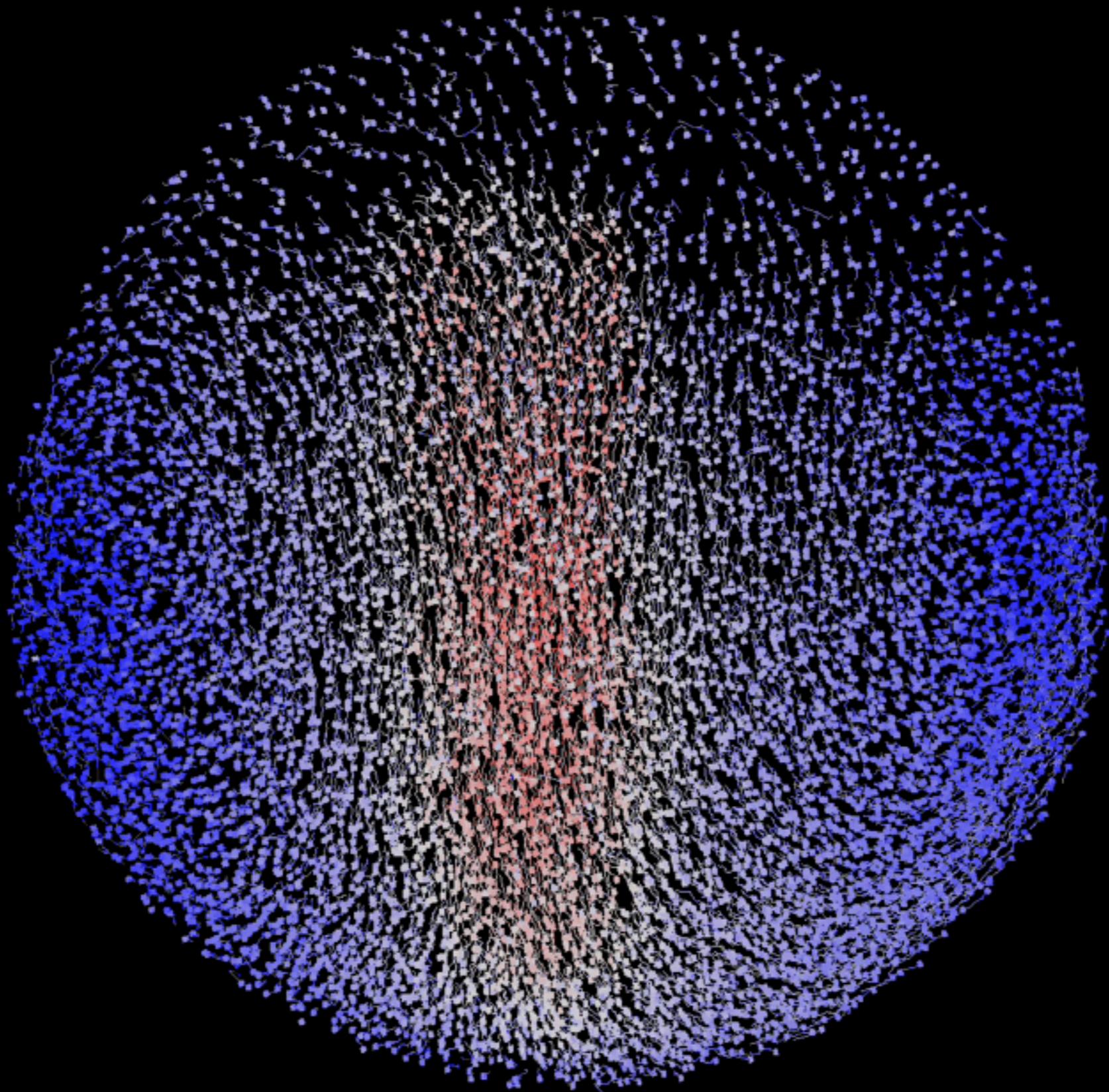
Mikula 2014

Divisions

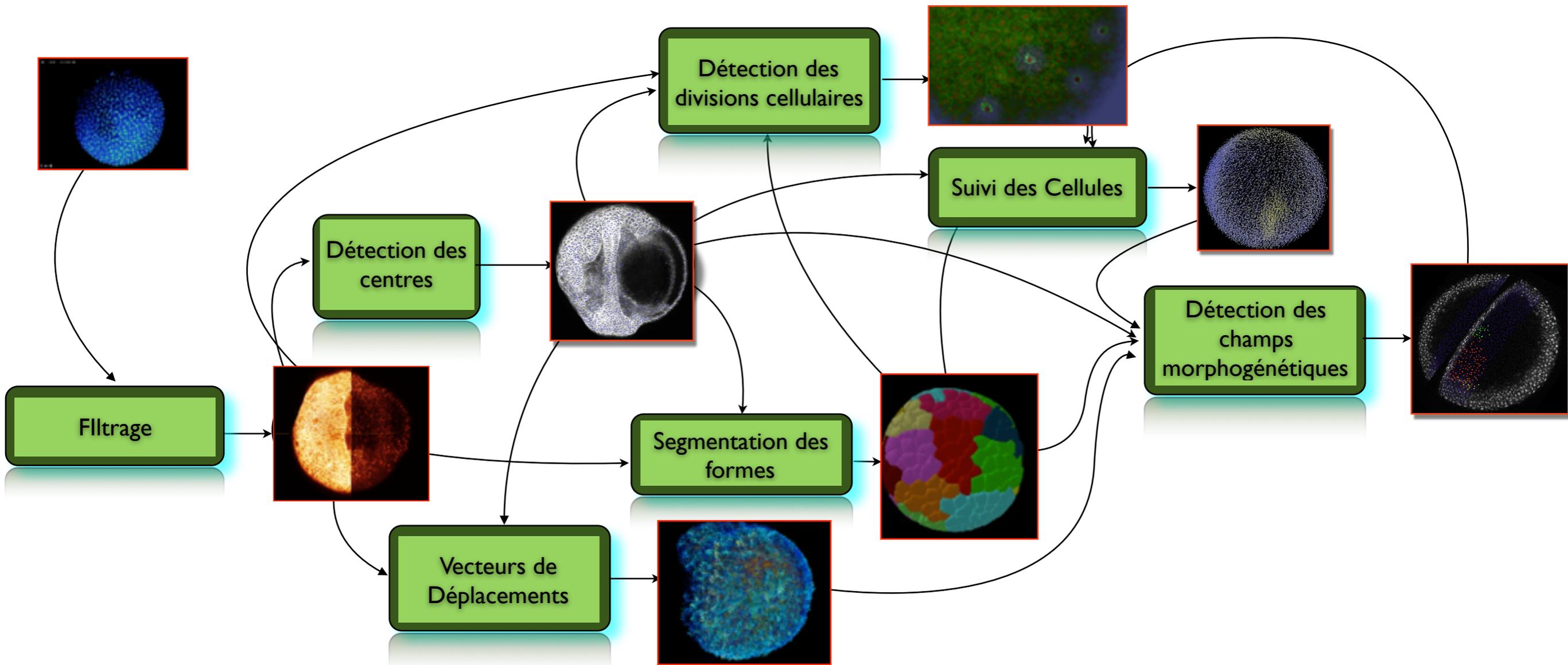


2 trajectoires

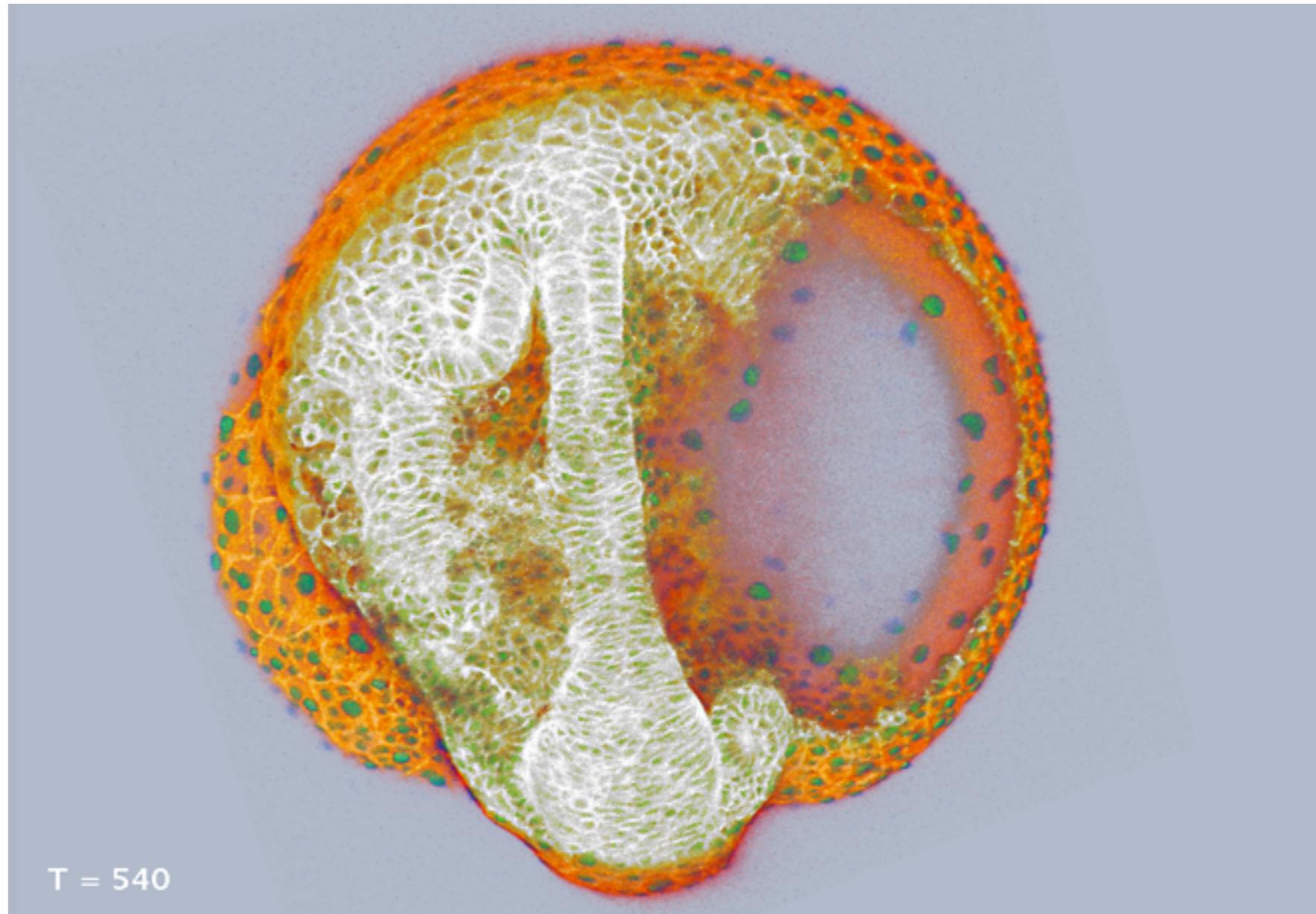




