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# Cancer as an evolutionary process

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## Outline

- Introduction
- Cancer initiation
- Cancer progression
- Genetic heterogeneity and treatment
- Dependencies among clonal mutations

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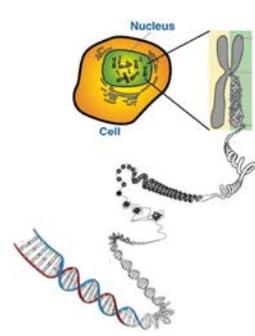
## Introduction

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## Cancer is a genetic disease

- Cancer cells harbor various types of genetic alteration, including
  - point mutations
  - insertions
  - deletions
  - chromosome rearrangements
  - mitotic recombination
  - loss or gain of whole chromosome arms

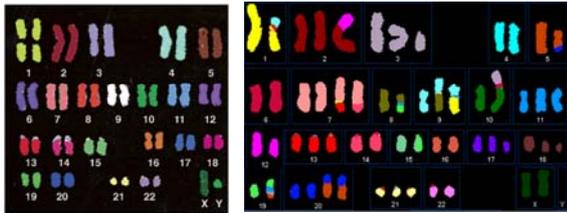


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## Most cancer cells are *aneuploid*



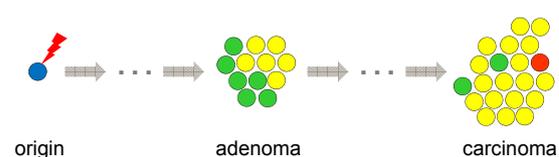
normal karyotype      karyotype of a colon cancer cell

