

# *in silico* strategies for target discovery

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A **target** is something you aim at.

In the context of a pathology, a **target** can be

- a **biological entity** ← **Ontology**
- and/or a **biological phenomenon** ← **Functional representations**

**Intervention** on the target depends on

- the **level of knowledge of the disease**,
- the **cultural background of the physician**
  - **traditional medicine** will focus more on the symptoms in order to **end the symptoms**.
  - **allopathic** (western, orthodox) **medicine** will focus on the causes (*i.e.* the **etiology**) in order to **oppose the causes**.
  - **homeopathic medicine** (in the sense of non-allopathic medicine) will focus on the etiology in order to **treat likes with likes**. In the widest sense of the word, vaccination is a category of homeopathic treatment.
- the **technics and technologies for the intervention**
  - **without exogenous agents**
    - musculoskeletal manipulations, surgery, diets
  - **with exogenous biochemicals**
    - **natural substances** based on **traditional pharmacopoeia**
    - **drugs** derived of traditional pharmacopoeia or from synthetic chemistry
    - **vaccines**
  - **with exogenous genes** (**gene therapy**)

# Malaria ?

*In silico* strategies for target discovery, should :

## 1) comply with knowledge of the disease in ETIOLOGIC terms

- **objects**, attributes and **data** that are **consistent with**
  - **biological entities** (as defined or understood by bench biologists in reductionist terms); i.e. genes, RNA, proteins, solutes, cellular structures, tissues, organs, etc.

**Ex.: a protein “E”**

- and their **measures**, scales, i.e. concentration
- **processes** and functions that represent biological operations
  - alterations over time (growth, transformations, movements) :

**Ex.: a reaction “A → B”**

- **relations** between objects and processes

**Ex.: “C” controls “E” that catalyzes “A → B”**

**If “B” is related to the pathology, “C” might be a target**

## 2) benefit from biological knowledge in widest terms

- **access** to all data, experiments, literature
- **mining, comparisons, knowledge transfers**

 3) include, enable, enhance, maximize creative approaches

 4) comply with feasibility of a treatment

- genetic variability of target? drug? vaccine? gene therapy?

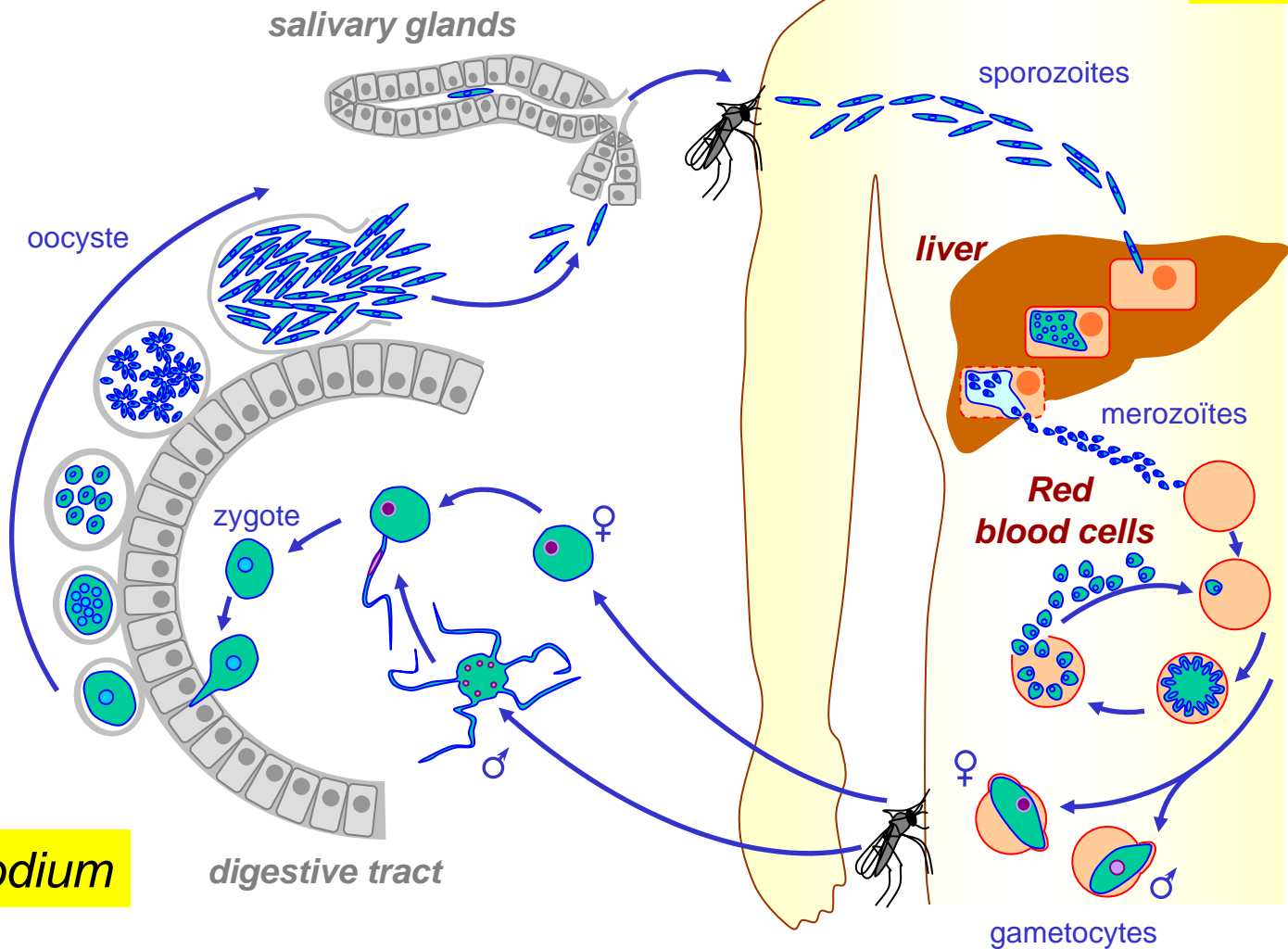
# Malaria

**Anopheles**

Sexual reproduction  
(*Anopheles* intestine)

Asexual multiplication  
(*Human red blood cells*)

