Rates, dates and traits.

The comparative method in evolutionary genomics

Nicolas Lartillot

June 19, 2014

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Variation of the substitution rate among lineages



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Variation of the substitution rate among lineages



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Variation in GC content



Romiguier et al, 2010, Genome Res 20:1001 🗆 🕨 < 🖻 🕨 < 🗏

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rRNA and proteome composition vs. temperature





Figure: Procaryotic protein content in amino acids IVYWREL correlated to the species optimal growth temperatures (OGT) [Zeldovich et al., 2007].

Zeldovich et al 2007

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Introduction

Ancestral growth temperatures inferred using rRNA



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The problem of phylogenetic inertia



Felsenstein, 1985, Am Nat 125:1

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The comparative method



- Assume traits follow bivariate Brownian motion
- data $X = (x_{ik}), i = 1..N_{taxa}, k = 1, 2$
- parameter Σ (3 independent parameters)
- maximize likelihood $L(\Sigma) = p(X \mid \Sigma)$
- estimate Σ₁₂, test whether Σ₁₂ < 0, etc

Brownian process



$$x_t \sim Normal(x_0, \sigma^2 t)$$

or, for a bivariate or multivariate process:

 $X_t \sim Normal(X_0, t\Sigma)$

The comparative method



Algorithm

- take two sister species i and j, with traits X_i and X_j
- T: time since their last common ancestor
- $\Delta X = X_j X_i \sim Normal(0, 2T\Sigma).$
- define normalized *contrast* $\Delta Y = \Delta X / \sqrt{2T}$: $\Delta Y \sim Normal(0, \Sigma)$.
- do it for all tips and then recursively up to the root

The comparative method



- *P* taxa \rightarrow *P* 1 normalized contrasts ΔY_j , for j = 1..P 1
- normalized contrasts are iid: ΔY_j ~ Normal(0, Σ)
- or equivalently, likelihood reduces to

$$p(X \mid \Sigma) \propto \prod_{j=1}^{P-1} Normal(\Delta Y_j; 0, \Sigma)$$

Independent Contrasts



Whitney and Garland Jr, 2010, PLoS Genetics 6:e1001080

- contrasts are statistically independent
- equivalent to asking whether traits show correlated variations

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Genome size and effective population size revisited



Phylogenetically-corrected regression in Archaea



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Ancestral growth temperatures inferred using rRNA



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Brownian model



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Brownian model



Regressing the trait against the predictor

- trait x (e.g. temperature)
- predictor y (e.g. GC content)
- trait evolution over time $t: \Delta x \sim N(0, \eta^2 t)$
- predictor evolution: $\Delta y = \alpha \Delta x + \epsilon$, with $\epsilon \sim N(0, \kappa^2 t)$

•
$$\Delta x \mid \Delta y, \alpha, \eta, \kappa \sim N(m, \lambda^2)$$

A phylogenetic Kalman filter



Regressing the trait against the predictor

- trait x (e.g. temperature)
- predictor y (e.g. GC content)
- trait evolution along branch *I*: $\Delta x_l = x_l x_n \sim N(0, \eta^2 t_l)$
- predictor evolution: $\Delta y_l = y_l y_n = \alpha \Delta x_l + \epsilon_l$, with $\epsilon_l \sim N(0, \kappa^2 t_l)$
- $\Delta x \mid \Delta y, \alpha, \eta, \kappa \sim N(m, \lambda^2)$

A phylogenetic Kalman filter



Conditional likelihoods

• at node *n*, and for $x_n \in \mathbb{R}$, $L_n(x)$ is Gaussian:

$$L_n(x_n) = K_n e^{-\frac{1}{2\sigma_n^2}(x_n-\mu_n)^2}$$

- calculate K_n , σ_n , μ_n as functions of K_l , K_r , σ_l , σ_r , μ_l , μ_r ...
- backward: likelihood computation; forward: stochastic traceback
- combined with conjugate sampling of covariance matrix
- Lartillot, 2014. Bioinformatics, 30:486-496 (see also Ho and Ané, Syst Biol 2014)