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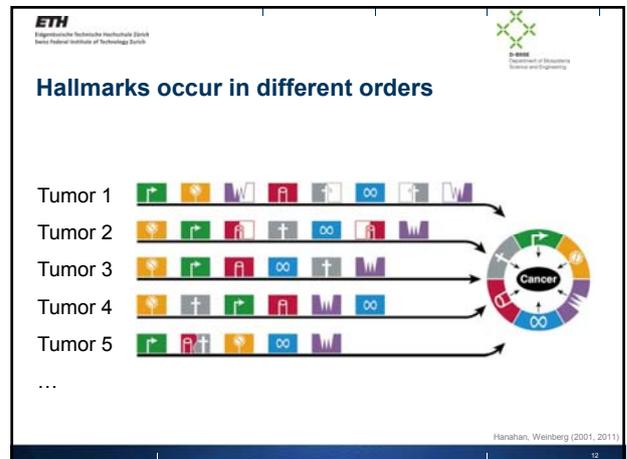
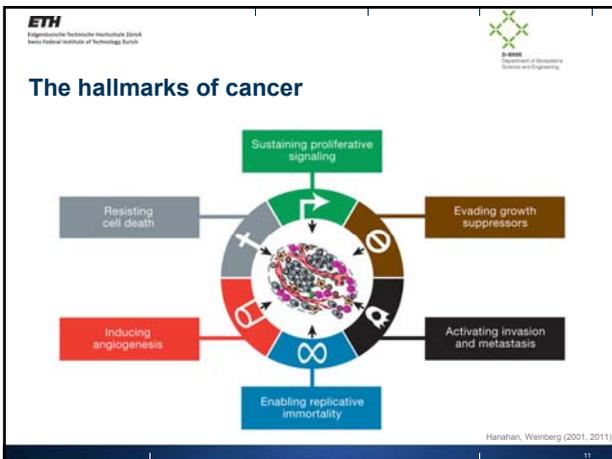
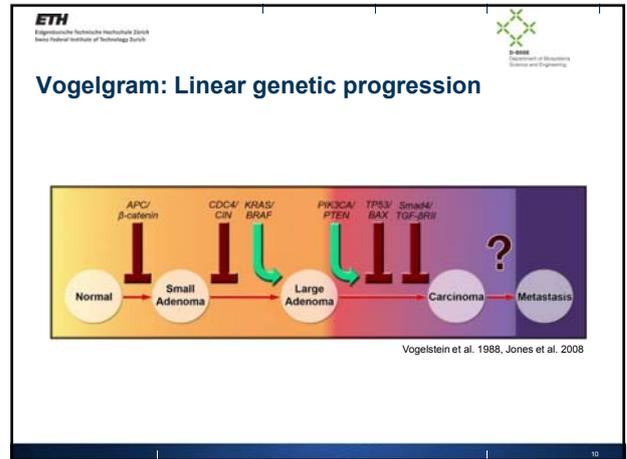
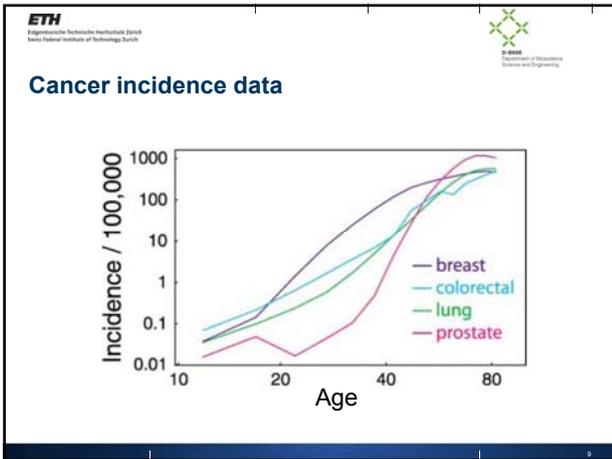
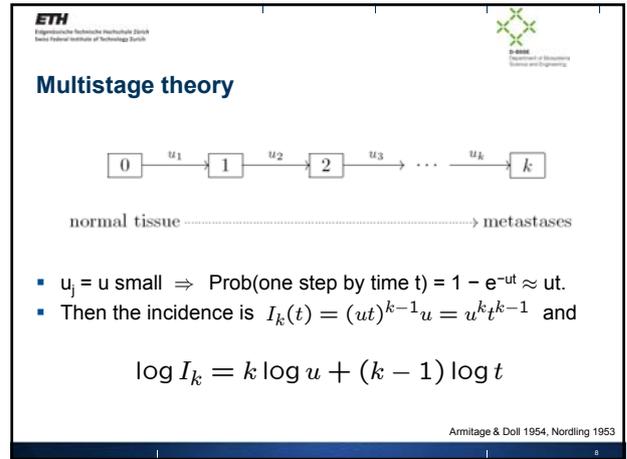
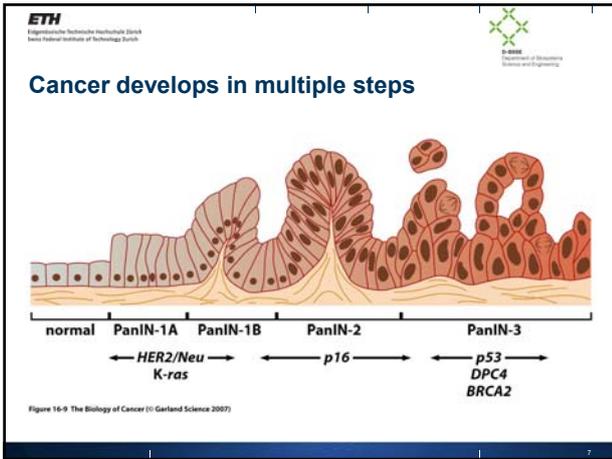
## Cancer progression



origin      adenoma      carcinoma

Genetic progression (accumulating mutations)

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**Cancer is heterogeneous at multiple levels**

Population    Intra-patient  
Spatial, temporal    Intra-tumor  
Tissue    Intra-tumor  
Genetic

Florian Markowitz

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**Inter-patient and intra-patient diversity**

Inter-tumour heterogeneity  
Intra-tumour heterogeneity

Dominance of clone 1    Dominance of clone 2    Mixed dominance

Manuyak et al (2012)

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**Cancer as an evolutionary process (Nowell, Science 1976)**

**The Clonal Evolution of Tumor Cell Populations**

Permits stepwise selection earlier tumor progression.