**Human**



**Plasmodium**

# Malaria

Anopheles

FlyBase

Fly

Ensembl  
VectorBase

CyanoBase

Synechocystis

PlasmoDB  
ApiDB

Cryptosporidium

Plasmodium

Integr8  
UniProt

Bacillus

E. coli

Arabidopsis

Euglena

Toxoplasma

Babesia

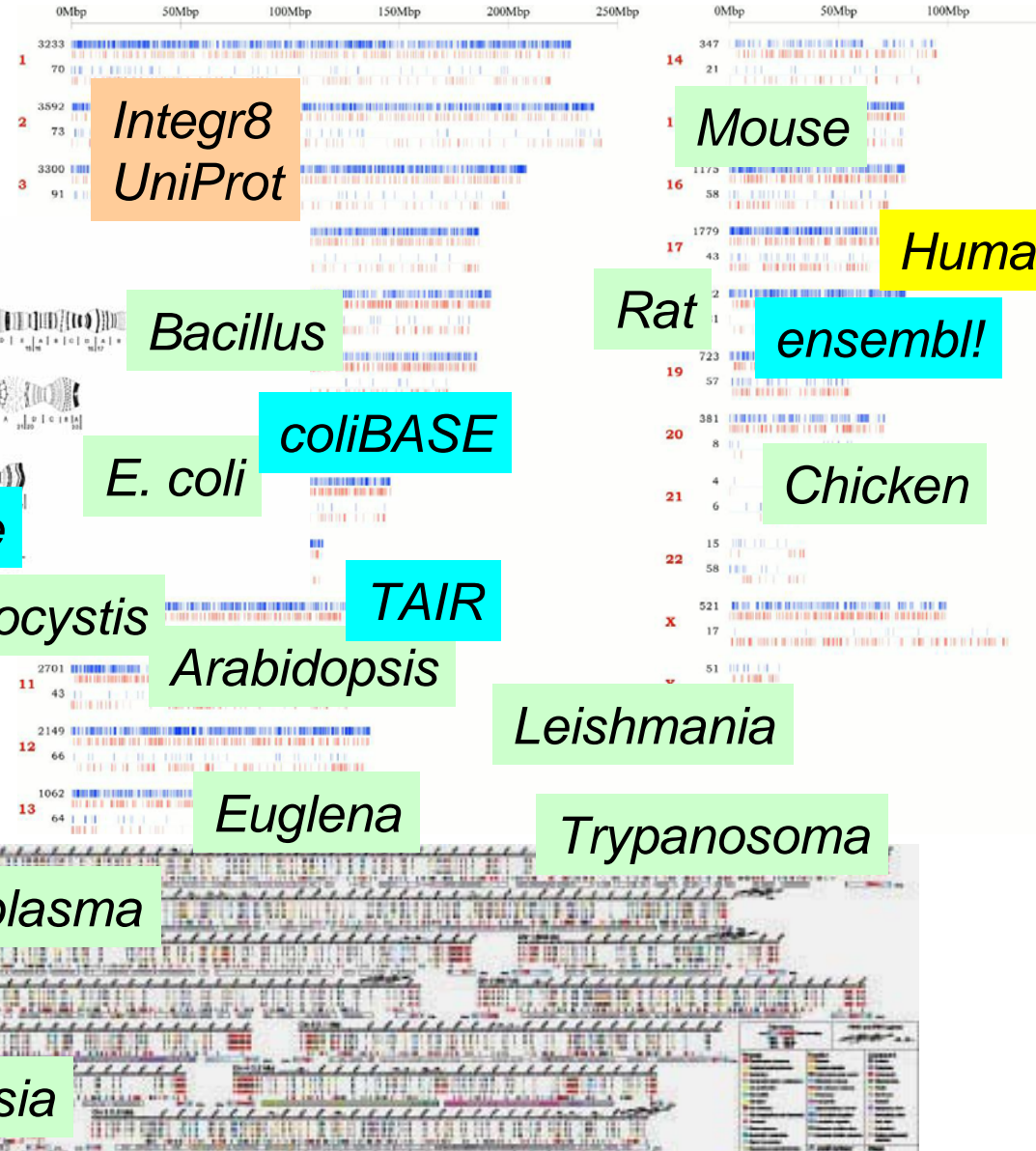
coliBASE

TAIR

Leishmania

Trypanosoma

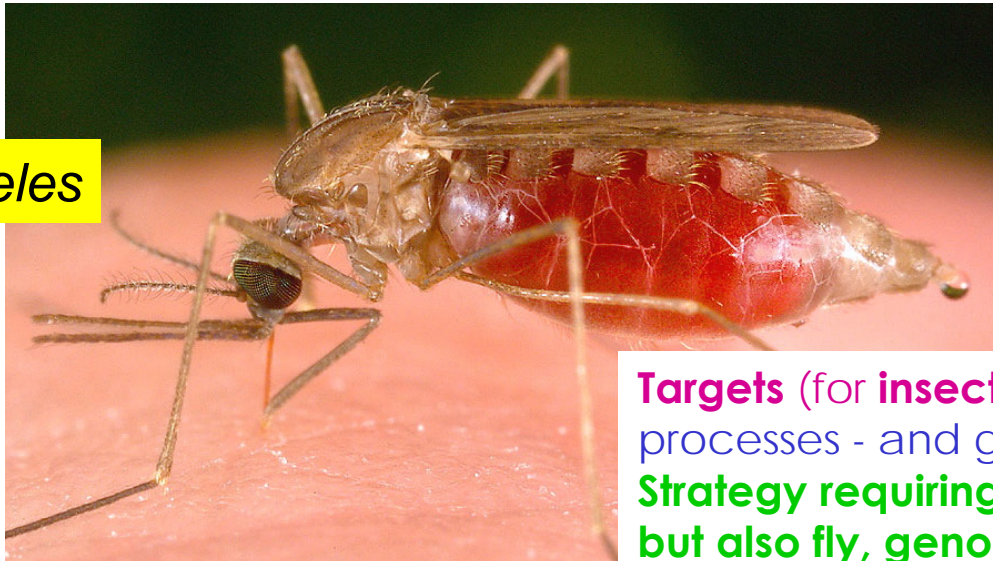
Desp. waiting for Google Bio-earth...





# Malaria

*Anopheles*



*Human*

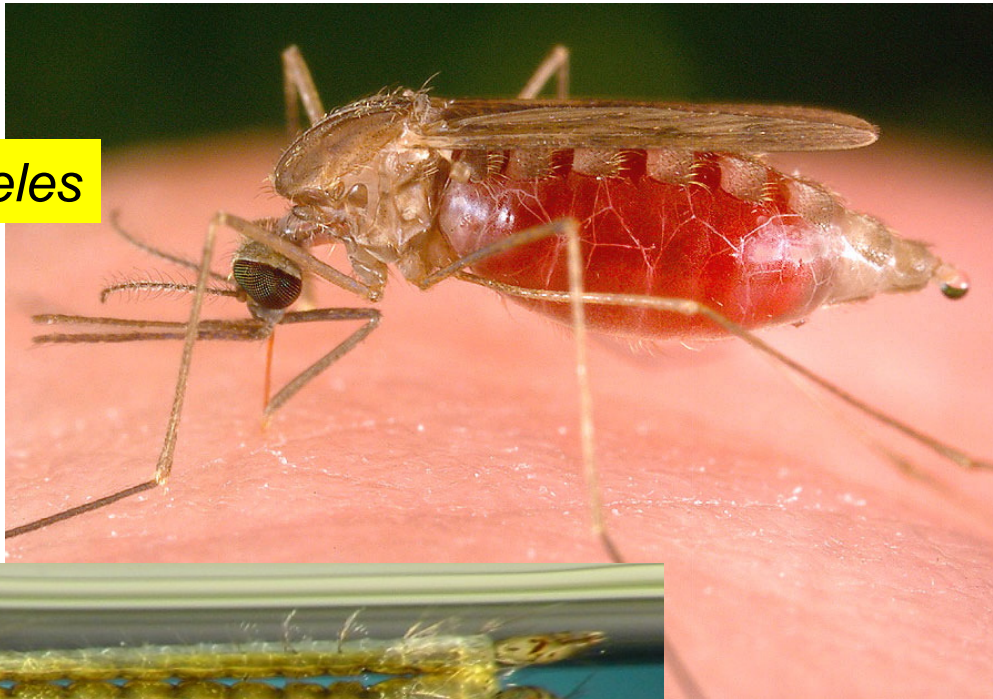
**Targets** (for **insecticides**): mostly vital processes - and genes -  
**Strategy requiring an exploration of mosquito, but also fly, genomic/postgenomic molecular, and functional data**



*Plasmodium*

# Malaria

*Anopheles*



Human



*Plasmodium*

**Targets** (for **vector resistance and population control**):  
Anopheles **genes introducing resistance to *Plasmodium***  
(with possible genetic manipulations to introduce this resistance) + **fitness genes** => promote the introduction of competing populations that resist to *Plasmodium*.  
**Strategy requiring an exploration of mosquito genomic/postgenomic molecular, and functional data + *Plasmodium* genomic/postgenomic data focusing on parasitic stages within mosquitoes.**

# Malaria

*Anopheles*

Targets ?

Human

*Plasmodium*





# Malaria

*Anopheles*

**Targets (for resistance):**

Human **genes introducing resistance to *Plasmodium*** (with possible genetic manipulations to introduce this resistance) => promote the human processes involved in resistance (with immune response enhancer, but also including gene therapy approaches on a long, long, long term...)

**Strategy requiring an exploration of human genomic/postgenomic molecular, and functional data + mouse/rat models requiring comparative tools (homologies, syntenies) + epidemiological information.**

Chromosome 5q31-q33

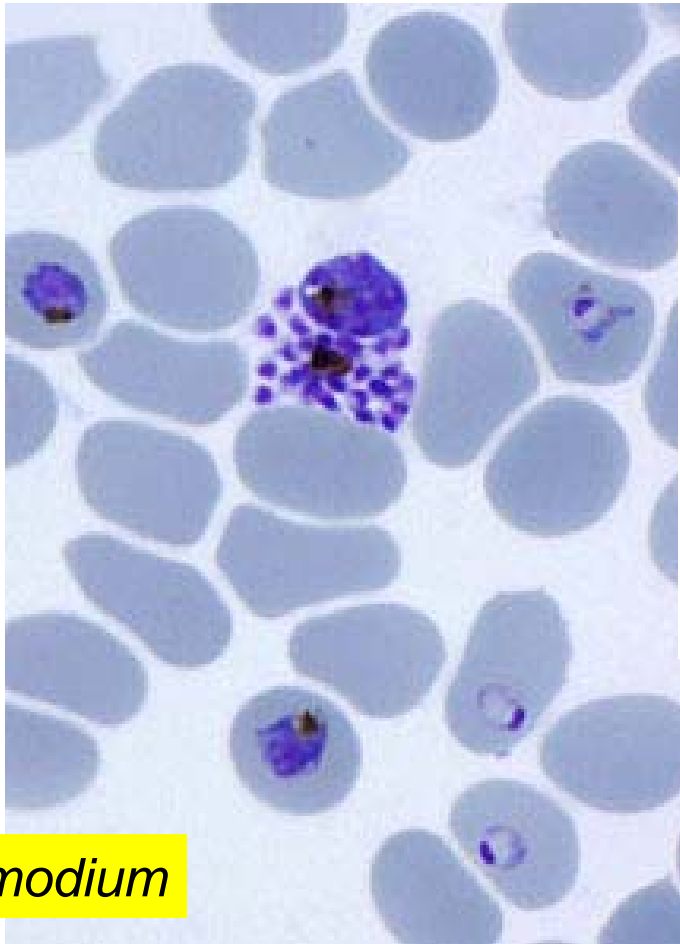
*Plasmodium*



Human

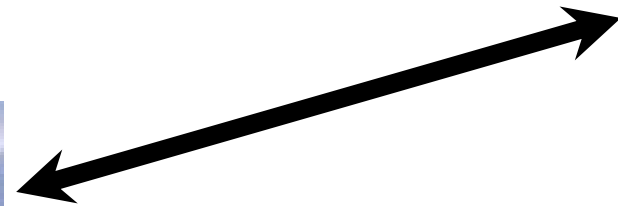
# Malaria

*Anopheles*



*Plasmodium*

Human



**Targets** (for **drugs/vaccines**): mostly vital processes - and genes -  
**Strategy** requiring an exploration of *Plasmodium* (*P. falciparum* and other malaria species), genomic/postgenomic molecular, and functional data + a study of genetic variations of the target candidate + a sufficient difference with human features to avoid toxicity.

