











Automatic Characterization of the Cell Organization in Light Microscopic Images of Wood

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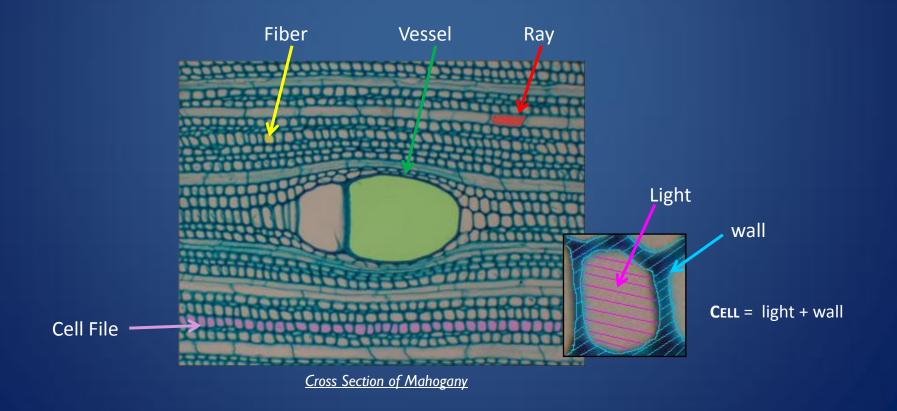
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Biology

- Environmental modifications on the functioning plasticity or production of plants
- Cell organization studies

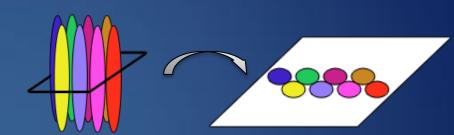


Preparation and digitization

- Vibratome¹; Sections of 20-25μm
- Coloured; methylene blue, safranine...
- Digitized; camera and microscope²
- Resolution ; 1600x1400 pixels
- Pixels size :