# Reconstruction Image 4D



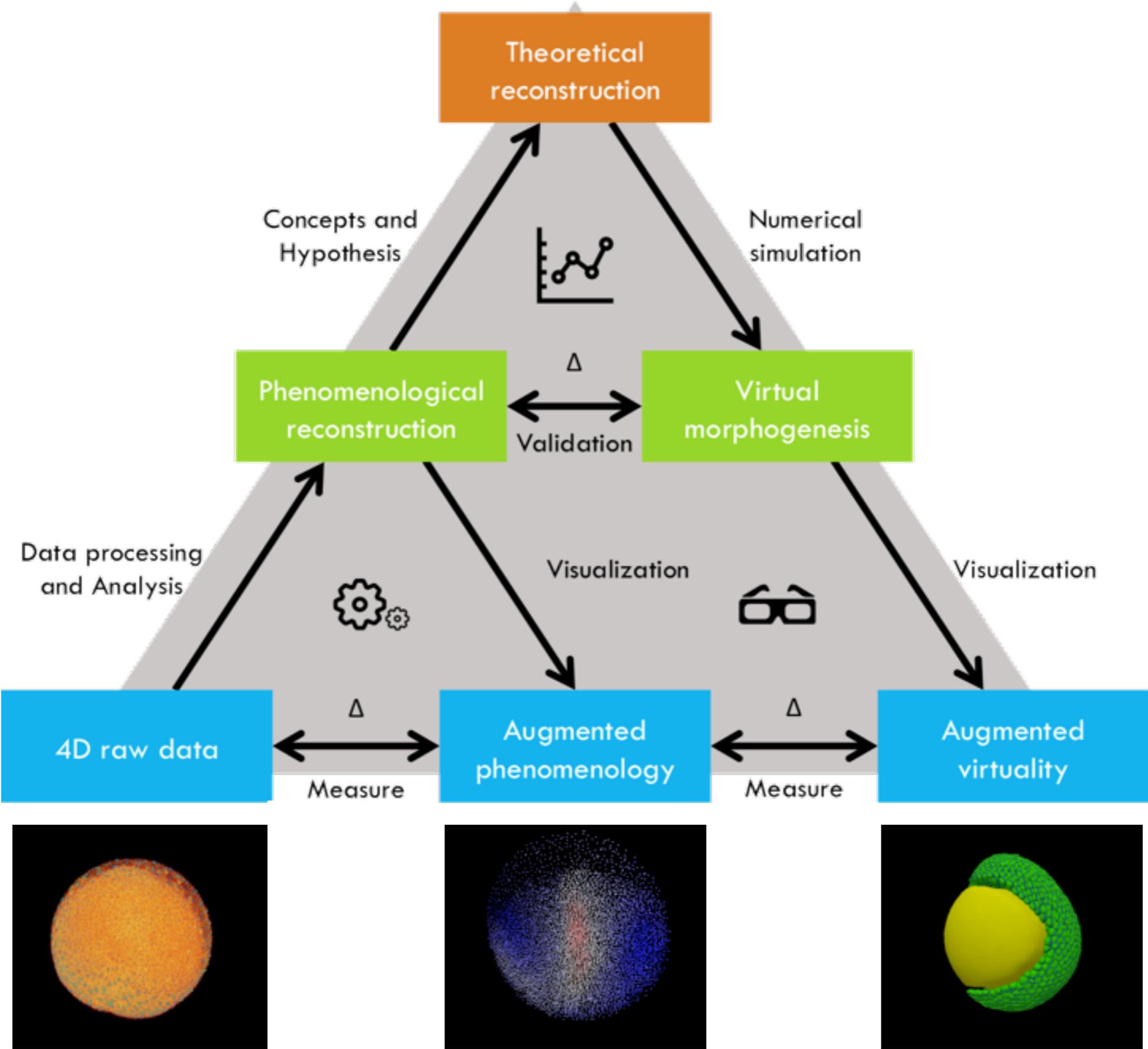
# Champs morphogénétiques



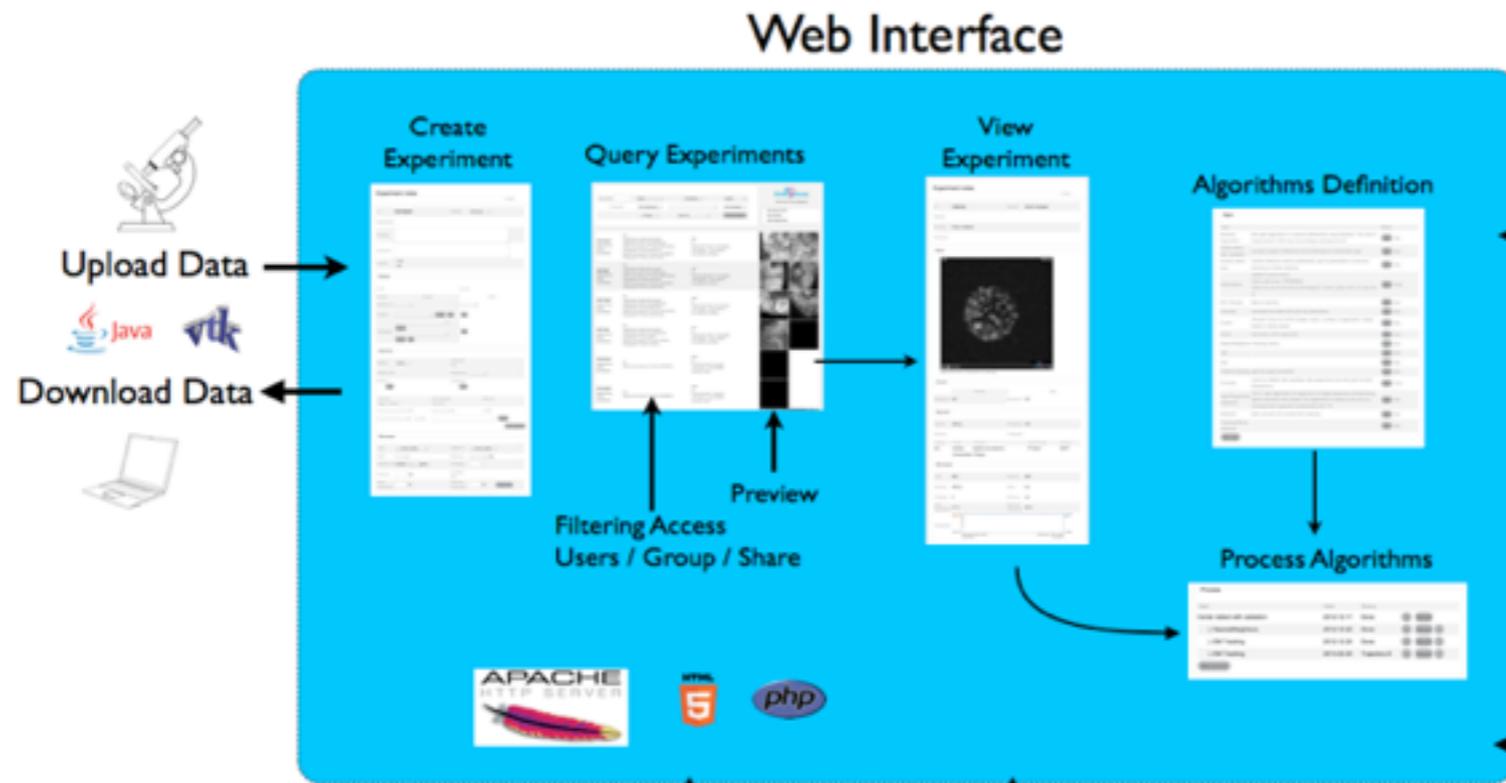