# Malaria

## Anopheles

**Targets** (for **insecticides**): mostly vital processes - and genes -  
**Mosquito + fly, genomic/postgenomic molecular, and functional data**

## Human

**Targets** (for **vector resistance and population control**): Anopheles **genes** introducing resistance to *Plasmodium* + **fitness genes**

**Mosquito genomic/postgenomic molecular, and functional data + Plasmodium genomic/postgenomic data focusing on parasitic stages within mosquitoes + population genetic data**

- + Experiments for Target validation (can be knock out but not only)
- + Experiment to develop treatment (drug discovery + development; vaccine development)

**Targets** (for **resistance**):  
Human **genes** introducing resistance to *Plasmodium*  
**Human genomic/postgenomic molecular, and functional data + mouse/rat models requiring comparative tools (homologies, syntenies) + epidemiological information.**

## Plasmodium

**Targets** (for **drugs/vaccines**): mostly vital processes - and genes -  
**Plasmodium (*P. falciparum* and other malaria species), genomic/postgenomic molecular, and functional data + human genomic/postgenomic data + a study of genetic variations of the target candidate**

# Malaria

*Anopheles*

**Targets (for insecticides):** mostly vital processes - and genes -  
**Mosquito + fly, genomic/postgenomic molecular, and functional data**

**Targets (for vector resistance and population control):** *Anopheles* g introducing resistance to *Plasmodium* fitness genes

**Mosquito genomic/postgenomic molecular, and functional data - *Plasmodium* genomic/postgenomic focusing on parasitic stages with mosquitoes + population genetic**

- + Experiments for Target validation (knock out but not only)
- + Experiment to develop treatment discovery + development; vaccine development)

*Plasmodium*

**Targets** genes -

***Plasmodium* (*P. falciparum* and other malaria species), genomic/postgenomic molecular, and functional data + human genomic/postgenomic data + a study of genetic variations of the target candidate**

**Invaluable** collaborative work to generate/collect/control data and put some order in this avalanche of information. Endorsed by all individual scientists + dedicated teams having sufficient critical masses. No cost evaluation (gigantic); access considered as a « universal human right »; only limit is the investment in the maintenance, improvement and development of wise data storage models and systems, navigation maps and mining tools (DATABASES, KNOWLEDGE BASES, KNOWLEDGE SPACES) and accesses (internet portals and grid distribution)

*Human*

**e):** **ucing resistance**

**stgenomic**  
**tional data +**  
**quiring**  
**omologies,**  
**iological**

esses - and

# Malaria

**Targets (for insecticides):** mostly vital processes - and genes -

**Mosquito + fly, genomic/postgenomic molecular, and functional data**

*Anopheles*

*Human*

**Targets (for vector resistance and population control):** *Anopheles* genes introducing resistance to *Plasmodium* + fitness genes

**Mosquito genomic/postgenomic molecular, and functional data + *Plasmodium* genomic/postgenomic data focusing on parasitic stages within mosquitoes + population genetic data**

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## Linear workflows of experiments

that can be stopped as soon as failure indicators appear. Cost evaluation includes the cost of failures (>70 % of drug discovery/development projects fail), *i.e.* 800 M€.

Endorsed by small teams.

Time scale is ~10 yr per project.

This is clearly the LIMITING STEP.

*In silico* information are non limiting by comparison.

ce to

information.

*Plasmodium*

**Targets (for drug genes -**

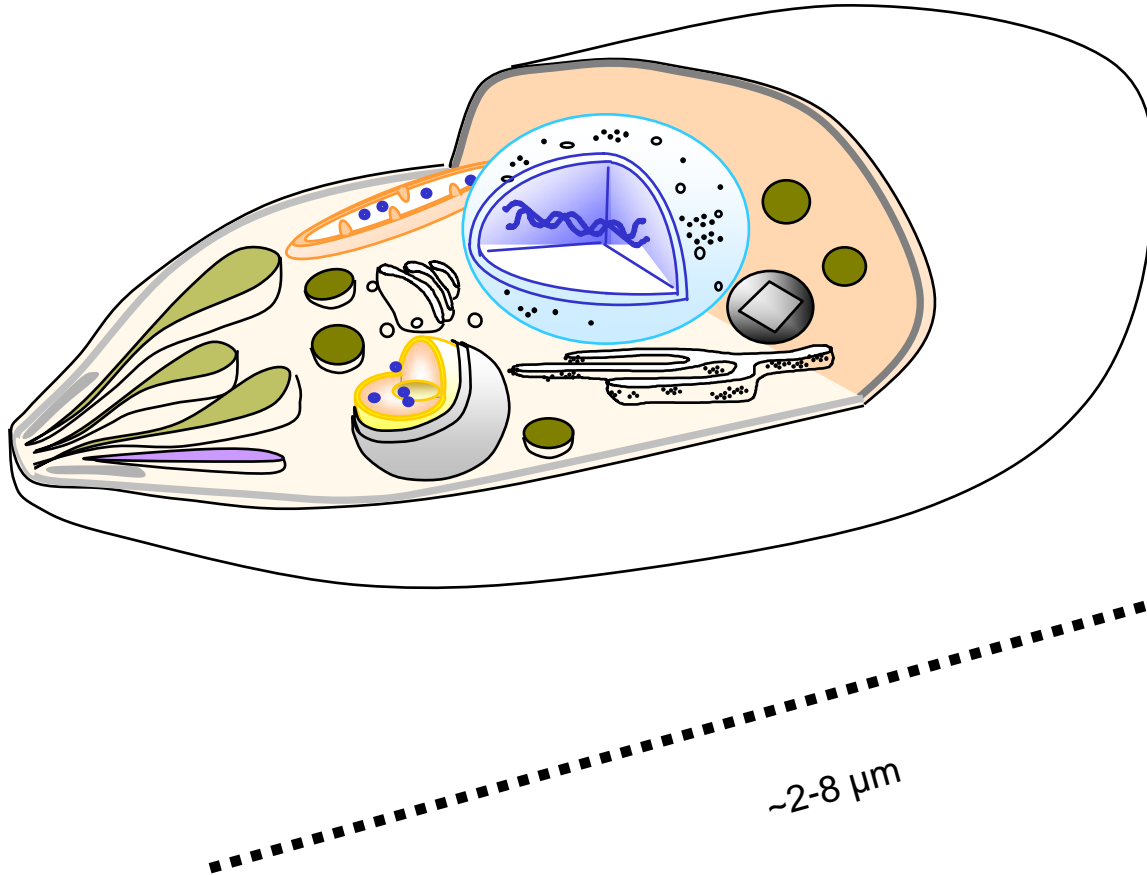
***Plasmodium* (genomic/postgenomic) + human genomic variations of the**





## Malaria targets for new drugs ?

An example of a drug discovery project,  
combining bioinformatics and experiments.



*Plasmodium falciparum* an apicomplexan

**Malaria targets for new drugs ?**  
**An example of a drug discovery project,**  
**combining bioinformatics and experiments.**

1991

*J. Protozool.* 1991 38:243-5.

The putative mitochondrial genome of *Plasmodium falciparum*.

**Feagin JE, Gardner MJ, Williamson DH, Wilson RJ.**

Intraerythrocytic stages of mammalian malarial parasites employ glycolysis for energy production but some aspects of mitochondrial function appear crucial to their survival since inhibitors of mitochondrial protein synthesis and electron transport have antimalarial effects. Investigations of the putative mitochondrial genome of *Plasmodium falciparum* have detected organellar rRNAs and tRNAs encoded by **a 35 kb circular DNA. Some features of the organization and sequence of the rRNA genes are reminiscent of chloroplast DNAs.** The 35 kb DNA also encodes open reading frames for proteins normally found in chloroplast but not mitochondrial genomes. An apparently unrelated 6 kb tandemly repeated element which encodes two mitochondrial protein coding genes and fragments of rRNA genes is also found in malarial parasites. The malarial mitochondrial genome thus appears quite unusual. Further investigations are expected to provide insights into the possible functional relationships between these molecules and perhaps their evolutionary history.

**Malaria targets for new drugs ?  
An example of a drug discovery project,  
combining bioinformatics and experiments.**

1996

*J. Mol. Biol.* (1996) **261**, 155–172

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**JMB**



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**Complete Gene Map of the Plastid-like DNA of the  
Malaria Parasite *Plasmodium falciparum***

**R. J. M. (Iain) Wilson\***, Paul W. Denny, Peter R. Preiser  
Kaveri Rangachari, Kate Roberts, Anjana Roy, Andrea Whyte  
Malcolm Strath, Daphne J. Moore, Peter W. Moore and  
Donald H. Williamson