Peter C. Nowell

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**Cancer ecology and evolution**

Selective pressures    Tx

Ecosystem 1    Ecosystem 2    Ecosystem 3    Ecosystem 4

Single founder cell (stem or progenitor)

Confined (CIS)    Diffuse    Metastases

Recurrence

ecosystem = tissue microenvironment

● ● ● Subclones with unique genotype / 'driver' mutations

Greaves and Maley (2012)

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**Molecular profiling of tumors**

Tumour

DNA    RNA    Protein    ChIP

Van't Veer et al (2002)    http://mss.kcl.gov    Ross-Innes et al (2012)

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**Cancer initiation**

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**Finite populations: The Moran process**

$P_{i,i+1} = \frac{i}{N} \left(1 - \frac{i}{N}\right)$   
 $P_{i,i-1} = \left(1 - \frac{i}{N}\right) \frac{i}{N}$   
 $P_{i,i} = \left(\frac{i}{N}\right)^2 + \left(1 - \frac{i}{N}\right)^2$

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**Key results**

- Fixation probability of a single allele with selective advantage  $r$ :  $\rho = \frac{1 - 1/r}{1 - 1/rN}$
- Mean waiting time:  $\tau \propto N^2$
- Rate of loss of diversity:  $2/N^2$

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**Oncogenes**

- Oncogenes increase fitness, if one allele is mutated or inappropriately expressed. They are activated by:
  - a specific point mutation
  - a gene amplification
  - or chromosomal fusion

Nowak (2006)

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**Fixation of oncogene mutations**

- The probability that a mutant with selective advantage  $r$  has been fixed in a (small) population of size  $N$  by time  $t$  is
 
$$P(t) = 1 - e^{-N\mu r t}$$
- Observation: Large compartments accelerate the accumulation of advantageous mutations, small compartments slow it down.
- Most tissues with high cell turnover are organized in many small compartments.
- For example, colonic crypts...

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**The colon is organized into  $10^7$  crypts, each consisting of 1,000 to 4,000 cells.**

colon wall  
 apoptosis  
 cell division and migration  
 stem cell

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**Colon cancer arises in a crypt**

polyp  
( $1\text{mm}^3$ , or  $10^6$  cells)

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**The linear process of cancer**

The mutation has to occur here

$\rho = 1/N$  and  $P(t) = 1 - e^{-ut}$ , independent of  $r$

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**Tumor suppressor genes (TSGs)**

- Somatic mutations in TSGs are recessive: inactivation of one allele is (nearly) neutral, while inactivating the second allele confers a fitness advantage. TSGs are inactivated by
  - two point mutations
  - one point mutation followed by loss of heterozygosity (LOH).

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**TSG inactivation in small populations: two hits**

frequency

time

2 hits

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**TSG inactivation in intermediate populations: one hit ("stochastic tunneling")**

frequency

time

1 hit

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**TSG inactivation in large populations: two hits (not rate limiting)**

frequency

time

2 hits

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**Summary: three dynamic laws for TSG inactivation**

$\text{Log } T_{1/2}$

Time until 50% chance

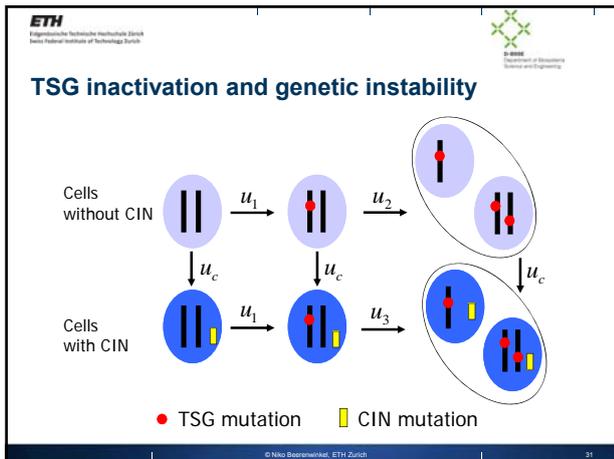
$\text{Log } N$   
Population size

2 hits

1 hit

0 hits (2 non-limiting)