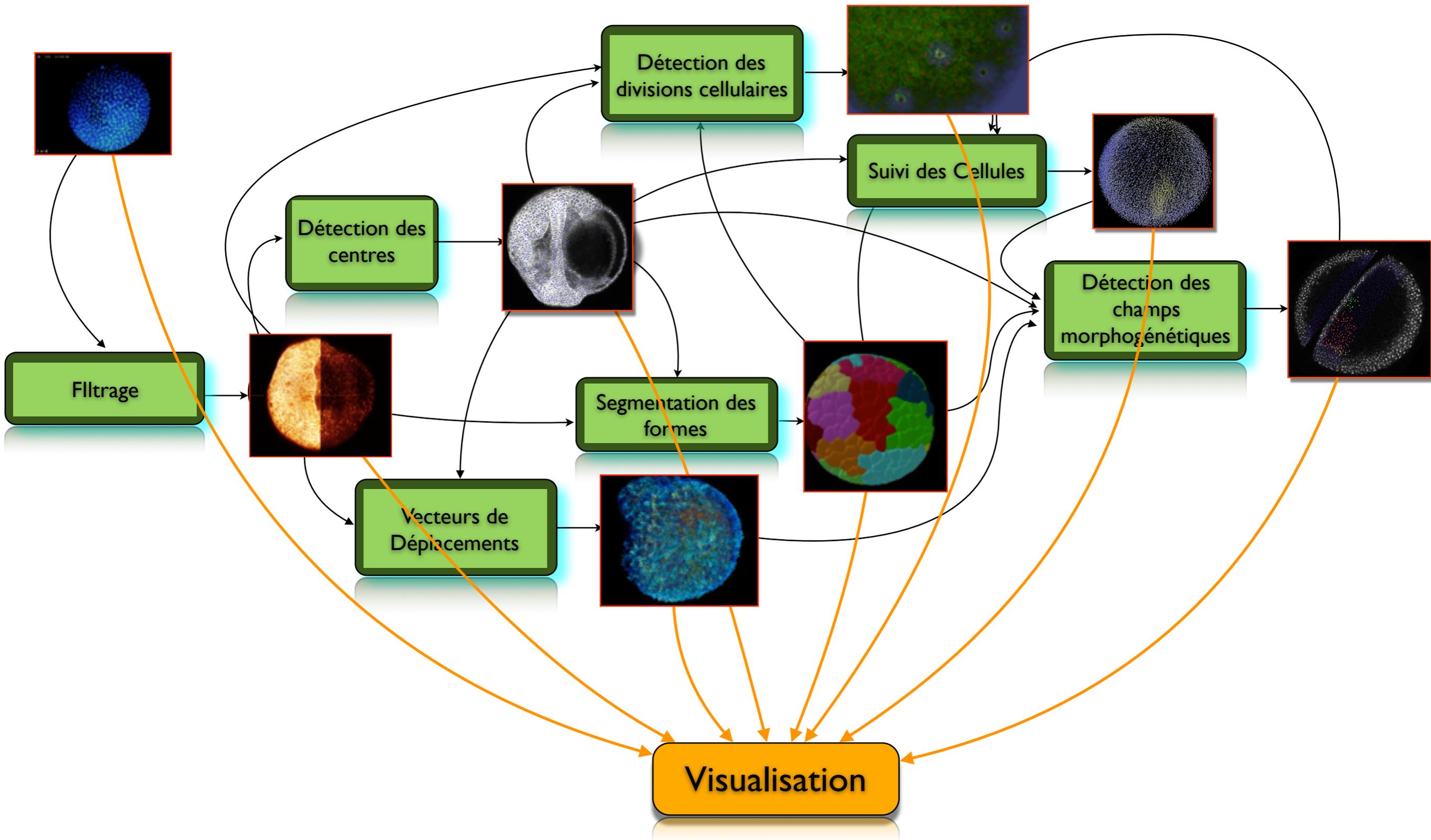
# Approche épistémologique



# Traitement des données 4D



# Visualisation d'images 4D



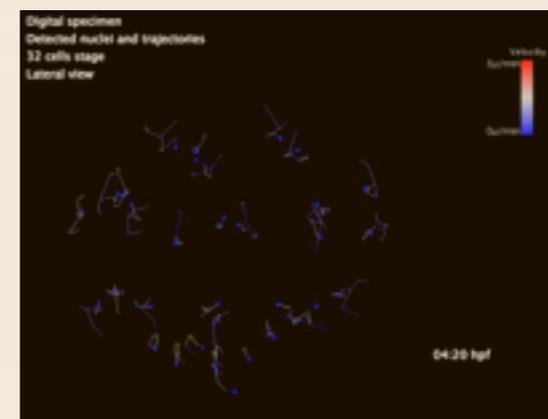