*National Institute for Medical  
Research, Mill Hill, London  
NW7 1AA, UK*

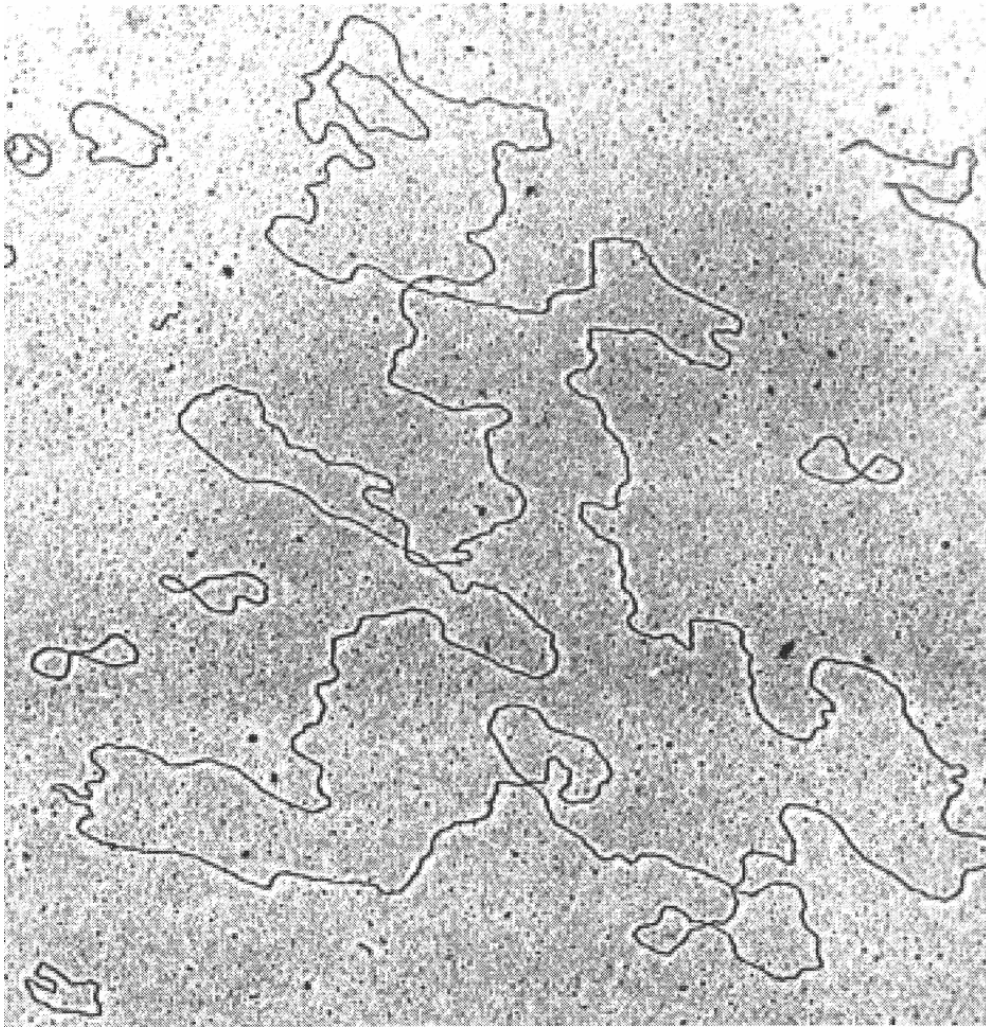
Malaria parasites, and other parasitic protists of the Phylum Apicomplexa, carry a plastid-like genome with greatly reduced sequence complexity. This 35 kb DNA circle resembles the plastid DNA of non-photosynthetic plants, encoding almost exclusively components involved in gene expression. The complete gene map described here includes genes for duplicated large and small subunit rRNAs, 25 species of tRNA, three subunits of a eubacterial RNA polymerase, 17 ribosomal proteins, and a translation elongation factor. In addition, it codes for an unusual member of the Clp family of chaperones, as well as an open reading frame of unknown function found in red algal plastids. Transcription is polycistronic. This plastid-like DNA molecule is conserved in several genera of apicomplexans and is conjectured to have been acquired by an early progenitor of the Phylum by secondary endosymbiosis. The function of the organelle (plastid) carrying this DNA remains obscure, but appears to be specified by genes transferred to the nucleus.



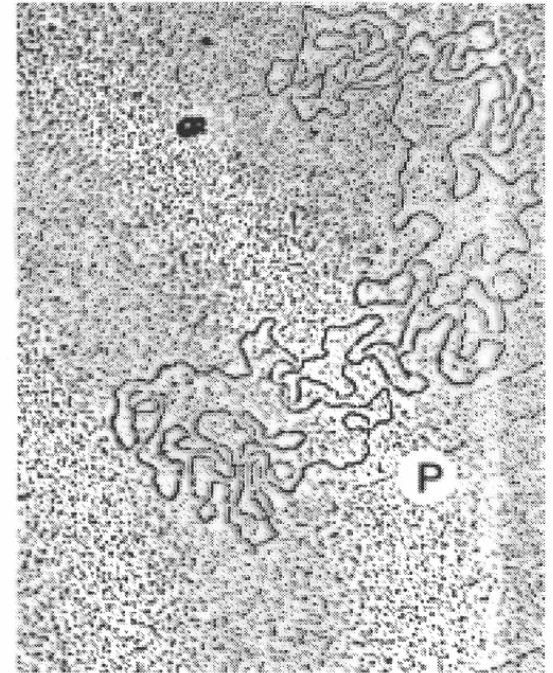
## Malaria targets for new drugs ?

An example of a drug discovery project,  
combining bioinformatics and experiments.

1997



Lettuce circular CpDNA ~140 kb  
(Kolodner and Tewari, 1975)



*Plasmodium berghei* ~35 kb  
(Yap et al., 1997)

<b>Synechocystis</b>	<b>3500 kb</b>
<b>Lettuce</b>	<b>140 kb</b>
<b>Plasmodium</b>	<b>35 kb</b>

## Malaria targets for new drugs ? An example of a drug discovery project, combining bioinformatics and experiments.

### A Plastid of Probable Green Algal Origin in Apicomplexan Parasites

Sabine Köhler,\*† Charles F. Delwiche,\*‡ Paul W. Denny,  
Lewis G. Tilney, Paul Webster, R. J. M. Wilson,  
Jeffrey D. Palmer, David S. Roos§

Protozoan parasites of the phylum Apicomplexa contain three genetic elements: the nuclear and mitochondrial genomes characteristic of virtually all eukaryotic cells and a 35-kilobase circular extrachromosomal DNA. In situ hybridization techniques were used to localize the 35-kilobase DNA of *Toxoplasma gondii* to a discrete organelle surrounded by four membranes. Phylogenetic analysis of the *tufA* gene encoded by the 35-kilobase genomes of coccidians *T. gondii* and *Eimeria tenella* and the malaria parasite *Plasmodium falciparum* grouped this organellar genome with cyanobacteria and plastids, showing consistent clustering with green algal plastids. Taken together, these observations indicate that the Apicomplexa acquired a plastid by secondary endosymbiosis, probably from a green alga.

Apicomplexan parasites contain two maternally inherited extrachromosomal DNA elements (1). The mitochondrial genome is a multicopy element of ~6 to 7 kb encod-

ing three proteins of the respiratory chain and extensively fragmented ribosomal RNAs (2). In addition, these parasites contain a 35-kb circular DNA molecule with no significant similarity to known mitochondrial genomes. The 35-kb element is similar to chloroplast genomes, containing an inverted repeat of ribosomal RNA genes and genes typically found in chloroplasts but not mitochondria (*rpoB/C*, *tufA*, and *clpC*) (3). The 35-kb DNA is also predicted to encode a complete set of tRNAs, numerous ribosomal proteins, and several unidentified open reading frames (3).

We used in situ hybridization to determine whether the 35-kb DNA is found within the parasite nucleus, mitochondrion, or cytoplasm or, alternatively, whether this

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S. Köhler, L. G. Tilney, D. S. Roos, Department of Biology, University of Pennsylvania, Philadelphia, PA 19104, USA.

C. F. Delwiche and J. D. Palmer, Department of Biology, Indiana University, Bloomington, IN 47405, USA.

P. W. Denny and R. J. M. Wilson, National Institute for Medical Research, Mill Hill, London NW7 1AA UK.

P. Webster, Department of Cell Biology, Yale University School of Medicine, New Haven, CT 06520, USA.

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\*These authors contributed equally to this work.

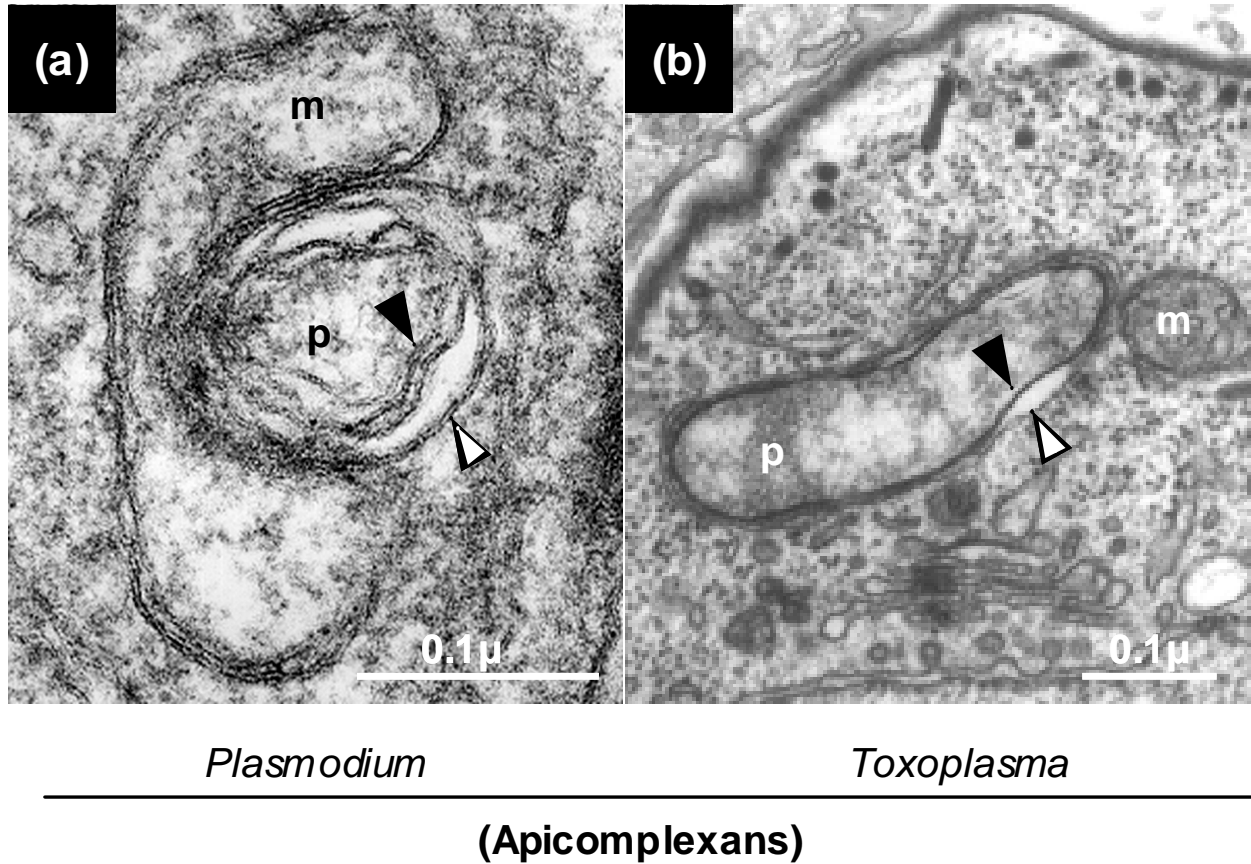
†Present address: School of Pharmacy, University of California, San Francisco, CA 94143, USA.