A phylogenetic Kalman filter



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$$L_n(x_n) = \left[\int_{-\infty}^{\infty} p(x_n \to x_l \mid t_l) L_l(x_l) \, dx_l\right] \left[\int_{-\infty}^{\infty} p(x_n \to x_r \mid t_r) L_r(x_r) \, dx_r\right]$$

analytical integrals: all factors are Gaussian, result is Gaussian

archaeal rRNA dataset



Inferred temperature for archaeal ancestor

- 95% credible interval: (91,110) Celsius.
- without molecular information: (60,96) Celsius

The comparative method – Summary



independent contrast traits follow bivariate Brownian motion

- more generally: statistical models of the evolutionary processes
- a large variety of questions: bursts, trends, jumps, correlations.

Variation of the substitution rate among lineages



The relaxed molecular clock



Branch lengths = times x rates

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The Brownian relaxed molecular clock



- substitutions occur at rate r_t
- rt modeled as Brownian motion along branches
- Brownian model induces rate autocorrelation across branches
- joint estimation of rates and times by Bayesian MCMC

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Estimating divergence times: the relaxed clock model



Data and constraints

- multiple alignment D (here, nuclear coding genes in mammals)
- tree topology *T*, and fossil calibrations Φ

Principle of the method

- build a hierarchical model, with a prior on its parameters
- sample from the posterior distribution using MCMC algorithms

Diversification process





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birth-death with subsampling

- speciation rate λ , extinction rate μ , sampling fraction ρ
- t: vector of divergence times
- gives you a probability distribution on times: $p(t \mid \lambda, \mu, \rho)$

Brownian process



gives you a probability distribution on rates: $p(r \mid t, \sigma^2)$

Model of sequence evolution by point substitutions

Substitution rate matrix Q (4 x 4)

$$Q = \begin{pmatrix} A & C & G & T \\ \hline A & - & \frac{\gamma}{2} & \kappa \frac{\gamma}{2} & \frac{1-\gamma}{2} \\ C & \frac{1-\gamma}{2} & - & \frac{\gamma}{2} & \kappa \frac{1-\gamma}{2} \\ G & \kappa \frac{1-\gamma}{2} & \frac{\gamma}{2} & - & \frac{1-\gamma}{2} \\ T & \frac{1-\gamma}{2} & \kappa \frac{\gamma}{2} & \frac{\gamma}{2} & - \end{pmatrix}$$

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κ: transition-transversion ratio

• γ : equilibrium GC (*GC*^{*})

Substitution process (rate matrix Q)



branch length approximated as:

$$I = \bar{r}\Delta t$$
, where $\bar{r} = \frac{r_j + r_{j_{up}}}{2}$

- length *I*: expected number of point substitutions along the branch
- gives you a probability distribution on sequences: p(D | r, t, Q)

Complete model



- diversification process (e.g. birth-death, parameters λ , μ , ρ)
- substitution rate: Brownian log-normal process (variance σ²)
- substitution process (4x4 substitution matrix Q)
- complete model configuration: $\theta = (\lambda, \mu, \rho, \sigma, t, r)$

posterior distribution proportional to joint probability:

 $p(\lambda)p(\mu)p(\rho)p(\sigma^2)$ $p(t \mid \lambda, \mu, \rho)$ $p(r \mid t, \sigma^2)$ $p(D \mid r, t, Q)$

Bayesian inference and Monte Carlo sampling



Metropolis Hastings on rates

$$\alpha = \frac{p(D \mid r', t, Q) p(r' \mid t, \sigma^2)}{p(D \mid r, t, Q) p(r \mid t, \sigma^2)}$$