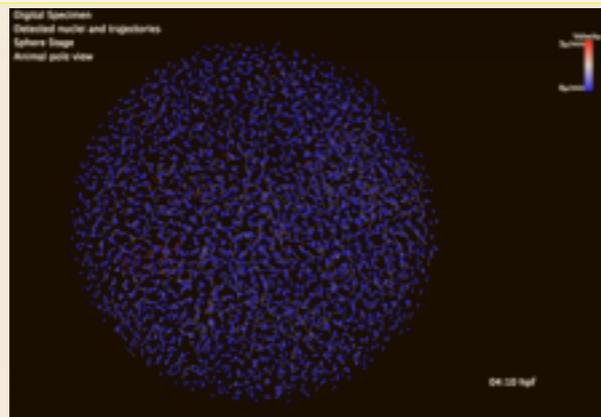
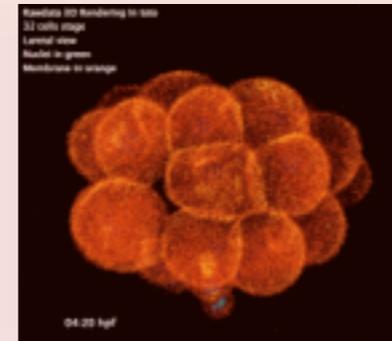
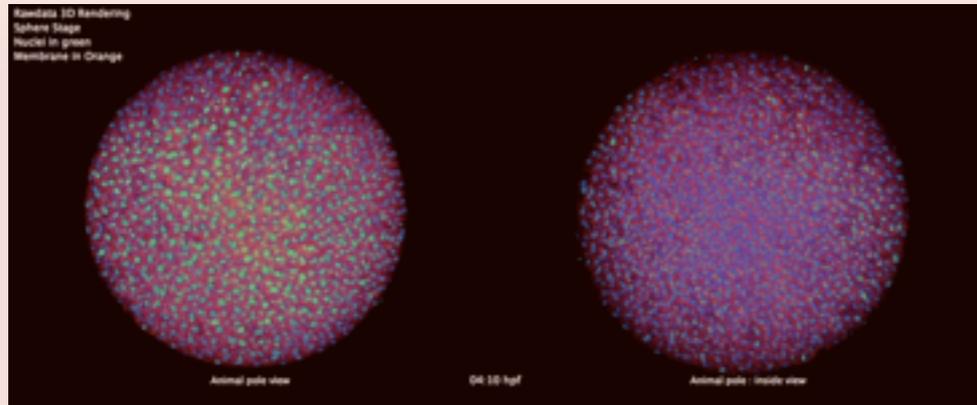
# MovIT : Interface de Visualisation Interactive