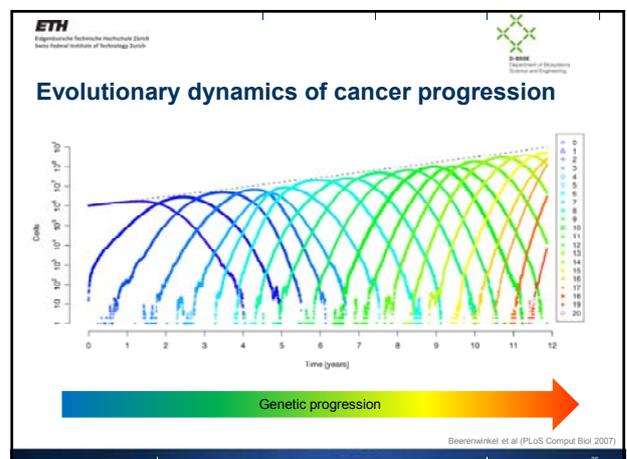
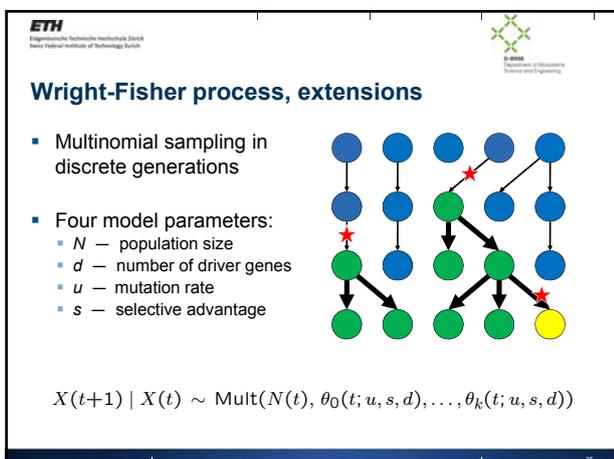
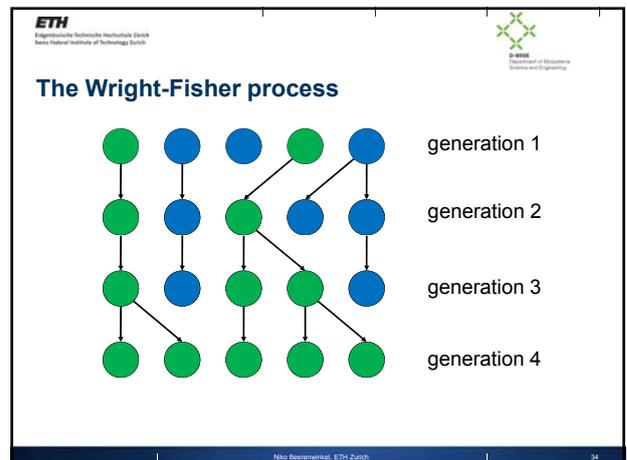
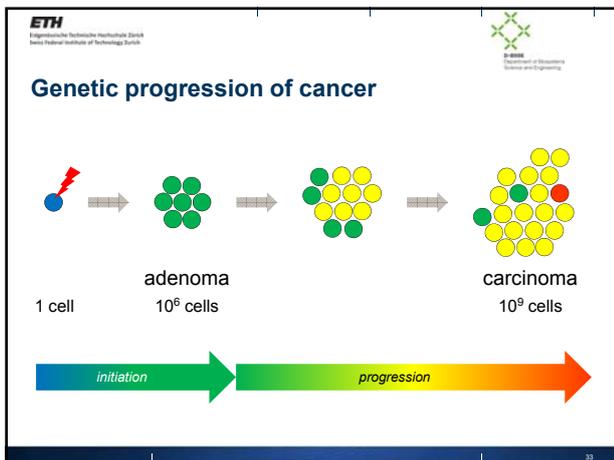
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**Cancer progression**

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### The waiting time to cancer

- How long does it take until the first cell with any 20 out of 100 mutations occurs?

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### The speed of adaptation

- The time until the first cell with  $k$  mutations appears is

$$\tau_k \approx \frac{k \log^2 [s/(ud)]}{2s \log N}$$

- Thus, the waiting time to cancer is dominated by the selective advantage  $s$ .