 $\alpha > 1$: accept move

 α < 1: accept with prob. α

Bayesian inference and Monte Carlo sampling



Metropolis Hastings on divergence times

$$\alpha = \frac{p(D \mid r, t', Q) p(r \mid t', \sigma^2) p(t' \mid \lambda, \mu, \rho)}{p(D \mid r, t, Q) p(r \mid t, \sigma^2) p(t \mid \lambda, \mu, \rho)} \quad \begin{array}{l} \alpha > 1: \text{ accept move} \\ \alpha < 1: \text{ accept with prob. } \alpha \end{array}$$

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Bayesian inference and Monte Carlo sampling



Metropolis Hastings on σ^2 , λ, μ, ρ

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Posterior mean times and rates



Correlating rates and traits



- sequential method: error propagation problems
- circularity in the way phylogenetic inertia is dealt with
- suggests a more direct integrative approach

Coupling trait evolution and substitution process



Lartillot and Poujol, 2011, Mol Biol Evol, 28:729

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Hierarchial Bayesian model (parameter estimation by MCMC)

- diversification process t (birth-death, parameters λ, μ, ρ)
- Brownian multivariate process X (covariance matrix Σ)
- time-dependent codon model Q

Coupling substitution process with life-history evolution



(Lartillot and Poujol, 2011, Molecular Biology and Evolution)

posterior proportional to joint probability:

 $p(\lambda, \mu, \rho) p(t \mid \lambda, \mu, \rho) p(\Sigma) p(X \mid t, \Sigma) p(D \mid X, t)$

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Generalization

time-dependent substitution parameters

- rate of synonymous substitution (r)
- non-synonymous / synonymous ratio (ω)
- equilibrium GC composition (γ)

time-dependent quantitative traits

- sexual maturity (proxy of generation time)
- adult body mass
- maximum recorded lifespan (proxy of longevity)
- metabolic rate
- genome size
- karyotypic number (number of chromosomes 2n)

Joint inference of rates, dates and traits



Alignment of 13 genes 4800 coding positions



red: positive correlation blue: negative correlation

Lartillot and Delsuc, 2012, Evolution 66:1773

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Equilibrium GC (GC^*) in nuclear genomes





Lartillot, 2013, Molecular Biology and Evolution, 30:356

- negative correlation between GC* and body size
- positive correlation between GC* and number of chromosomes

Biased conversion during meiosis



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Biased conversion during meiosis



Biased gene conversion (BGC) during meiosis



GC overtransmission

- meiotic distorsion bias $b \iff$ like positive selection for GC
- *b* proportional to local recombination rate $(b = b_0 r)$

Duret and Galtier 2009, Annu Rev Genomics Hum Genet 2009, 10:285

Biased gene conversion explains variation in GC*



de Villena and Sapienza, 2001, Mamm Genome 12:318

Positive correlation GC* / chromosome number

- ~ 1 recombination event per chromosome arm per meiosis
- more fragmented karyotype = smaller chromosomes
 - = higher recombination rate = stronger gene conversion

Biased gene conversion explains variation in GC*



red: positive blue: negative light shade: not significant

Negative correlation GC* / body mass

- larger animals = smaller population = less efficient selection
- also less efficient BGC (lower GC*)

Romiguier and Galtier 2010, Genome Res 20:1001

GC-biased gene conversion interferes with selection



ADCYAP1 gene, Ratnakumar et al, 2010, Phil Trans R. Soc. B 365:2571

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Fixation probability in the presence of BGC GC overtransmission



Relative fixation probability: 2Nep

mutation from AT to GC

 $2N_ep=\frac{B}{1-e^{-B}}>1$

mutation from GC to AT

$$2N_ep=rac{-B}{1-e^B}<$$

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 N_e : effective population size $B = 4N_eb$: scaled conversion coefficient