[ Faure & al Nature Communication 2016]

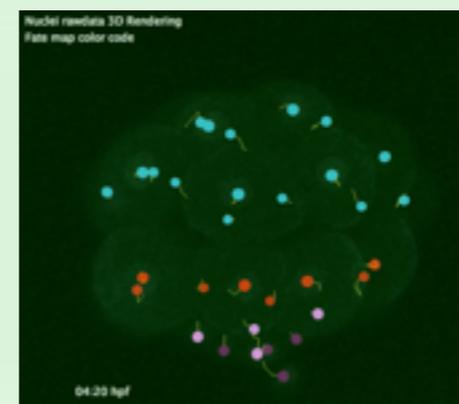
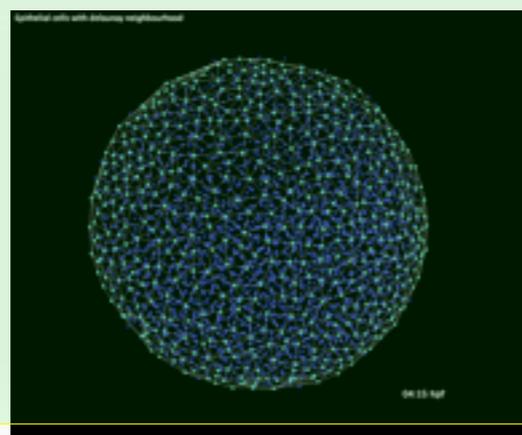
## Zebrafish