‡Present address: Department of Plant Biology, University of Maryland, College Park, MD 20742, USA.

§To whom correspondence should be addressed. E-mail: droos@sas.upenn.edu



**Malaria targets for new drugs ?**  
**An example of a drug discovery project,**  
**combining bioinformatics and experiments.**



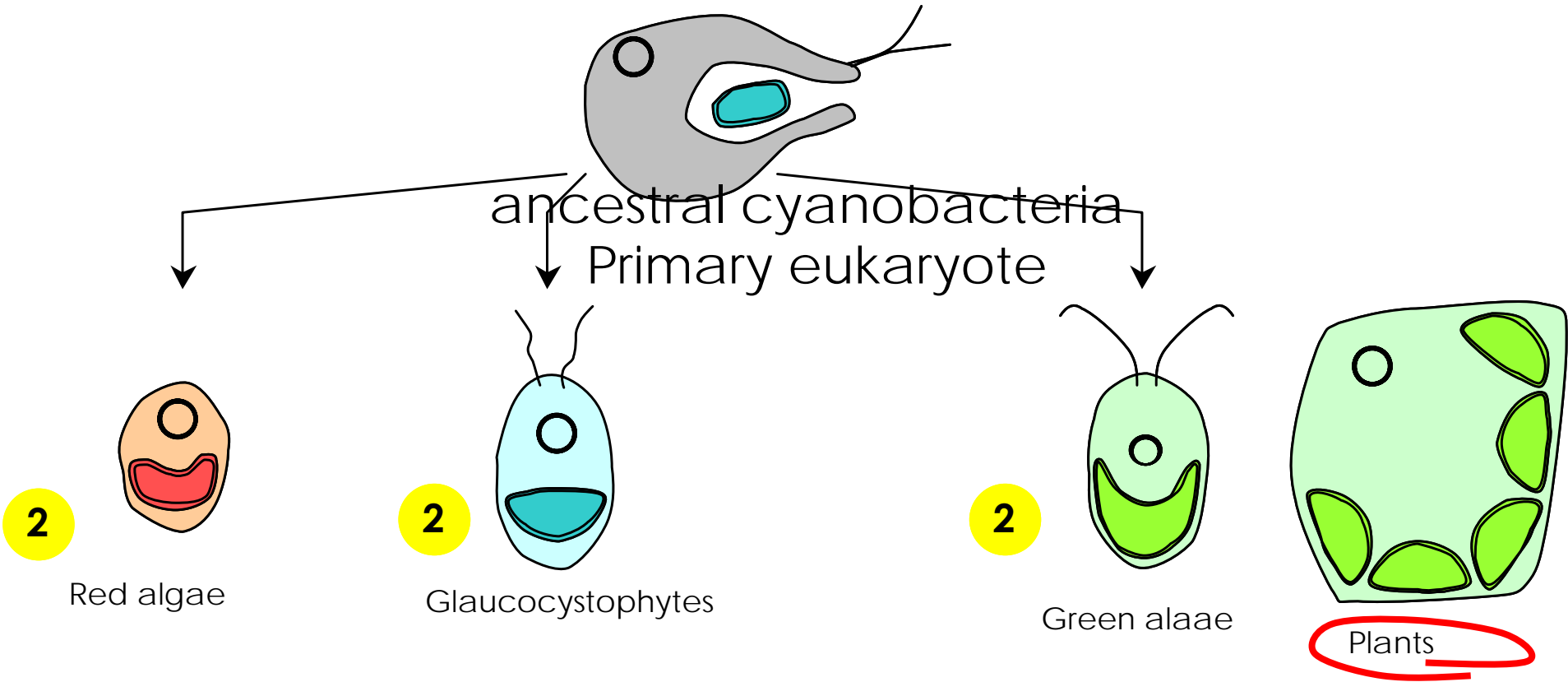
**The apicoplast, a plastid limited by 3-4 membranes**

**Malaria targets for new drugs ?**

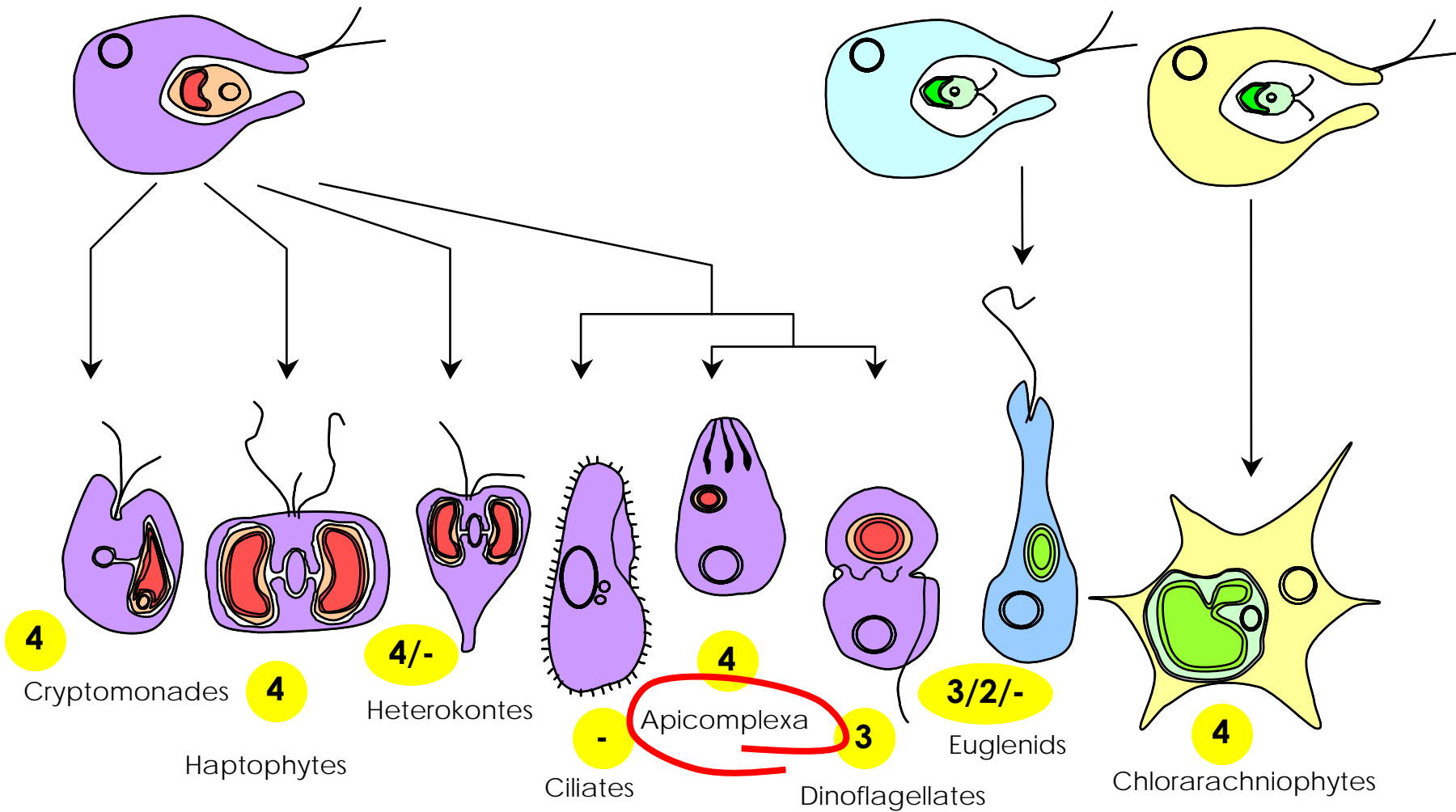
**An example of a drug discovery project,  
combining bioinformatics and experiments.**

**What is the apicoplast origin ?**

*primary endosymbiosis*



*secondary endosymbiosis*



The two contentious groups include parasites that cause diseases such as malaria and sleeping sickness; Adl hopes his classification will aid drug development. “Placing these organisms in the wrong group is in part responsible for the fact that we do not have specific drugs for these diseases, because of wrong assumptions about their biochemistry,” he says.

arily troubled, but the evidence was not clear enough for a consensus on a new regime, and fierce disagreements became common.

“I’ve seen people throw things at each other,” says Sina Adl, a soil-organism specialist at Dalhousie University in Halifax, Canada, who coordinated the group of 28 protist experts that produced the new classification. It was commissioned by the International Society of

trypanosomes (purple), which cause sleeping sickness, were one of the hardest protists to classify.

Protistologists and is published in *The Journal of Eukaryotic Microbiology* (S. M. Adl et al. *J. Eukaryot. Microbiol.* 52, 399–451; 2005).

The experts have given protists a mighty four kingdoms out of six. Animals do not even get their own group — fungi and animals have



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## Malaria targets for new drugs ?