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### The probability of developing cancer

- normal mutation rate
- genetic instability

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### Accumulation of driver and passenger mutations

- Branching process model:

- Data:

$$n = \frac{v}{2s} \log \frac{4ks^2}{u^2} \log k$$

Bozic et al (PNAS 2010) 40

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### Intra-tumor genetic diversity and treatment

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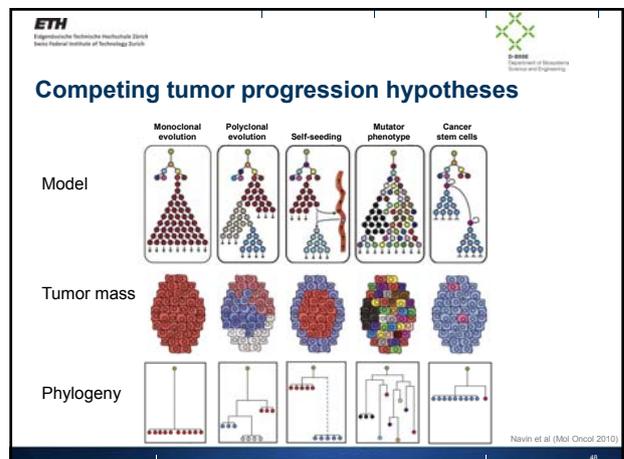
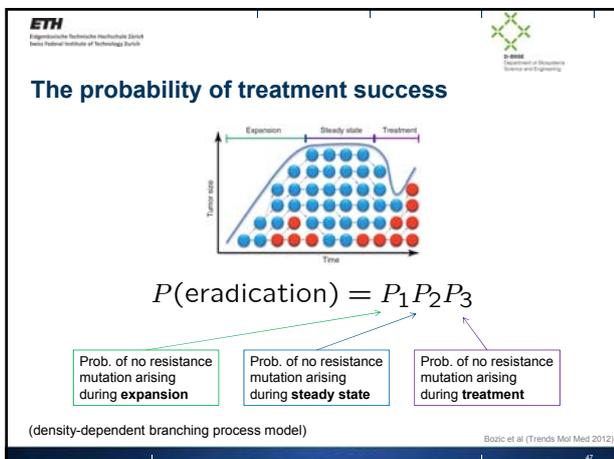
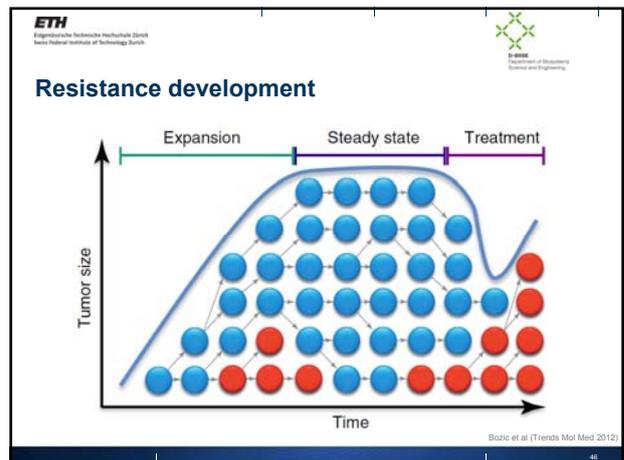
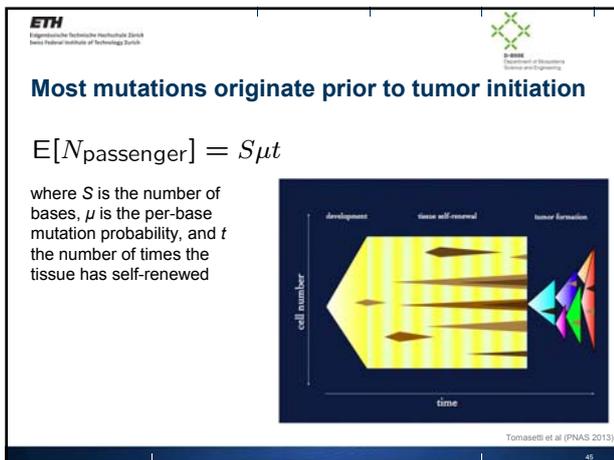
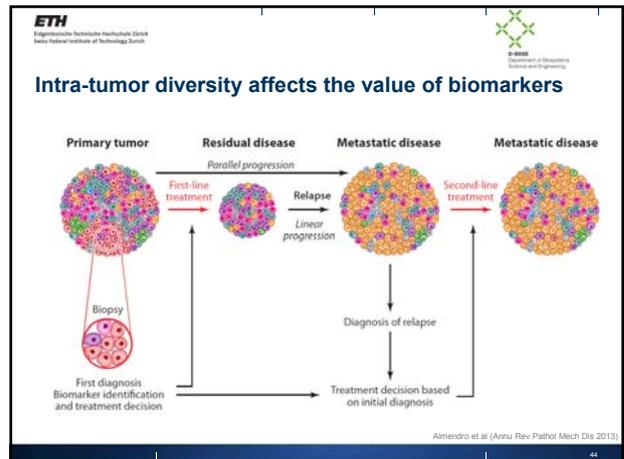
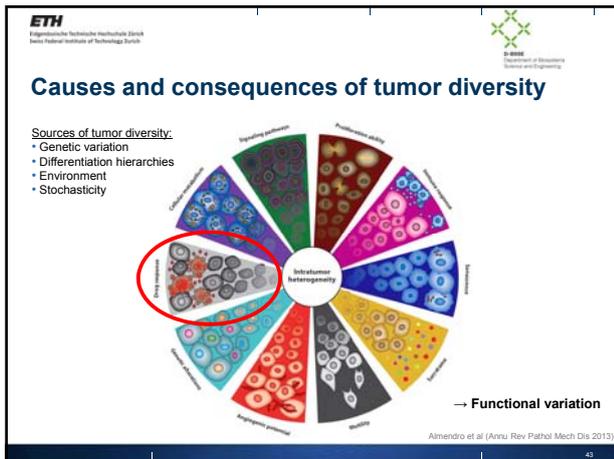
### Intra-tumor diversity itself has prognostic value