## Sea Urchin

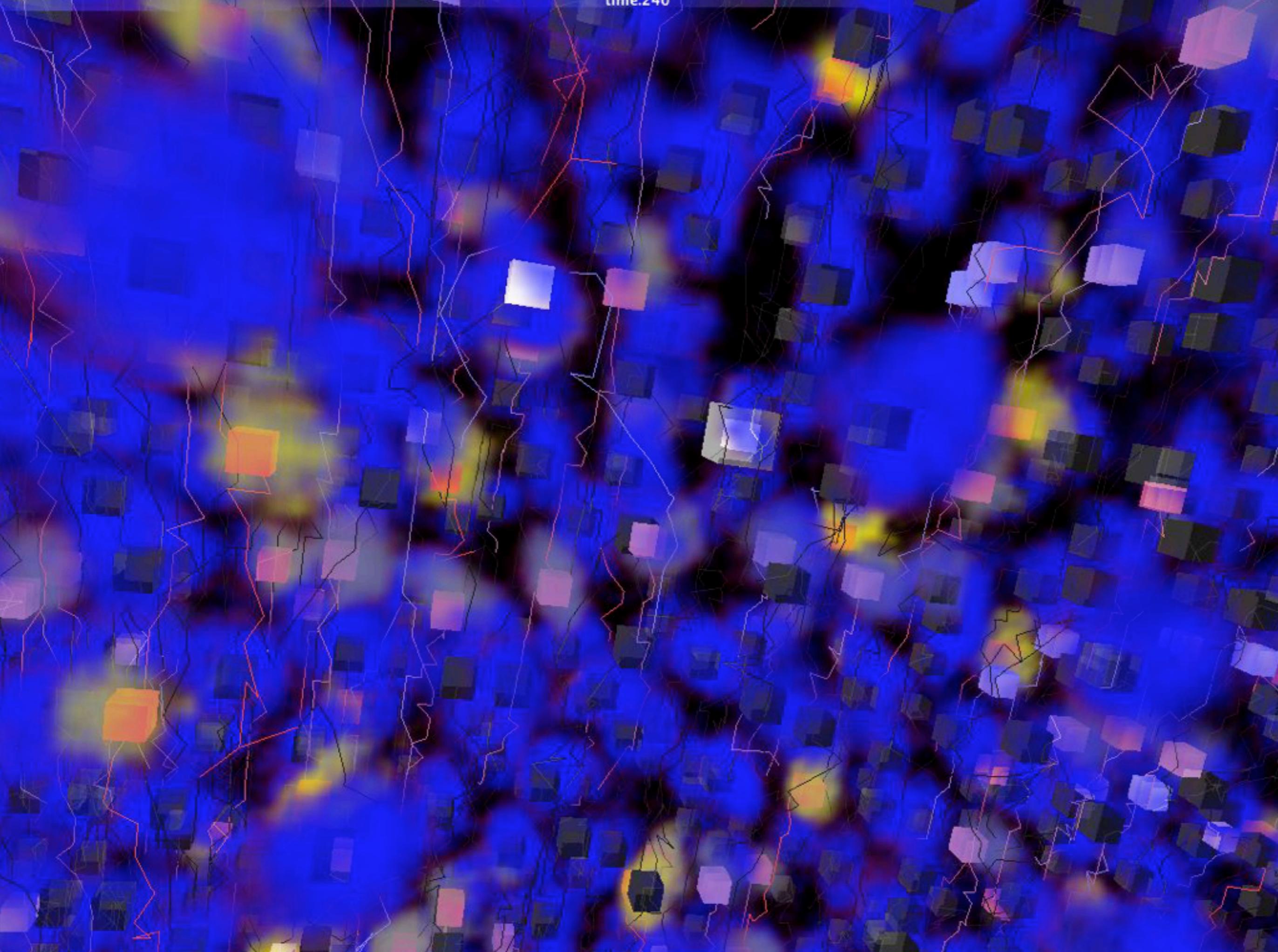
Observation



Digitizing



Abstraction



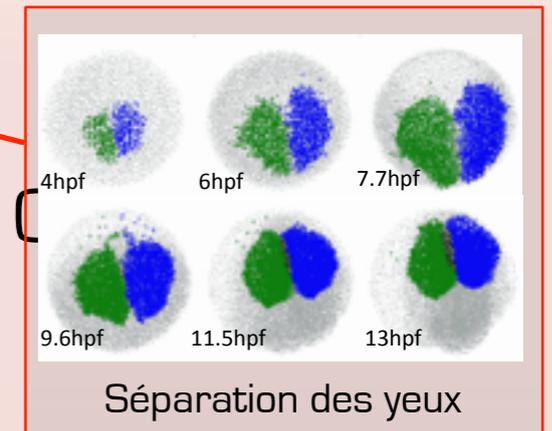
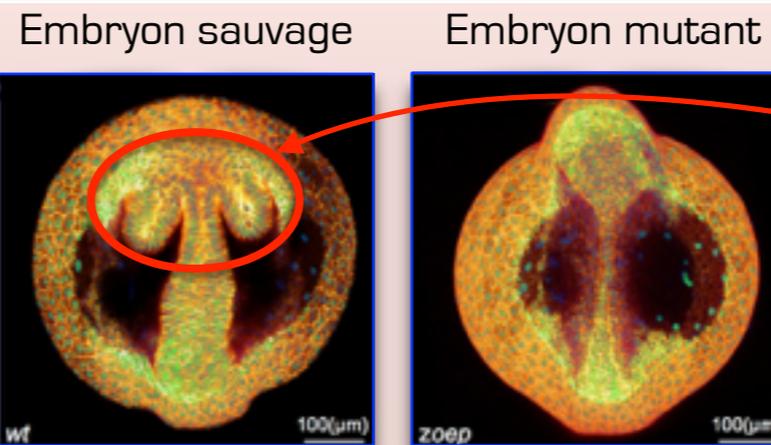
# Reconstruction des dynamiques multi-échelles de la morphogenèse animale

[ Faure & al Nature Communication 2016 ]



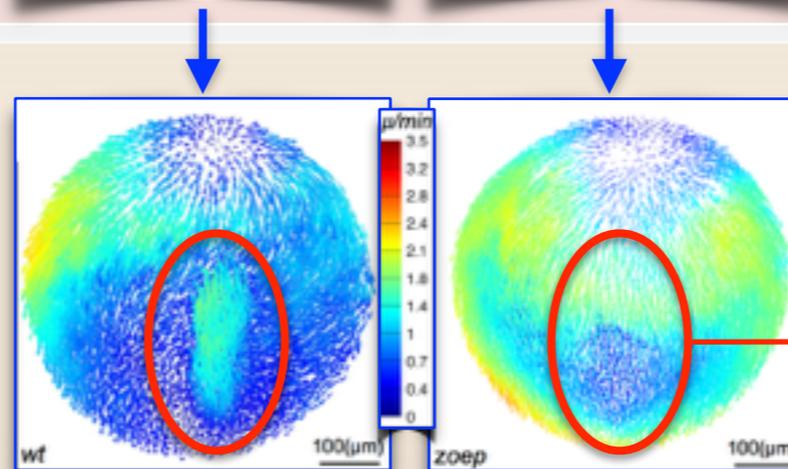
## Résultats :

**Enjeu** : Compréhension des dynamiques cellulaires



**Objectif** : Comparaison d'embryons numériques

**Méthode** : Reconstruction automatisée par un web-service

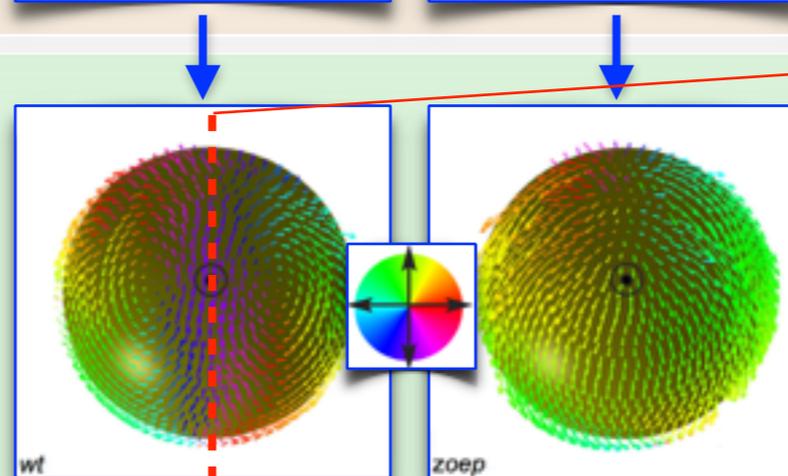


**Digitalisation**  
zoep dépourvu d'une partie de l'hypoblaste

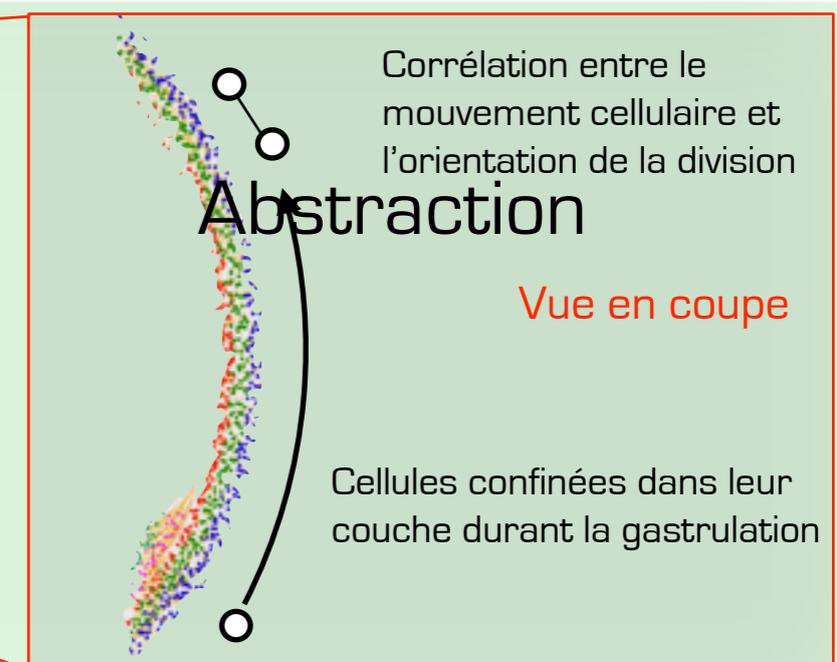
**Objectif** : Construction de prototypes (développement moyen)