1; Vibratome Series 1000 Sectioning System

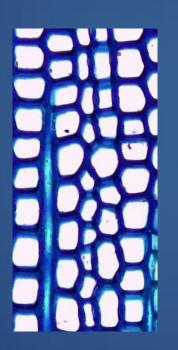


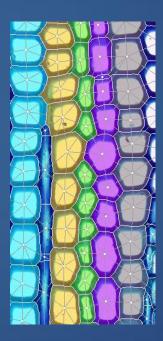


2 ; Olympus DP71 LCD camera mounted on a Olympus BX51 microscope

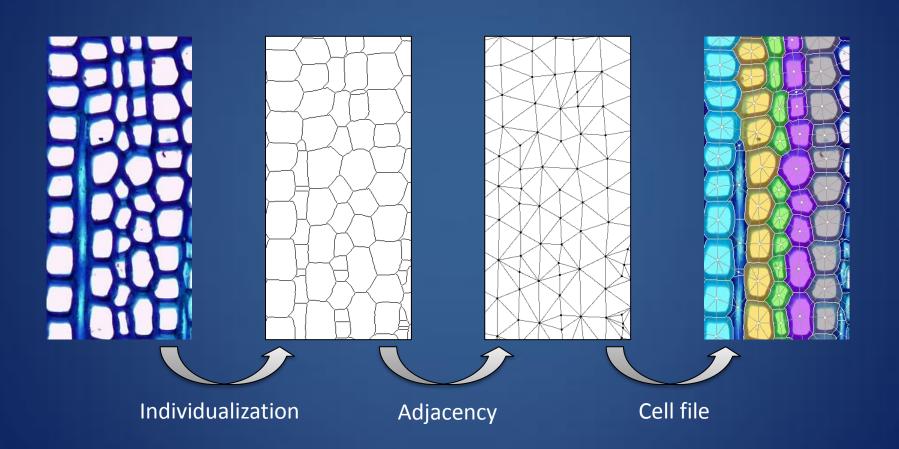
Method: Concepts

- Stability
- Similarity
- Validity



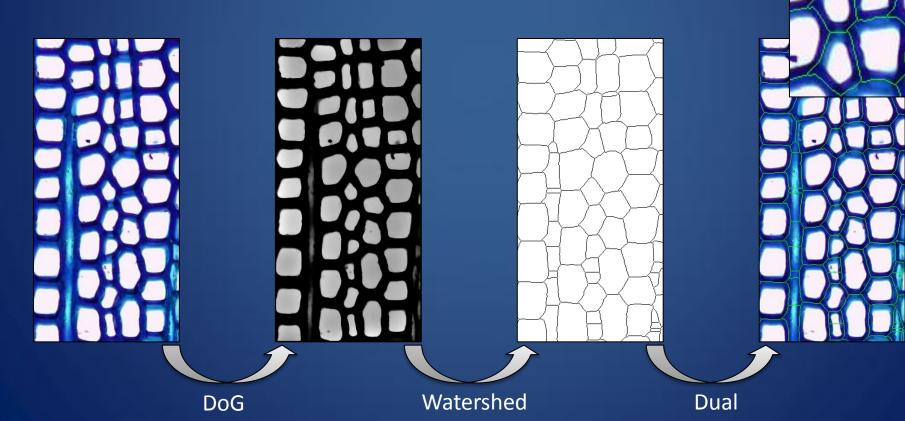


Method: Overview



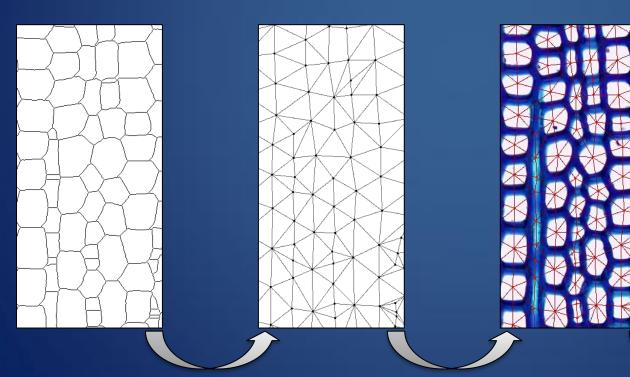
Method: Individualization

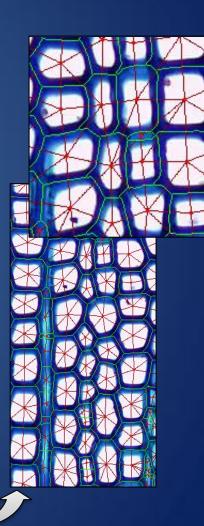
- Increase the contrast
- Segmentation
- Geometric modeling



Method: Adjacency

- Topological modeling
- Connections between neighboring cells





Graph

Dual

Graph & Watershed lines

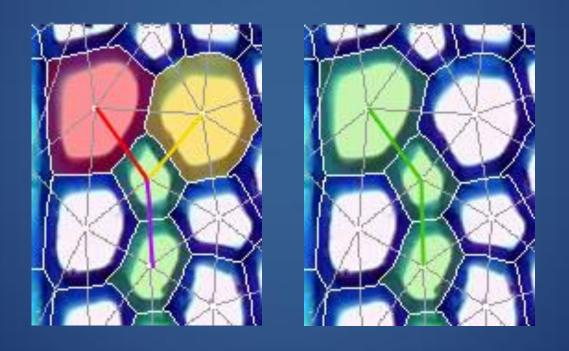
Method: Alignement Principle



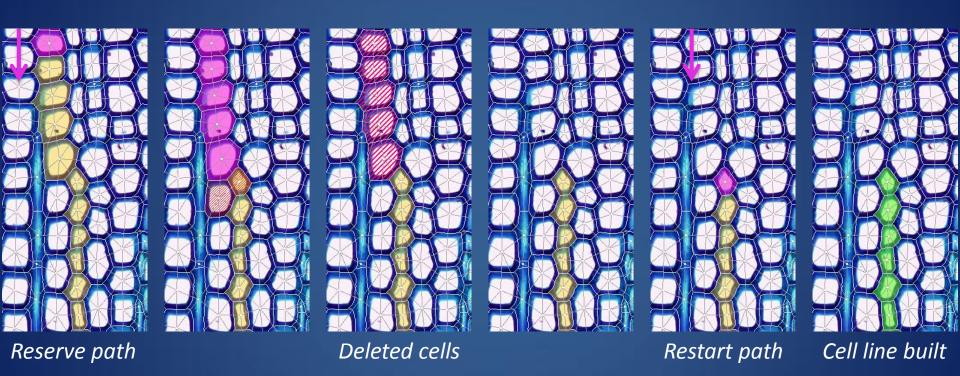
Path in the direction of the cell file

How find the better neighbor?