**Diagnostics**

**Prognostics**

→ Personalized medicine

Maruyak et al (Nat Rev Cancer 2012), Maley et al (Nat Genet 2006) 42



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**Strategies for intra-tumor sampling**

- Deconvolution of pooled sample
- Laser capture microdissection
- Cell sorting
- Single-cell analysis

- Tree reconstruction methods:
  - Rearrangement phylogeny (Greenman et al., 2011)
  - TuMult (Letouze et al., 2010)
  - MEDICC (Schwarz et al.)

→ Roland Schwarz' talk!

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**Sector Ploidy Profiling**

Correlation among copy number profiles

NJ tree

Navin et al (Genome Res 2010)

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**Estimating intra-tumor diversity from next-generation sequencing data**

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**Wright-Fisher model: Expected mutant distribution**

- Include neutral mutations (passengers)
- Trace individual genotypes

→ Most genetic variants occur at low frequencies

Frequency

Clone size

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**Next-generation ultra-deep sequencing**

Mixed sample

deep sequencing

Aligned reads

- Single nucleotide variants
- Sequencing error

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**Calling single-nucleotide variants (SNVs)**

Mixed sample

deep sequencing

Aligned reads

- Single nucleotide variants
- Sequencing error

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## Challenges in NGS-based diversity estimation

1. Most SNVs are expected to occur at low frequencies
2. Sample processing and sequencing errors are not uniform
3. Need to test many positions

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## deepSNV: Likelihood ratio test

Test mixed population  
Control Clonal

Ultra-deep sequencing

Position  $i$  Reference

Strand  $\beta = 1$   
 $\beta = 0$

SNVs  
Error

Aligned reads

Mean rate (mu) + SNVs  
Coverage  
Binomial nucleotide counts  
Dispersion =

Mean rate  
Coverage  
Binomial nucleotide counts  
Dispersion =

deepSNV algorithm

1. Test for each strand & position  $i$  and nucleotide  $b$   
Null hypothesis: no snv  
 $P_{i,b} = \binom{C}{k} p^k (1-p)^{C-k}$   
Alternative: SNV A, frequency?  $P_{i,b} = \binom{C}{k} p_A^k (1-p_A)^{C-k}$
2. Combine P-values from each strand
3. Adjust P-values for multiple testing

$$-2 \log \frac{L(X_{s,i,b}, Y_{s,i,b} | H_0)}{L(X_{s,i,b}, Y_{s,i,b} | H_1)} \sim \chi_1^2$$