**Méthodes** :

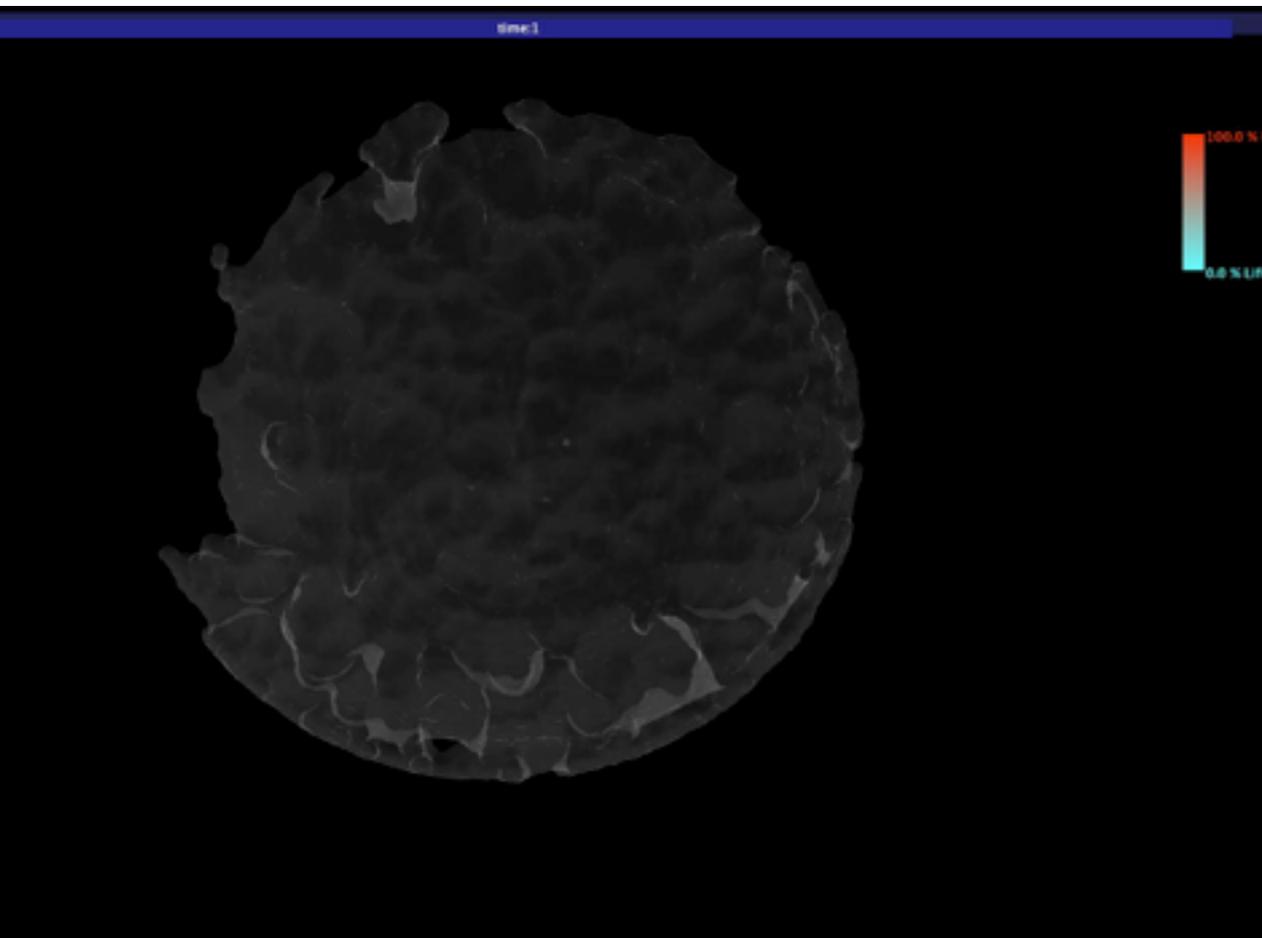
- Recalage 3D (Images et objets)
- Recalage temporel (Dynamiques)
- Moyennage local



Prototypes



# Division Prototype for zebrafish early embryo cell lineage



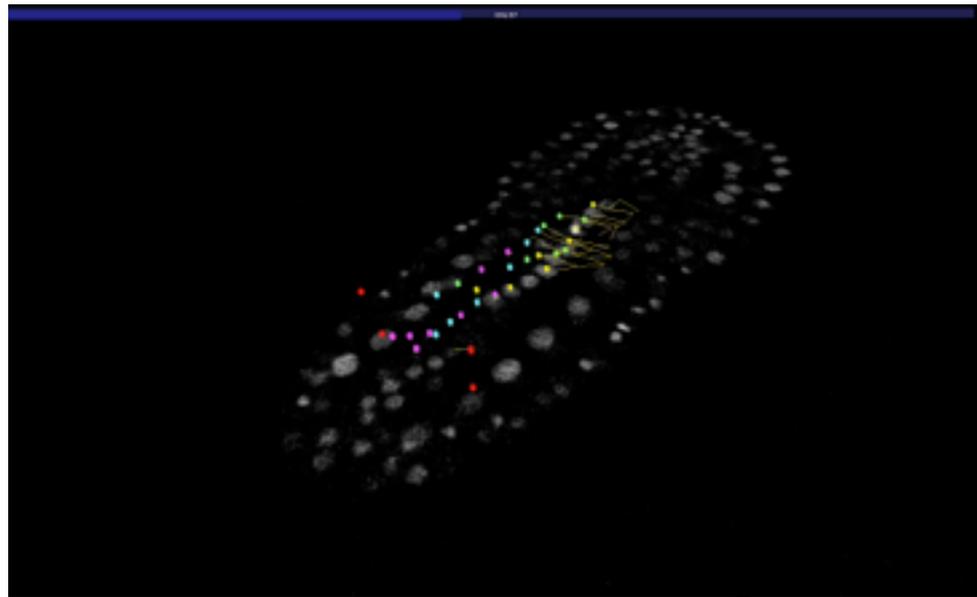
Time 0 min



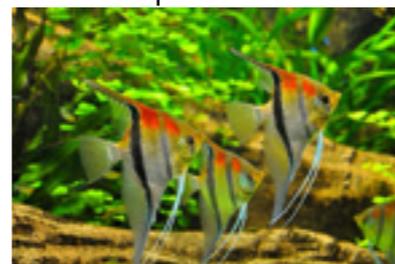
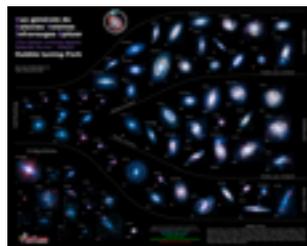
[ Olivier & al Science 2010 ]