An example of a drug discovery project,  
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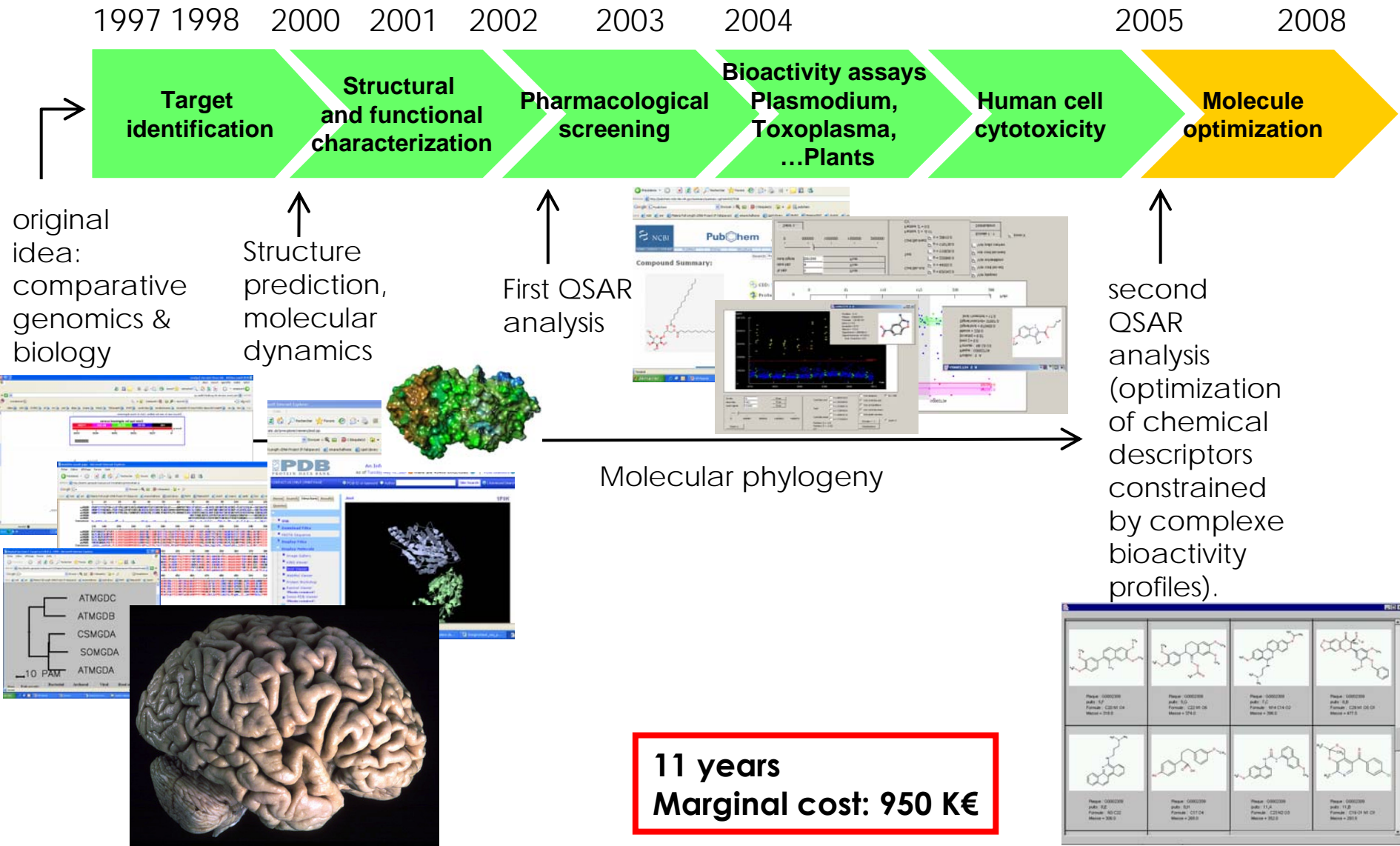
1999

### Is the apicoplast a target ?

- **Fosmidomycin**, a **herbicide** targeting a plant chloroplast process (*i.e.* the non-mevalonate isoprenoid pathway), kills the parasite (Jooma et al., 1999, *Science*).
- **Genetic impairment of apicoplast division** leads to cell with no apicoplast... that die (He et al., 2001, *EMBO J*)

# Malaria targets for new drugs ?

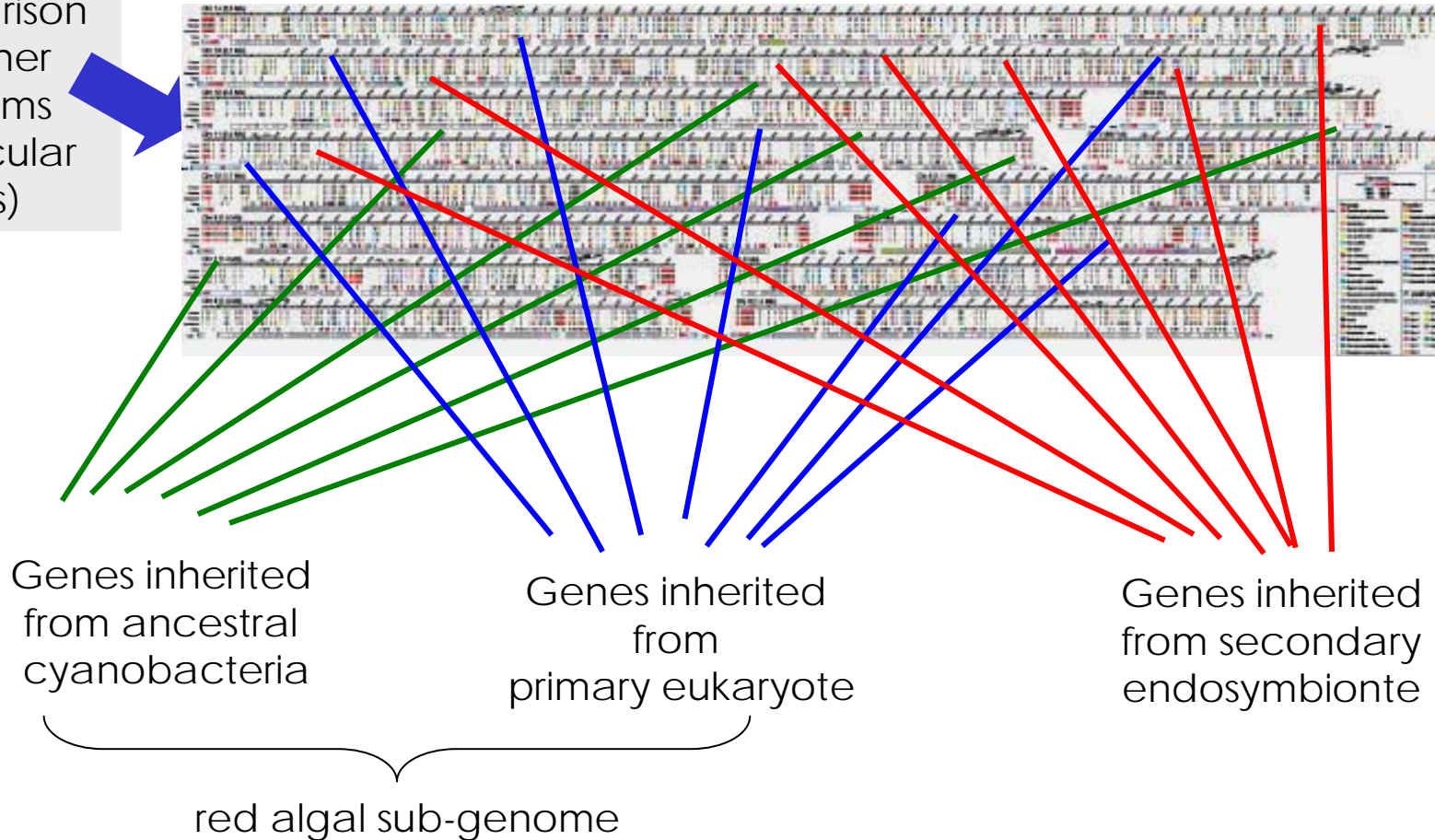
## An example of a drug discovery project, combining bioinformatics and experiments.



# Is the apicoplast a target ? (in more general term any plant-like process in Plasmodium)

Available molecular data  
PlasmoDB, etc.  
+ comparison with other organisms  
(in particular plants)