A mechanistic phylogenetic covariance model

substitution rate = mutation rate x fixation probability

$$\begin{pmatrix} - & \mu_{AC} & \mu_{AG} & \mu_{AT} \\ \mu_{CA} & - & \mu_{CG} & \mu_{CT} \\ \mu_{GA} & \mu_{GC} & - & \mu_{GT} \\ \mu_{TA} & \mu_{TC} & \mu_{TG} & - \end{pmatrix} + B \implies \begin{pmatrix} - & \mu_{AC} \frac{B}{1-e^{-B}} & \mu_{AG} \frac{B}{1-e^{-B}} & \mu_{AT} \\ \mu_{CA} \frac{-B}{1-e^{B}} & - & \mu_{CG} & \mu_{CT} \frac{-B}{1-e^{B}} \\ \mu_{GA} \frac{-B}{1-e^{B}} & \mu_{GC} & - & \mu_{GT} \frac{-B}{1-e^{B}} \\ \mu_{TA} & \mu_{TC} \frac{B}{1-e^{-B}} & \mu_{TG} \frac{B}{1-e^{-B}} & - \end{pmatrix}$$

 $B = 4N_e b$: scaled conversion coefficient



Lartillot, 2013, Molecular Biology and Evolution, in press

Overall modeling strategy

- only 4-fold degenerate third codon positions
- modeling joint variation in *B*, body mass (*M*) and karyotype (2*n*)
- modeling among-gene variation (recombination seascapes)

Data

Exon-rich dataset

- 180 exons from Orthomam, with at least 30 taxa
- 1000 exons (30 jacknife replicates of 100 exons)

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- only 4-fold degenerate positions
- analysis replicated using non-CpG positions

Taxon-rich dataset

• 17 single-exon genes 73 mammals

Reconstructed history of $B = 4N_eb$



Lartillot, 2013, Molecular Biology and Evolution, 30:489

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Phylogenetic history of population-genetic regimes



B = 0.1 2.8 5.5

large mammals, small N_e : drift dominates (B < 1)

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Phylogenetic history of population-genetic regimes



B = 0.1 2.8 5.5

small mammals, large N_e : BGC dominates (B > 1)

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Lartillot, 2013, Molecular Biology and Evolution, in press

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Allometry and covariance

- (In B, In M, In n) follow a trivariate Brownian motion
- ${\it B} \sim {\it M}^{\gamma} \, {\it n}^{lpha}$ for some coefficients of allometry γ and lpha

Estimated allometric scaling of $B = 4N_eb$

$B \sim M^{\gamma} n^{\alpha}$

M: body mass (prediction: $\gamma < 0$)

n: number of chromosomes (prediction: $\alpha = 1 > 0$)

$$\begin{array}{ccc} \gamma & \alpha \\ \\ & & \\$$

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Reconstructed history of $B = 4N_eb$



Divergence times and body mass evolution



last common ancestor: between 150 g and 3.5 kg

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Reconstructing past population-genetic regimes

- mutation rate per generation u (substitution rate)
- effective population size N_e (dN/dS, GC)
- scaled conversion coefficient $B = 4N_e b$ (GC)
- evolutionary dynamics of recombination landscapes (GC)
- useful for understanding mechanisms of genome evolution

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Molecular dating and diversification studies

Diversification studies: current approach

• likelihood (time-calibrated tree T, diversification parameters θ):

$$L(\theta) = \rho(T \mid \theta)$$

Diversification studies: integrative approach

• use diversification model as your prior on divergence times:

 $p(D \mid r, T) p(r \mid T) p(T \mid \theta) p(\theta)$

- compare models based on their marginal likelihoods
- avoids circularity and overconfidence
- dating and diversification: two sides of a same coin

Integrative models for macroevolutionary studies

Toward a unified probabilistic framework for

- reconstructing divergence times
- fitting / testing diversification models
- fitting / testing models of trait evolution
- understanding driving forces of molecular evolution
- correlating diversification / traits / substitution patterns
- see also total evidence dating (Ronquist et al, Syst Biol 61:973)

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- Nicole Uwimana, Benoit Nabholz
- Benjamin Horvilleur (Brownian paths)
- many others...

Software availability (coevol)

www.phylobayes.org