- Angle deviation
- Similarity Bray Curtis criterion

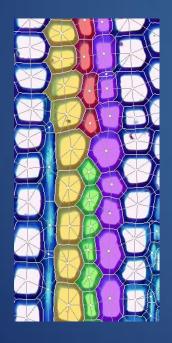


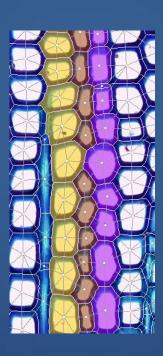
Method: Feedback check principle



Method: Merge

- Concatenate several part of line
- Topological rules





Section: S1 and S2

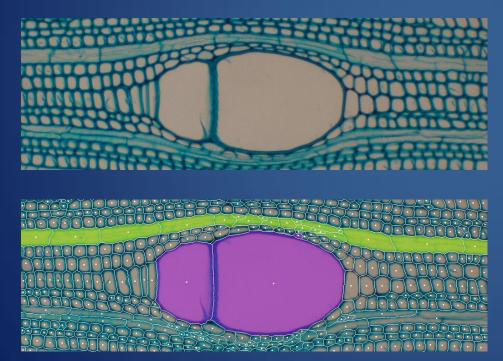
File: F3 and F4

[F3;F4] O S1 [F3;F4] O S2 F1,2 = S1 U S2

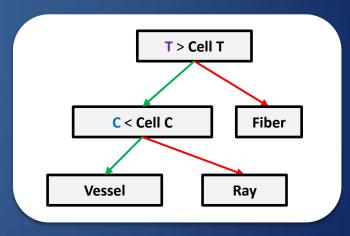
New File: F1,2

Method: Classification

- Classify different cells
- Geometric and densitometric characterization



Top, Cross Section of Picnanthus. Down, Classification Vessels et Rays



Decision Tree. T represents the threshold of the perimeter and C the threshold of circularity

Score

$$Score_f = file(length, State ...) + rac{\sum_k^p Cell(Blur_k, Size_k ...)}{p}$$

$$Methods fiability \qquad \qquad Cells quality$$

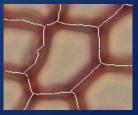
$$size_k \begin{cases} If S_k < threshold; t = \frac{S_k}{threshold} \\ Else t = 1 \end{cases}$$

$$length egin{cases} If NbCell < threshold; l = rac{NbCell}{threshold} \ Else \ l = 1 \end{cases}$$

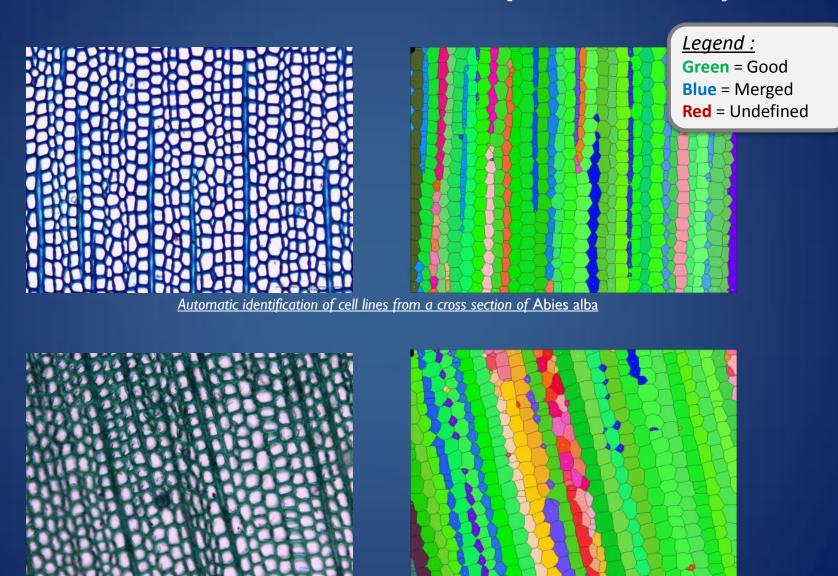
$$state \begin{cases} 1 \ \textit{if direct} \\ 0.75 \ \textit{if reconstruct} \\ 0.5 \ \textit{if isolated} \end{cases}$$