sequence comparisons



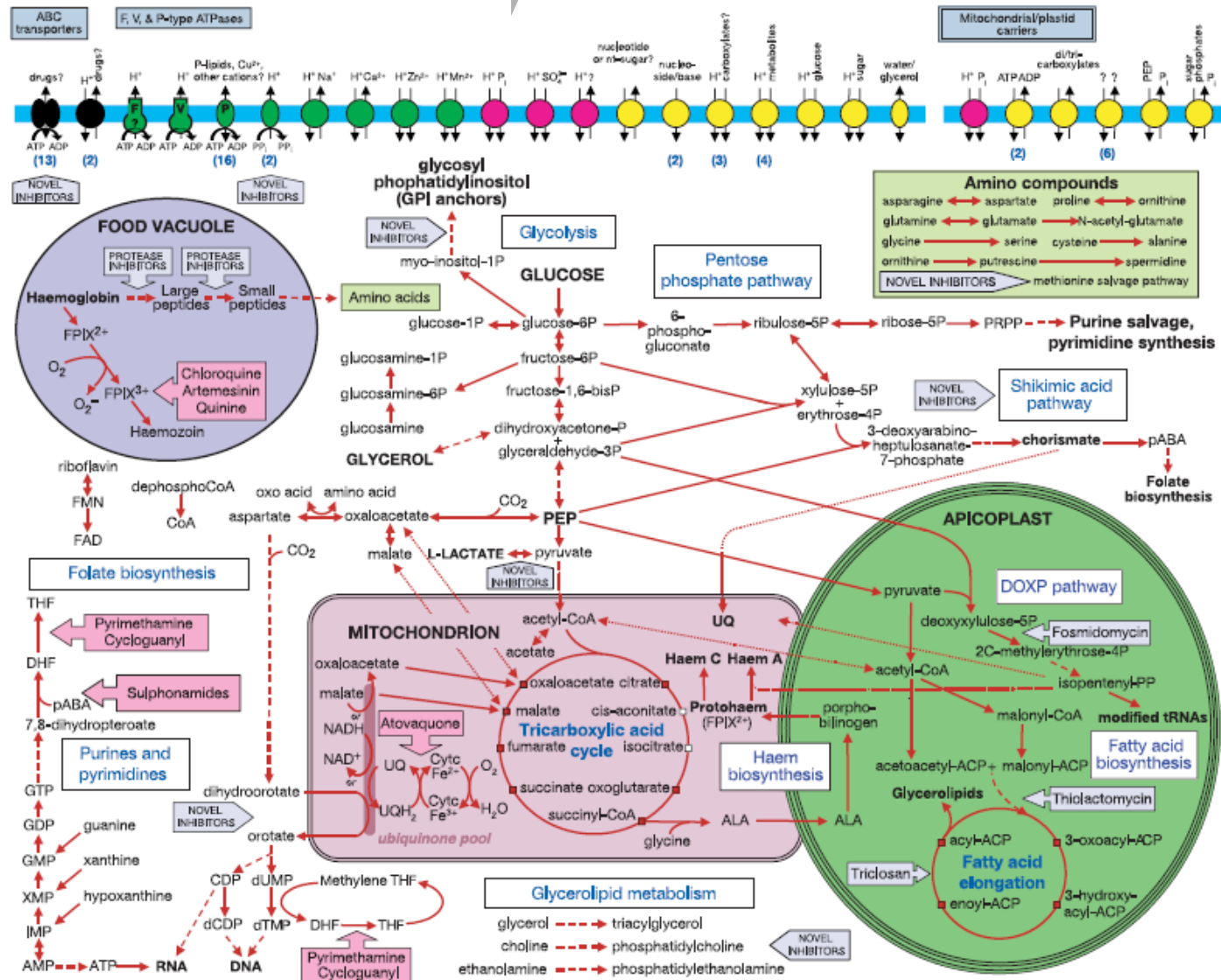
**Plant side ?**



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sequence comparisons  
analyses of biological processes

Target identification

Structural and functional characterization

Pharmacological screening

Bioactivity assays  
Plasmodium, Toxoplasma, ...Plants

Human cell cytotoxicity

Molecule optimization

Are there more we might have missed ?

# CURRENT LIMITS

Available molecular data  
PlasmoDB, etc.  
+ comparison with other organisms  
(in particular plants)



**sequence comparisons**  
**analyses of biological processes**

## Blast (+ motifs) searches

	Number of genes predicted to code for proteins (in original paper)	hit with genes with known function	hit with genes with no known function	no hit
--	--	------------------------------------	---------------------------------------	--------

**Human**  
(Craig et al., 2001)

~30,000



**Arabidopsis**  
(A.G.I., 2000)

~25,000



**Plasmodium**  
(Gardner et al., 2002)

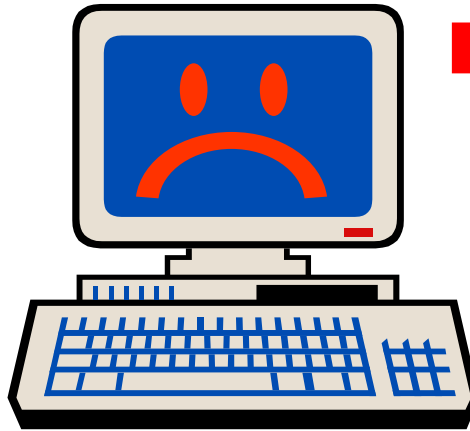
~5,000



**~3200 sequences**

# CURRENT LIMITS

Available molecular data  
PlasmoDB, etc.  
+ comparison with other organisms  
(in particular plants)



sequence comparisons

analyses of biological processes

## Blast (+ motifs) searches

```
>87088036|chr11_s259
      Length = 17,424
```

Plus Strand HSPs:

```
Score = 79 (32.9 bits), Expect = 4.1, Sum P(2) = 0.98
Identities = 23/76 (30%), Positives = 42/76 (55%), Frame = +2
```

```
Query:   441 IAGQEAGNVPYVIENGIG--KYLKSPKEIAKTVSQWFGPKANELQIMSQNALKHARPDV 498
          I G+  G++  ++ N +   KY KSP  + KTVS  FG ++ E  ++      +H + +
Sbjct:  2423 INGNMGLDQSM LNNLLNSEKYKSPYTLDKTVSSGFGSRSKESMLLEY---QH KSEQ- 2590
```

```
Query:   499 FKIVHDLDELV-RQKI 513
          +I+ +L EL+ + KI
Sbjct:  2591 -EILRELQELINKKI 2635
```

```
Score = 54 (24.1 bits), Expect = 4.1, Sum P(2) = 0.98
Identities = 21/95 (22%), Positives = 45/95 (47%), Frame = +2
```

```
Query:   27 NSSLHGNNNGYSSFFSSNSVHFGGLATQNRKFKVNSLSFSKEGSNLKRILSDFNRVIRL- 85
          N++ + NN+N  SS +++S +   ++ N      N+ + +K  + +++ + F+ +
Sbjct:  1745 NNNNNNNNNNNSSNNSSNNNNSSNNNNSSNNNNNNNAKAAANPMEQLTLF SHINND 1924
```

```
Query:   86 HCDRI----PLGFSSIGLNSGESNGVSDNGHGVLE 116
          H D      PL      G +NG +  G+  LE
Sbjct:  1925 HHDEKGDGHPLEHIMLFGQHNGDAKGGNNPLE 2029
```

~3200 sequences

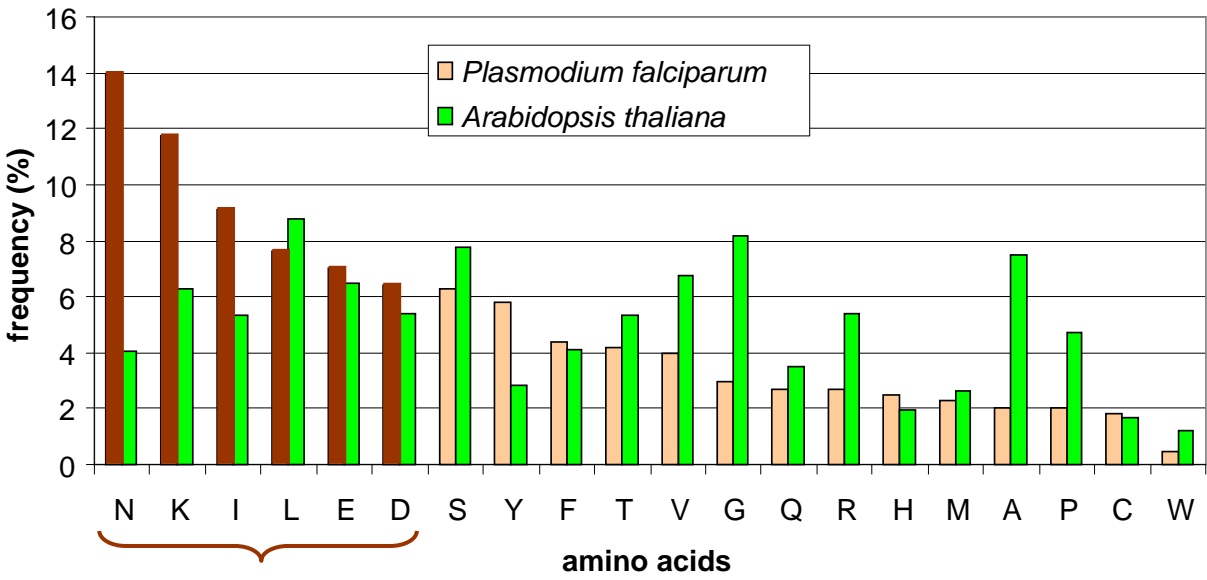
# CURRENT LIMITS

Available molecular data  
PlasmoDB, etc.  
+ comparison with other organisms  
(in particular plants)



**sequence comparisons**  
**analyses of biological processes**

**Blast (+ motifs) searches**



**> 50% protein content**

**~3200 sequences**

# CURRENT LIMITS

Available molecular data  
PlasmoDB, etc.  
+ comparison with other organisms  
(in particular



sequence comparisons

analyses of biological processes

A R N D C

A	5	-3	-2	-3	0
R	-1	6	-4	-1	-2
N	-1	0	4	1	-1
D	-2	-3	0	5	-4
C	1	-3	-2	-4	8

*abidopsis thaliana*

Blast (+ motifs) searches

E	G	H	I	L	K	M	F	P	S	T	W	Y	V
-3	0	-2	-2	-2	-4	-2	-3	-1	0	0	-1	-3	0
-2	-3	0	-3	-5	1	0	-7	-1	-1	0	-3	-5	-3
0	0	0	-3	-2	0	-2	-3	0	0	0	-2	-2	-2
1	-1	0	-3	-5	0	-1	-5	0	0	-1	-6	-5	-3
-5	0	-2	-1	0	-5	-1	-2	-2	0	0	0	0	0
0	-2	0	-5	-1	0	0	-6	0	0	-2	0	0	-3
4	-1	-1	-4	-3	0	-2	-5	0	0	0	0	-4	-2
-4	7	-1	-3	-5	-4	-4	-5	-3	-2	-2	-5	-4	-5
0	-2	7	-4	-2	-3	-2	-1	-1	0	-1	0	1	-2
-2	-4	-3	4	1	-2	0	0	-1	-2	0	0	-2	2
-3	-4	-2	1	4	-3	2	0	-5	-2	-1	0	-1	0
0	-2	0	-3	-2	4	0	-5	0	0	0	-5	-2	-2
-4	-3	0	0	2	-2	6	0	-1	-1	-1	-4	-3	0
-3	-3	-1	0	0	-4	0	6	-3	-3	-4	1	2	0
-1	-5	-1	-7	-3	-1	-4	-1	7	-1	-1	-4	-2	-3
0	0	-1	-3	-5	0	-2	-2	0	4	0	-3	-3	-3