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## Overdispersion

phiX replicates

$\alpha = 1000$

tumor vs. normal

$\alpha = 137$

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## Application: Renal cell carcinoma

1x multiple lesions  
Primary 1  
Primary 2  
Metastasis

3x tumor-normal:  
Tumor 1  
Tumor 2  
Tumor 3

TP53

SNV Frequency

Position

VHL

SNV Frequency

Position

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## Loss of heterozygosity

- 14 hemizygous SNPs
- Allele frequency ratio  $r = f_A / f_a \approx 2$  on chr. 3

$f_A = 7/10$   $f_a = 3/10$   $\rho = 4/7$

$$\rho = \frac{r/r_0 - 1}{r/r_0 + n - 1}$$

43% cancer cells in all tumor samples!

Fraction of cells with LOH

Genomic position [Chr3]

Primary 1  
Primary 2  
Metastasis

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## Evolutionary history

Seven subclonal mutations

VHL p.E189\*

-3p

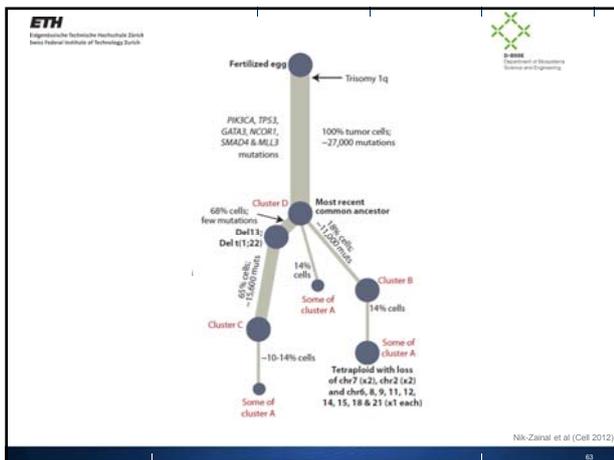
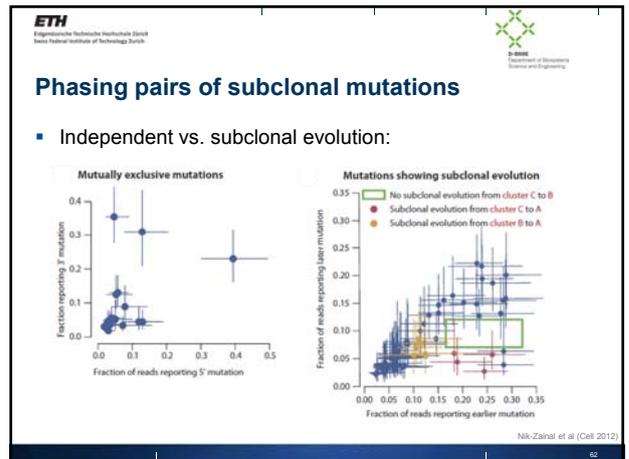
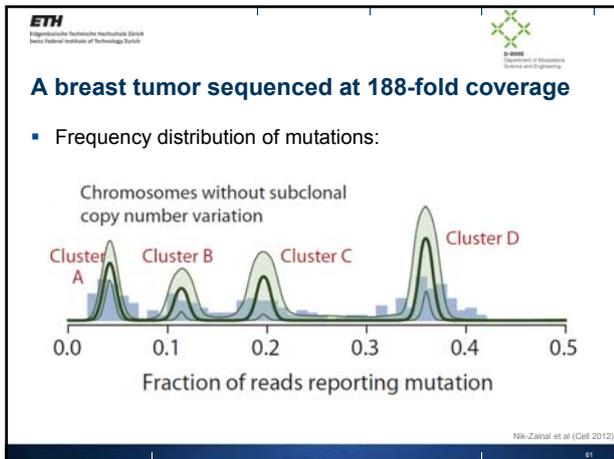
Normal

Frequency

Time

Diagnosis

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**Dependencies among clonal mutations (order constraints)**