# Ground truth

- analyses des données
- amélioration des algorithmes
- évaluation des méthodes



## Plateforme de validation



- détection
- classification
- segmentation
- tracking



# Jeux Sérieux : Validation d'images scientifiques

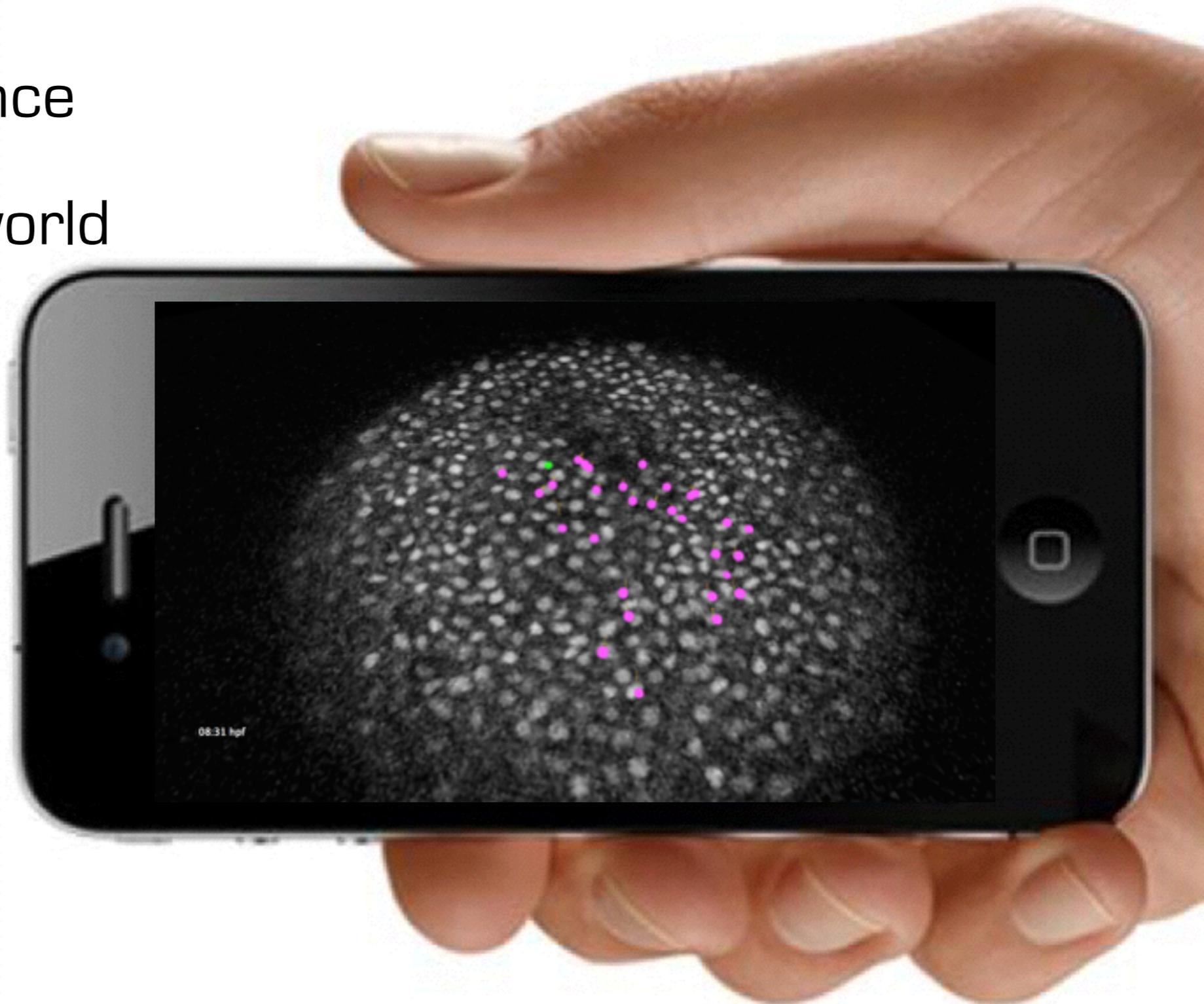
 Citizen Science

 Varieties of world

 Education

 Levels

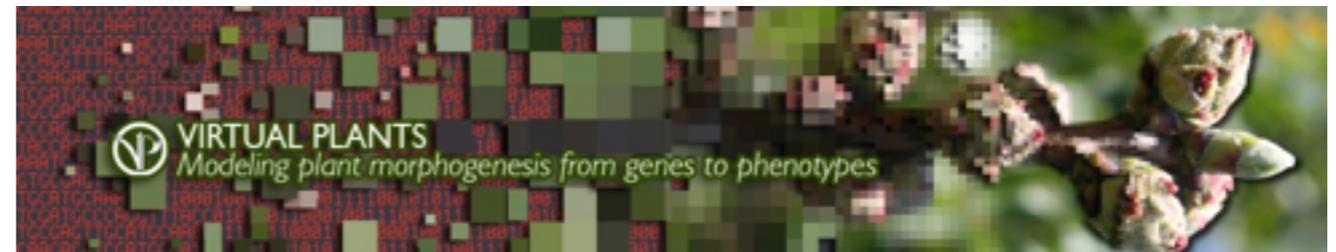
 Diffusion



# Merci !

**BiO  
Emergences**

**Nadine Peyri ras  
Paul Bourguine**



**Christophe Godin  
Leo Guignard**



**Patrick Lemaire**