P/c

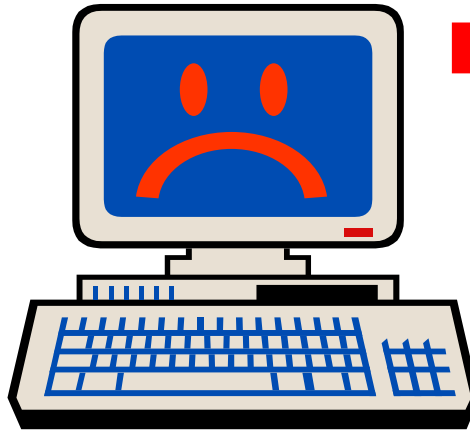
T	0	0	-1	-1	-1	0	0	-1	-1	-1	-1	0	-2	-1	1	5	-4	-5	0	
W	-6	-1	-5	-3	0	-4	-4	-3	-1	-2	0	-3	0	3	-5	-3	-3	11	2	-4
Y	-1	0	-2	-2	-1	-1	-1	-2	1	-2	0	-2	0	3	-2	-1	-1	3	6	-1
V	0	-3	-5	-7	0	-3	-2	-2	-4	2	0	-2	0	0	-2	-3	0	-2	-2	4

~3200 sequences



## CURRENT LIMITS

Available molecular data  
PlasmoDB, etc.  
+ comparison with other organisms  
(in particular plants)



sequence comparisons

analyses of biological processes

Blast (+ motifs) searches

⇒ **Sequence alignment methods ?**

✓ **Substitution matrices ?**

• **Non-symmetric matrices ⇒ in progress**

✓ **Score statistics?**

• **E-value vs Z-score**

⇒ **Z-score**

⇒ **Use at massive scale in progress**

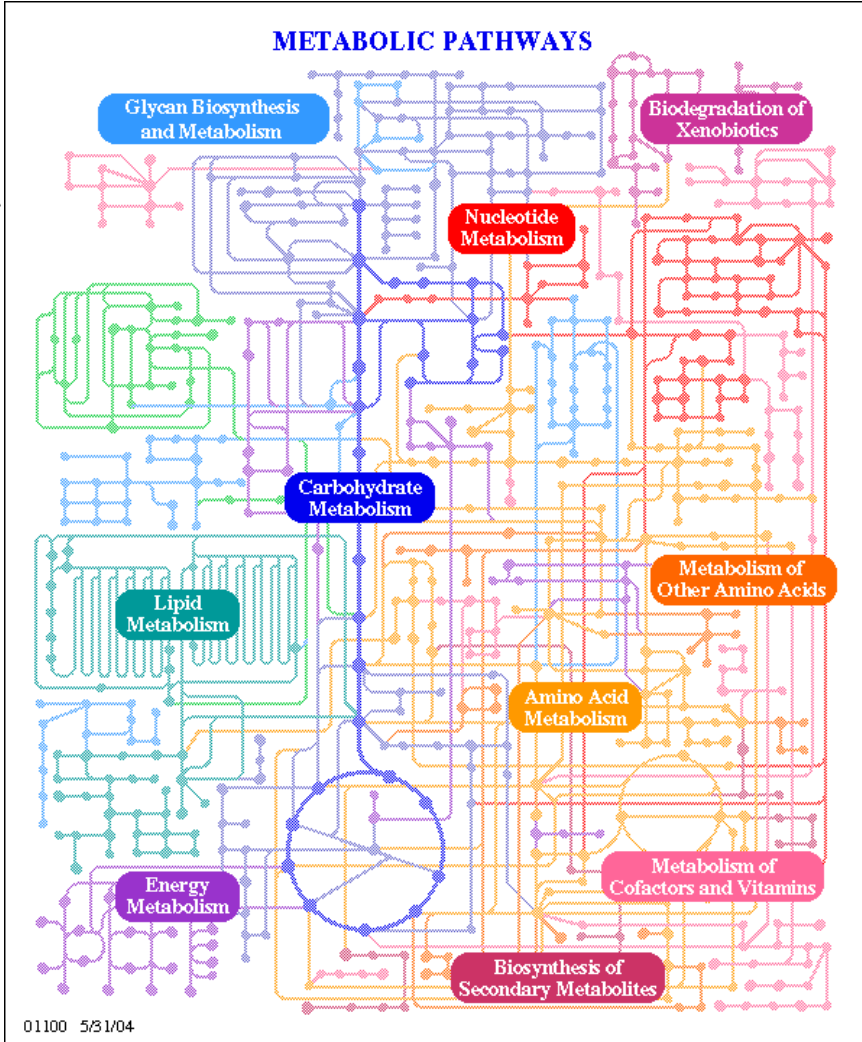
**~3200 sequences**

# CURRENT LIMITS

Available molecular data  
PlasmoDB, etc.  
+ comparison with other organisms  
(in particular plants)



- sequence comparisons
- analyses of biological processes



## FUTURE CHALLENGES

Potential of  
*in silico*  
approaches  
for target  
discovery ?



- ⇒ more integration of genomic/post-genomic molecular and functional data
- ⇒ from user-friendly, but rigid, web portals to flexible & creativity-oriented knowledge accesses (with user-designed workflows, implying a strong interoperability)
- ⇒ improved and new analyzing and mining tools
  
- ⇒ integration of biological and chemical information (chemogenomic knowledge) and models for biological and chemical space intersects
  - what biological space ? what chemical space ?
  - bridging biological targets → ligand
    - bridge #1: recorded effect of drugs/small molecules on known targets
    - bridge # 2: protein structure –small molecule structure (docking)
  - bridging small molecule → biological responses
    - OMIC functional responses
  - implementing new predictions (*i.e.* druggability)

*Comparative chemogenomics* **CEA Grenoble**

**C Botté, O Bastien, N Saïdani,  
D. Grando, A. Zoppé, J Jouhet, H Valadié  
MA. Block, E Maréchal**

*AI for chemogenomics+cheminformatics*

**CEA Grenoble**

**S. Wiczorek, S. Aci, S Roy**

**IMAG Grenoble**

**G. Bisson, M. Gordon**

**Université de Lille**

**D Horvath**

*Functional chemogenomics* **CEA Grenoble**

**L Lafanechère**

*Automated screening* **Cerep, France**

**E Nicolai, F Revah**

*Virtual screening* **IN2P3 Clermont-Ferrand**

**V Kasam, V Breton**

*Diversity oriented chemistry* **CEA Saclay**

**M Deligny, AL Bonneau, R Lasselin,**

**B Rousseau, R Lopez**

**Faculté de Pharmacie Meylan**

**YS Wong**

*Structural biology* **Cermav, Grenoble**

**C Jouanneau, M Audry**

**A Imberty, C Breton**

*Parasitology* **Grenoble**

**C. Mercier, C. Bisanz,**

**MF. Cesbron-Delauw**

**Montpellier**

**JF Dubremetz, H Vial**

**Mexico**

**R Mondragon**

**Seattle**

**J Feagin,**

**Marburg**

**N. Azzouz, R Schwarz,**

*Ontological status of plants*

**Univ. J Fourier Grenoble**

**N Aumonier**

*Bioinformatics + Post genomics analyses*

**LIRMM Montpellier**

**O. Gascuel, L. Bréhélin & team**

**Gene-IT, France**

**K Metayer, JJ Codani**

**Pretoria, South Africa**

**LM Birkholtz, F Joubert, B. Louw**

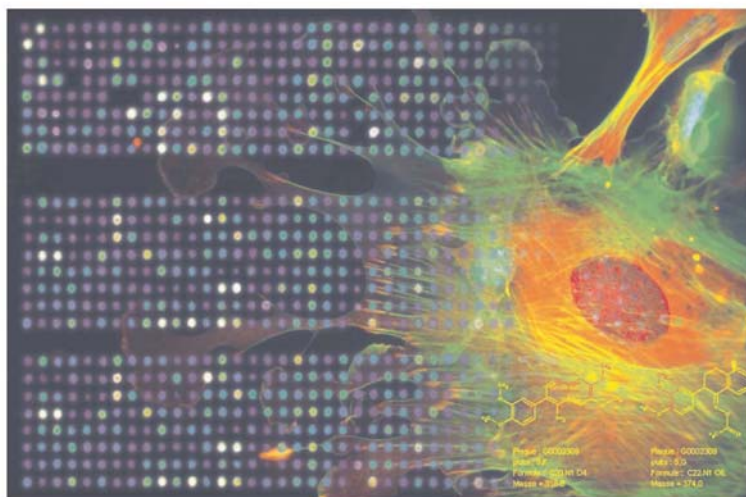
**CEA Cadarache**

**P Ortet**

# CHEMOGÉNOMIQUE

DES PETITES MOLÉCULES  
POUR EXPLORER LE VIVANT

■ sous la direction de  
**Eric MARÉCHAL**  
**Sylvaine ROY**  
**Laurence LAFANECHÈRE**



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(Eds. Maréchal, Roy, Lafanechère)  
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