$$blur_k = 1 - rac{\sigma_k - \sigma_{min}}{\sigma_{max}}$$



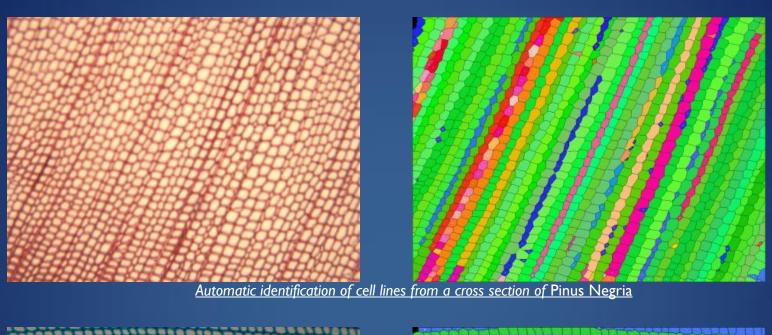


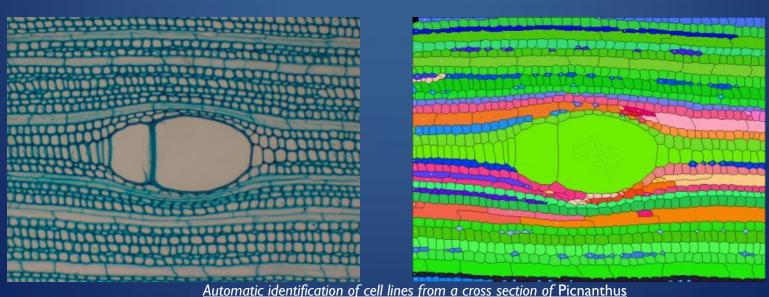
Experimental results: simple examples



Automatic identification of cell lines from a cross section of Pinus Caribensis

Experimental results: more difficult





Result

Summary of some significant results

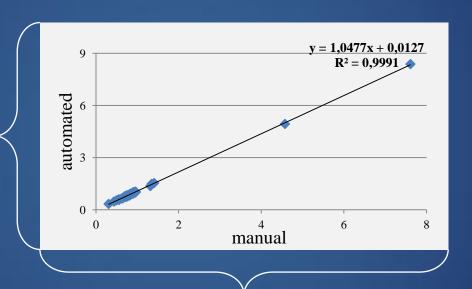
Species	Size (pixels)	Cells numbers	Times (sec)	Total Quality (%)
Mahogany	1024x768	1359	14.3	83
Fir	1360x1024	800	12.4	92
Black Pine 1	1600x1200	1873	23.2	73
Caribean Pine	1360x1024	828	11.5	91
Black Pine 2	1600x1200	1458	16.1	93

Summary of some significant results: the size of images processed, the number of cells extracted, the CPU time obtained on a machine with an Intel Xeon at 2.3GHz

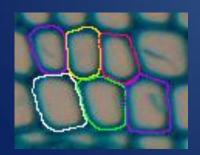
Experimental results: accuracy

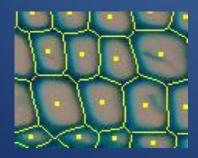
- Strongly correlated areas
- Weak over-valuation of the automated method
- Medium error of 5%

normalized
automated areas



normalized expert areas





Experimental results: sharpness studies

- Strongly correlated areas
- Weak under-valuation of the automated method
- Medium error of 2.8%

numerical detection on image blurred

y = 0,9963x + 0,0037

R² = 0,998

1.5

0.7

0.3

0.3

0.5

0.7

0.9

1.1

1.1

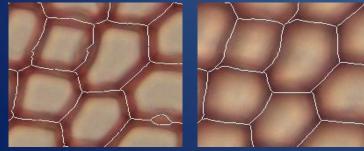
1.1

1.3

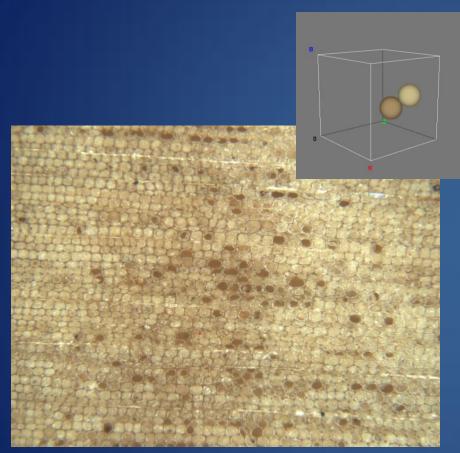
1.5

1.7

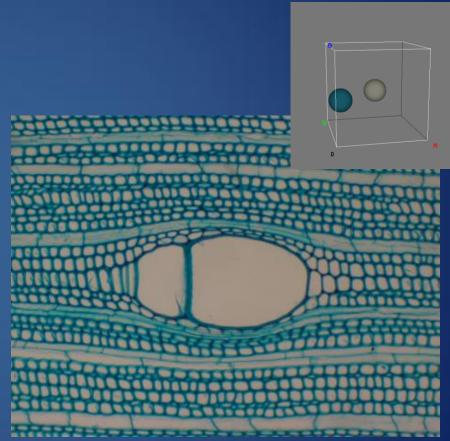
numerical detection on image sharp



Limits: segmentation



Cross section of Pinus Negria sanded



Cross section of Picnanthus colored with toluidine blue

Conclusion

Fully-automated method to characterize the cell organization

Self conditioned algorithm

Morphological and Topological analysis

Thanks for your attention